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<p>(54) Title: PEPTIDES AND ANTI-SENSE PEPTIDES WITH BROAD NEOPLASTIC SPECIFICITY (57) Abstract An analogue peptide comprises the variable regions of the light or heavy chains of an antibody of a first species selectively binding to a tumor antigen, such as a carcinoma antigen, has 1 to 46 amino acids of the framework regions per chain substituted with amino acids such as those present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 variable region CDRs per chain and optionally flanking regions thereof of 1 to 10 or more amino acids, alone or with an N-terminal fragment of 1 to 10 or more amino acids, combinations or mixtures thereof. The polypeptide may also comprise an effector agent and/or be glycosylated, and is presented as a composition with a carrier. The analogue peptides are used in diagnostic kits for neoplasms such as carcinomas and methods for in vivo imaging and treating a primary or metastasized neoplasm such as a carcinoma, and in vitro diagnosing a neoplasm, ex vivo purging neoplastic cells from a biological fluid. RNAs and DNAs encode the analogue peptide, and a hybrid vector carrying the nucleotides and transfected cells express the peptides and a method produces the analogue peptide. An anti-idiotypic polypeptide comprises polyclonal antibodies raised against an antineoplastic antibody or the analogue peptide of this invention, monoclonal antibodies thereof, Fab, Fab', (Fab')₂, CDR, variable region, or analogues or fragments thereof, combinations thereof with an oligopeptide comprising a TRP trimer, tandem repeats thereof, or combination or mixtures thereof. An anti-idiotypic hybrid polypeptide with an effector agent and the anti-idiotypic polypeptide, an anti-neoplastic vaccine, an anti-neoplastic vaccination kit, a method of vaccinating against neoplasms such as carcinomas and a method of lowering the serum concentration of a circulating antibody or polypeptide are provided.</p>		

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PEPTIDES AND ANTI-SENSE PEPTIDES WITH BROAD NEOPLASTIC SPECIFICITY

BACKGROUND OF THE INVENTION

Field of the invention

This invention relates to the in vitro and in vivo diagnosis, immunization, and therapy of neoplastic tumors, particularly carcinomas, by means of specifically targeted analogue peptides comprising amino acid sequences encompassing the complementarity determining regions (CDRs) of a different species, and analogues of the variable (F_v) region of anti-carcinoma antibodies, among others. The carcinoma specific peptides for use in one species, e.g., humans, are provided as a single amino acid chain having the specificity of F_v regions obtained in different species, e.g., murine antibody F_v regions of the light or heavy chains, or as paired chains. These peptides are provided either by themselves or bound to other molecules such as synthetic polymers or oligopeptides resulting in sequences of mixed species, and more particularly analogues of human/non-human chimeric antibodies or other polymeric constructs. The analogue peptides comprise sequences derived from the variable regions of heterologous antibodies specific for, e.g., human carcinoma antigens that elicit a lesser immunological response in humans than the whole heterologous antibodies. The anti-idiotypic polypeptides and their analogues are suitable for immunizing humans or other animals against carcinoma. Polynucleotide segments encoding the analogue peptide and anti-idiotypic polypeptides, and hybrid vectors and transfected host cells carrying the segments are useful for preparing the peptides disclosed herein.

Description of the Background

Carcinomas result from the carcinogenic transformation of cells of different epithelia. Two of the most damaging characteristics of carcinomas are their uncontrolled growth and their ability to create metastases in distant sites of the host, particularly a human host. It is usually these distant metastases that cause serious consequences to the host, since frequently the primary carcinoma may be, in most cases, removed by surgery. The treatment of metastatic carcinomas, that are seldom removable, depends on irradiation therapy and systemic therapies of different natures. The systemic therapies currently include, but not fully comprise, chemotherapy, radiation, hormone therapy, different immunity-boosting medicines and procedures, hyperthermia and systemic monoclonal antibody treatment. The latter can be labeled with radioactive elements, immunotoxins and chemotherapeutic drugs.

Radioactively labeled monoclonal antibodies were initially used with success in lymphomas and leukemia, and recently in some carcinomas. The concept underlying the use of labeled antibodies is that the labeled antibody will specifically seek and bind to the carcinoma and, the radioactive element, through

its decay, will irradiate the tumor in situ. Since radioactive rays travel some distance in tumors it is not necessary that every carcinoma cell bind the labeled antibody. The specificity of the monoclonal antibodies will permit a selective treatment of the tumor while avoiding the irradiation of innocent by-stander normal tissues, that could be dose limiting. Chemotherapy produces serious toxic effects on normal tissues, making the chemotherapy of carcinomas less than desirable, and the use of radiolabeled monoclonal antibodies a valid alternative.

Non-human antibodies raised against human epitopes have been used for the diagnosis and therapy of carcinomas as is known in the art. Also known are the methods for preparing both polyclonal and monoclonal antibodies. Examples of the latter are BrE-2, BrE-3 and KC-4 (e.g., US patent Nos. 5,077,220; 5,075,219 and 4,708,930..

The KC-4 murine monoclonal antibody is specific to a unique antigenic determinant, the "antigen", and selectively binds strongly to neoplastic carcinoma cells and not to normal human tissue (U.S. Patent No. 4,708,930 to Coulter). The antigen appears in two forms in carcinoma cells, only the smaller of these forms being expressed in the cell membrane. The larger form appears only in the cytoplasm and has an approximate 490 Kdalton molecular weight (range of 480,000-510,000). The second form occurs at a higher density of expression, is found both in the cytoplasm and the membrane of carcinoma cells and has an approximate 438 Kdalton molecular weight (range of 390,000-450,000) as determined by gel electrophoresis with marker proteins of known molecular weights. Labeled KC-4 was applied to the diagnosis and medical treatment of various carcinomas, particularly adenocarcinoma and squamous cell carcinoma regardless of the human organ site of origin.

The BrE-3 antibody (Peterson et al., Hybridoma 9:221 (1990); US patent No. 5,075,219) was shown to bind to the tandem repeat of the polypeptide core of human breast epithelial mucin. When the mucin is deglycosylated, the presence of more tandem repeat epitopes is exposed and the binding of the antibody increases. Thus, antibodies such as BrE-3 bind preferentially to neoplastic carcinoma tumors because these express an unglycosylated form of the breast epithelial mucin that is not expressed in normal epithelial tissue. This preferential binding combined with an observed low concentration of epitope for these antibodies in the circulation of carcinoma patients, such as breast cancer patients, makes antibodies having specificity for a mucin epitope highly effective for carcinoma radioimmunotherapy. A ^{90}Y -BrE-3 radioimmunoconjugate proved highly effective against human breast carcinomas transplanted into nude mice. Human clinical studies showed the ^{90}Y -BrE-3 radioimmunoconjugate to

- considerably reduce the size of breast tumor metastases without any immediate toxic side effects. Moreover, an ^{111}In -BrE-3 radioimmunoconjugate was successfully used for imaging 15 breast cancer patients, providing excellent tumor targeting in 13 out of 15 of the patients. Out of all the breast tumor metastases occurring in another study, 86% were detected by ^{111}In -BrE-3. Unfortunately, 2 to 3 weeks after treatment, the patients developed a strong human anti-murine antibody (HAMA) response that prevented further administration of the radioimmunoconjugate. The HAMA response, which is observed for numerous murine monoclonal antibodies, precludes any long-term administration of murine antibodies to human patients. Similarly, other heterologous antibodies, when administered to humans, elicited similar antibody responses. The anti-heterologous human response is, thus, a substantial limiting factor hindering the successful use of heterologous monoclonal antibodies as therapeutic agents, which could, otherwise, specifically annihilate breast carcinomas, causing little or no damage to normal tissue and having no other toxic effects.

- Chimeric antibodies are direct fusions between variable domains of one species and constant domains of another. Murine/human chimeric antibodies prepared from other types of B cells binding to other types of antigenic determinants have been shown to be less immunogenic in humans than whole murine antibodies. These proved to be less immunogenic but still in some cases an immune response is mounted to the rodent variable region framework region (FR). A further reduction of the "foreign" nature of the chimeric antibodies was achieved by grafting only the CDRs from a rodent monoclonal into a human supporting framework prior to its subsequent fusion with an appropriate constant domain, (European Patent Application, Publication No. 239,400 to Winter; Riechmann, et al., *Nature* 332:323-327 (1988)). However, the procedures employed to accomplish CDR-grafting often result in imperfectly "humanized" antibodies. That is to say, the resultant antibody loses avidity (usually 2-3 fold at best).

The ligand binding characteristics of an antibody combining site are determined primarily by the structure and relative disposition of the CDRs, although some neighboring residues also have been found to be involved in antigen binding (Davies, et al., *Ann. Rev. Biochem.* 59:439-473 (1990)).

- The technologies of molecular biology have further expanded the utility of many antibodies by allowing for the creation of class switched molecules whose functionality has been improved by the acquisition or loss of complement fixation. The size of the bioactive molecule may also be reduced so as to increase the tissue target availability of the antibody by either changing the class

from an IgM to an IgG, or by removing most of the heavy and light chain constant regions to form an F₁ antibody. Common to all of these potentially therapeutic forms of antibody are the required complementary determining regions (CDRs), which guide the molecule to its ligand, and the framework residues (FRs) which support the CDRs and dictate their disposition relative to one another. The crystallographic analysis of numerous antibody structures revealed that the antigen combining site is composed almost entirely of the CDR residues arranged in a limited number of loop motifs. The necessity of the CDRs to form these structures, combined with the appreciated hypervariability of their primary sequence, leads to a great diversity in the antigen combining site, but one which has a finite number of possibilities. Thus, its hypermutability and the limited primary sequence repertoire for each CDR would suggest that the CDRs derived for a given antigen from one species of animal would be the same derived from another species. Hence, they should be poorly immunogenic, if at all, when presented to a recipient organism.

Accordingly, there is still need for a product of high affinity and/or specificity for carcinoma antigens suitable for the detection and therapy of carcinomas which elicits a lesser antibody response than whole non-human antibodies or chimeric antibodies containing, for instance the entire non-human variable region.

SUMMARY OF THE INVENTION

This invention relates to an analogue peptide and its glycosylated derivative which specifically and selectively bind to the human mammary fat globule (HMFG) antigen and to an antigen found on the surface or in the cytoplasm of tumor cells such as carcinoma cells or that is released by the cells. the analogue peptide consisting essentially of at least one variable region of the light or heavy chains of an antibody of a first species having affinity and specificity for an antigen found on the surface or the cytoplasm of a carcinoma cell or released by the cell, wherein preferably about 1 to at least 46 amino acids in the FR are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain or 1 to 3 CDRs plus flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids, combinations thereof or combinations thereof with other variable regions or analogues thereof, wherein each analogue peptide is operatively linked to at least one other peptide or analogue thereof, or mixtures thereof.

Also provided herein are a fusion protein and a hybrid polymer comprising the analogue of the invention, the corresponding DNAs encoding them, hybrid vector thereof, transfected host thereof, and RNA.

- 5 This invention also encompasses a method of producing an analogue peptide or hybrid analogue peptide by recombinant technology, in vitro methods of diagnosing and immunohistochemistry of tissue slices, an ex vivo method of purging neoplastic cells, and in vivo methods for imaging and therapy of neoplasias such as carcinomas.

- Also disclosed herein are anti-idiotypic polypeptides comprising polyclonal
10 antibodies raised against the analogue peptide of this invention, analogues thereof, monoclonal antibodies thereof, fragments thereof selected from the group consisting of Fab, Fab', (Fab')₂, CDRs, variable regions and analogues thereof as described above, combinations thereof operatively linked to one another, an anti-carcinoma vaccine, a vaccination kit, a method of vaccinating
15 against neoplasias including carcinomas, and a method of lowering the serum concentration of circulating anti-tumor antibody with the anti-idiotypic polypeptide of this invention.

Other objects, advantages and features of the present invention will become apparent to those skilled in the art from the following discussion.

20

DESCRIPTION OF THE PREFERRED EMBODIMENTS

- This invention arose from a desire by the inventors to improve on antibody technology suitable for use in diagnostic, prognostic, vaccine and therapeutic applications in the field on neoplasmas and cancer. The present invention will be described for application to humans. However, it is also
25 suitable for use in other species. The monoclonal antibodies obtained up to the present time have been prepared by fusing immortalized cell lines with B-cells of non-human origin such as murine, rat, rabbit, goat, and the like. Many of these hybridomas can produce large quantities of monoclonal antibodies that have desirable binding properties such as high affinity and/or specificity and selectivity
30 for carcinoma antigens of a species, e.g., for human carcinoma antigens. However, in general, antibodies from other species, e.g., non-human antibodies, may only be administered once to a subject of a predetermined species, e.g., humans, due to the detrimental effects they produce. This is true for most heterologous antibodies being administered to mammalian animals. For example,
35 the repeated administration of murine antibodies to a human subject elicits a strong human anti-murine antibody (HAMA) response, which precludes their further utilization as therapeutic agents in humans. These non-human antibodies initiate an immediate adverse reaction in many human patients and are, thus,

rendered ineffective for further administration as therapeutic agents. In some cases, non-human-human chimeric antibodies and non-human CDR "grafted"-human antibodies may have lower affinity and/or specificity for their antigens than the corresponding non-human antibody. On the other hand, human monoclonal hybridoma cell lines have not been very stable and have, therefore, not been suitable for the large scale, repeated production of monoclonal antibodies.

The present invention is applicable to the manufacture of analogue peptides having a high number of amino acids in the respective positions found in the endogenous antibodies, and where at least the CDRs are of another xenogeneic species.

The present inventors, thus, have undertaken the preparation of antitumor non-human CDRs and non-human variable regions of antibodies, having affinity and specificity for an antigen found on the surface or the cytoplasm of a human neoplastic cell such as a carcinoma cell or that is released by the cell, wherein about 1 to 46 amino acids in the framework region (FR) are substituted per chain with amino acids present in equivalent positions in other human antibodies, or fragments thereof comprising 1 to 3 CDRs per chain or 1 to 3 CDRs per chain plus flanking regions thereof, each of about 1 to 10 or more amino acids, alone or plus an N-terminal fragment of about 1 to 10 or more amino acids, to lower or even circumvent the anti-xenogeneic response. To preserve substantial binding specificity in the molecules intended for use in humans, the present invention utilizes CDRs and/or analogues of varying lengths of the variable regions of light and/or heavy chains of antibodies of other species such as murine, rat, rabbit, goat, equine, primate, bovine, and guinea pig antibodies, among others. Heterologous human CDRs and/or analogues of their variable regions may be utilized are intended for use in other species.

The present inventors have found, surprisingly, that these analogue antibody fragments substantially preserve the binding, specificity and selectively characteristics of the whole non-human antibody while eliciting a lesser detrimental immunological response. However, the simple preservation of the binding region of an antibody does not by itself ensure that the binding characteristics of the antibody will be maintained. Antibodies are glycopolypeptides that are folded into specific conformations. When the glycoside portion of the molecule or portions of the amino acid sequence are perturbed or excised, the folding pattern of the molecule may be perturbed. Thus, any deletion or modification of the sequence of an antibody must be made taking into consideration that its folding-dependent properties may be diminished or even obliterated if the folding is substantially affected, even though the amino

acid sequences involved in the binding of the antigen are preserved.

The present inventors selected the following strategy for the preparation and manufacture of the analogue and hybrid peptides of this invention. The cDNAs that encode the variable chains of an antibody may be obtained by
5 isolation of mRNA from a hybridoma cell and reverse transcription of the mRNA, amplification of the cDNA by polymerase chain reaction (PCR) and insertion of the DNA into a vector for optional sequencing, and for restriction enzyme cutting. The cDNAs encoding the CDRs or variable chain (F_v) region fragments of the light (V_L) and heavy (V_H) chains of an antibody having affinity and
10 specificity for a carcinoma cell antigen may be reverse transcribed from isolated mRNA. The cDNAs encoding the CDRs may then be ligated to other segments encoding neighboring sequences, and the variable region cDNAs may then be modified with predesigned primers used to PCR amplify them or synthesized de novo, cloned into a vector optionally carrying DNA sequences encoding, e.g.,
15 a constant region(s), optionally sequenced, and then transfected into a host cell for expression of the analogue gene product. The binding specificity characteristics of the analogue peptides may then be determined and compared to those of the whole antibodies.

X-ray crystallographic studies demonstrate that the framework
20 structures of the F_v of different antibodies assume a canonical structure regardless of the species of origin, amino acid sequence, or ligand specificity. This is generally taken as evidence that the ligand-binding characteristics of an antibody combining site are determined primarily by the structure and relative disposition of the CDRs, although some neighboring framework residues may
25 also be involved in antigen-binding. Thus, if the fine specificity of an antibody is to be preserved, its CDR structures, and probably some of the neighboring residues, their interaction with each other and with the rest of the variable domains, must also be maintained. These crystallographic studies point to the possible need for retaining most, if not all, of the many interior and inter-domain
30 contact residues since the structural effects of replacing only a few of them cannot be predicted.

While at first the necessity of keeping these amino acids might seem to defeat the goal of decreasing immunogenicity by "humanization", the actual
35 number of amino acids that must be retained is usually small because of the striking similarity between human and murine variable regions. Moreover, many, if not most, of the retained amino acids possess side chains that are not exposed on the surface of the molecule and, therefore, may not contribute to its antigenicity. Clearly, it is most of the exposed amino acids that are good candidates for substitution since it is these amino acids that are exposed to the

immunological environment of a mammal and may form epitopes of increased immunogenicity.

The challenge in humanizing the variable regions of a non-human antibody, e.g., a murine antibody, thus begins with the identification of the "important" xenogeneic amino acids. "Important" amino acids are those, for example, that are involved in antigen binding, contact the CDRs and the opposite chains, and have buried side-chains. Ideally, these residues might be identified from a well characterized three-dimensional structure. However, when direct structural data are not available, it is, fortunately, still possible to predict the location of these important amino acids by analyzing other related antibody structures, especially those whose variable light and heavy regions belong to the same class. The classes of variable regions can be determined from their amino acid sequence.

A method by which these important amino acids are identified has been described for the case of the amino acids with buried side chains by Padlan, E.A. (Padlan, E.A., "A Possible Procedure for Reducing the Immunogenicity of Antibody Variable Domains While Preserving Their Ligand-Binding Properties", *Molecular Immunology*, 28:489-494 (1991)). In the present case, various antibody variable region structures were compared using a computer program that determines the solvent accessibility of the framework residues as well as their contacts with the opposite domain as described by Padlan, E.A. (1991), supra. Surprisingly, a close examination of the fractional solvent accessibility reveals a very close similarity in the exposure patterns of the V_H and the V_L domains. Put in simple terms, regardless of the particular antibody in question, and of its amino acid sequence, the buried residues occupy similar relative positions in most antibodies.

A similar analysis can be done by computer modeling, to determine which amino acids contact the CDRs and which contact the opposite domain. At this point, the Fab structures that are currently in the Protein Data bank (Bernstein, F.C., et al., *J. Mol. Biol.* 112:535-542 (1977)) may be examined to determine which FRs are probably important in maintaining the structure of the combining site. Thus, after a close inspection of many high resolution three-dimensional structures of variable regions, the positions of all important framework amino acids, that is, those that contact the CDRs, the opposite domain, and those whose side chains are inwardly pointed, may be tabulated. Keeping these amino acids, as well as those from the CDRs, and finally those FR amino acids that may be involved in ligand binding, should insure to a great extent the preservation of affinity. The precise identification of FR amino acids that are involved in ligand-binding cannot be generalized since it varies for

differe... antibodies. Nevertheless, conservative decisions can be made to preserve the amino acids located in FR that have a high probability of contacting the antigen. These regions are located immediately adjacent to the CDRs and at the N-terminus of both chains, because the surfaces of these regions are contiguous with the CDR surfaces.

Surprisingly, it is possible to keep all of these important amino acids in a heterologous humanized antibody and still increase dramatically the similarity with a human consensus sequence. That is, the final number of amino acids with murine identities differing from human identities that are kept is typically small.

This is usually possible because human frameworks that are similar to the murine frameworks, especially at the positions of the important amino acids, can be found. This is because many of the important amino acids have the same identities in both murine and human antibodies.

All the amino acids that are determined to be not important by the method described above may be completely replaced by their corresponding human counterparts. The surface of the finally humanized antibody should look very much like that of a human antibody except for the antigen binding surfaces. The original shape of those binding surfaces, however, is maintained by leaving the internal composition of the antibody intact, preserving inter-domain contacts and by keeping very few key amino acids that contact the CDRs.

a) Choosing the Best Human Framework to Use in the "Humanization" of an Antibody When Its Structure Is Known

At the present time, there are 11 Fab structures for which the atomic coordinates are known and have been placed in the Protein Data Bank as shown in Table 1 below, 2 from human and 9 from murine antibodies.

Table 1: Fab Structures for Which Coordinates are in the Protein Data Bank

	ANTIBODY	RESOLUTION (Å)	R-VALUE	PDB CODE
HUMAN:	NEWK	2.0	0.46	3FAB
	KOL	1.9	0.189	2FB4
MURINE:	McPC603	2.7	0.225	1MCP
	J539	1.95	0.194	2FBJ
	HyHEL-5	2.54	0.245	2HFL
	HyHEL-10	3.0	0.24	3HEM
	R19.9	2.8	0.30	1F19
	4-4-20	2.7	0.215	4FAB
	36-71	1.85	0.248	6FAB
	B1312	2.8	0.197	1IGF
	D1.3	2.5	0.184	1FOL

Figure 1. The effect of the number of trials on the mean accuracy of the responses ($n = 10$) as a function of the number of items presented at once. Error bars represent standard error.

5

POSITION	ANTIBIOT										KOL
	J535	MCPC603	MYHEL-10	MYHEL-5	R19.9	4-4-20	36-71	B1312	01.3	NEWN	
1	GLU (2)	ASP (5)	ASP (10)	ASP (3)	ASP (8)	ASP (4)	ASP (11)	ASP (13)	SER (1)		
2	ILE (11)	ILE (15)	ILE (17)	ILE (13)	ILE (5)	ILE (20)	VAL (9)	ILE (10)	VAL (2)		
3		VAL (3)	VAL (2)	VAL (3)	GLN (2)	VAL (2)	GLN (2)	LEU (5)	VAL (2)		
4	LEU (7)	MET (6)	LEU (6)	LEU (10)	MET (9)	MET (13)	MET (7)	MET (6)	LEU (4)	LEU (6)	
5		THR (1)			THR (1)	THR (2)			THR (1)		
6								THR (4)			
7								SER (5)			
22										CYS (1)	
23	CYS (1)	CYS (1)	CYS (2)	CYS (2)	CYS (1)	CYS (1)	CYS (1)	TRP (4)	TAP (4)	TRP (1)	
25	TRP (3)	TRP (2)	TRP (4)		TRP (2)	TRP (6)	TRP (1)	TRP (4)	TAP (4)	TRP (1)	
36	TYR (12)	TYR (16)	TYR (8)	TYR (10)	TYR (23)	TYR (13)	TYR (15)	TYR (8)	TYR (14)	TYR (13)	
37					LYS (12)	LYS (5)					
45	PRO (3)	LEU (6)	LEU (4)	ARG (15)	LEU (5)	VAL (14)	LEU (5)	LEU (10)	LEU (6)	LEU (2)	
46	ILE (1)	ILE (1)	ILE (1)		ILE (1)	ILE (2)	ILE (1)	VAL (1)	VAL (1)	ILE (1)	
48	TYR (20)	TYR (29)	LYS (13)	TYR (12)	TYR (40)	TYR (22)	TYR (22)	TYR (16)	TYR (25)	TYR (25)	
49	VAL (3)	VAL (3)	ILE (1)	VAL (6)	VAL (6)	VAL (5)	VAL (4)	VAL (5)	VAL (1)	VAL (6)	
50		ASP (1)				ASP (2)	ASP (4)			ASP (2)	
60											
62				PHE (1)		PHE (1)				LYS (13)	
64									LYS (2)		
67		SER (3)				THR (1)	THR (4)	THR (1)	SER (1)		
69		THR (3)	THR (3)			ASP (6)			SER (2)		
70		ASP (2)							ALA (2)	ALA (4)	
71	TYR (14)	PHE (23)	PHE (17)	TYR (17)	TYR (24)	PHE (17)	TYR (17)	PHE (19)	TYR (16)	CYS (1)	
84	CYS (1)		CYS (2)		CYS (1)	CYS (1)	CYS (1)	CYS (1)	CYS (2)	PHE (7)	
94	PHE (8)	PHE (8)	PHE (10)	PHE (5)	PHE (8)	PHE (14)	PHE (8)	PHE (14)	PHE (14)		

10 below.

Table 3:

[illegible]

the FR amino acids that contact the opposite domain and which presumably are the ones mainly responsible for the quaternary structure of the F_v domains are listed in Table 4 below.

5 **Table 4:** Framework Residues That Contact Framework Residues in the Opposite Domain in Fabs of Known Three-Dimensional Structure

IN V_L	POSITION	J558	MPCC603	HYHEL-10	HYHEL-5	RL1.3	4-4-20	36-71	81322	01.3	MEMM	EDL
	36	TYR(2)	TYR(4)	TYR(3)	TYR(5)	GLN(5)	TYR(13)	TYR(7)	TYR(11)	TYR(7)	GLN(7)	TYR(13)
	38	GLN(10)	GLN(11)	GLN(9)	GLN(5)	GLN(5)	GLN(3)	GLN(8)	GLN(13)	GLN(8)	ALA(5)	GLN(8)
	43	SEA(7)	PRO(11)	SEA(8)	SEA(5)	TYR(3)			SEA(3)	SEA(3)	PRO(7)	PRO(13)
	44	PRO(10)	PRO(11)	PRO(8)	PRO(11)		PRO(7)	ILE(20)	PRO(16)	PRO(16)	PRO(7)	PRO(13)
	46											
	48			MET(2)		TYR(3)			VAL(1)		ASP(12)	
	49	TYR(4)	TYR(4)	PHE(4)	TYR(2)		PHE(3)	TYR(10)	TYR(8)	TYR(8)	TYR(8)	TYR(14)
	50	PHE(11)	PHE(8)	PHE(7)	PHE(12)	PHE(12)	PHE(12)	PHE(8)	PHE(13)	PHE(12)	PHE(10)	PHE(13)
	100		ALA(2)									

IN V_H	POSITION	J558	MPCC603	HYHEL-10	HYHEL-5	RL1.3	4-4-20	36-71	81322	01.3	MEMM	EDL
	37	VAL(4)		ILE(2)	VAL(1)	VAL(4)	VAL(1)	VAL(2)	VAL(4)	VAL(4)	VAL(1)	VAL(4)
	39	GLN(10)	GLN(4)	LYS(8)	GLN(5)	GLN(5)	GLN(3)	GLN(8)	GLN(10)	GLN(8)	GLN(4)	GLN(7)
	43			ASN(4)		GLN(7)			LYS(6)		ASN(19)	
	44											
	45	LEU(13)	LEU(12)	LEU(8)	LEU(14)		LEU(8)	LEU(13)	LEU(13)	LEU(14)	LEU(11)	LEU(14)
	47	TRP(11)		TYR(2)		TRP(2)			TRP(2)		TRP(2)	
	51	TYR(8)	TYR(4)	TYR(3)	TYR(8)	PHE(2)	TYR(2)	PHE(4)	TYR(3)	TYR(5)	TYR(3)	
	103	TRP(13)	TRP(13)	TRP(14)	TRP(11)	TRP(4)	TRP(18)	TRP(24)	TRP(12)	TRP(19)	TRP(8)	TRP(19)
	105	GLN(3)										

The buried, inward-pointing FR amino acids in the V_L domains, i.e., those which are located in the domain interior, are listed in Table 5 below.

10 **Table 5:** Inward-Pointing, Buried Framework Residues in the V_L of Fabs of Known Three-Dimensional Structure

POSITION	J558	MPCC603	HYHEL-10	HYHEL-5	RL1.3	4-4-20	36-71	81322	01.3	MEMM	EDL
3	ILE	ILE	ILE	ILE	ILE	VAL	ILE	VAL	ILE	LEU	LEU
4	LEU	MET	LEU	LEU	MET	GLN	GLN	GLN	MET	GLN	GLN
6	GLN	GLN	GLN	GLN	GLN	LEU	LEU	LEU	LEU	VAL	ALA
11	THR	LEU	LEU	MET	LEU	VAL	ALA	VAL	ALA	VAL	VAL
13	ALA	VAL	VAL	ALA	VAL	ALA	VAL	ALA	VAL	VAL	VAL
15	VAL	VAL	VAL	VAL	VAL	ILE	ILE	ILE	ILE	ILE	ILE
21	ILE	MET	LEU	MET	ILE	CTG	CTG	CTG	CTG	CTG	CTG
23	CTG	CTG	CTG	CTG	TRP	TRP	TRP	TRP	TRP	TRP	TRP
25	TRP	TRP	TRP	TRP	TRP	LEU	LEU	LEU	LEU	LEU	LEU
27	GLN	GLN	GLN	GLN	GLN	LEU	LEU	LEU	LEU	LEU	LEU
41	TRP	LEU	LEU	TRP	LEU	ILE	ILE	ILE	VAL	PHE	ILE
43	ILE	ILE	ILE	ILE	VAL	ILE	ILE	ILE	VAL	PHE	ILE
45											
49	VAL	VAL	ILE	VAL	VAL	VAL	ARG	ARG	ARG	ARG	ARG
51	ARG	ARG	ARG	ARG	PHE	PHE	PHE	PHE	PHE	PHE	PHE
63	PHE	PHE	PHE	PHE	TYR	PHE	PHE	PHE	TYR	ALA	ALA
71	TYR	PHE	PHE	TYR	LEU	LEU	LEU	LEU	LEU	LEU	LEU
73	LEU	LEU	LEU	LEU	ILE	ILE	ILE	ILE	ILE	ILE	ILE
75	ILE	ILE	ILE	ILE	ILE	VAL	LEU	VAL	LEU	LEU	LEU
78	MET	VAL	VAL	MET	LEU	ASP	ASP	ASP	ASP	ASP	ASP
83	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	PHE	ASP	ASP
85											
87	ALA	ALA	ALA	ALA	ALA	TYR	TYR	TYR	TYR	TYR	TYR
89	CTG	CTG	CTG	CTG	CTG	CTG	CTG	CTG	CTG	CTG	CTG
91	CTG	CTG	CTG	CTG	THR	THR	THR	THR	THR	THR	THR
101	THR	THR	THR	THR	LEU	LEU	LEU	LEU	LEU	VAL	VAL
103	LEU	LEU	LEU	LEU	ILE	ILE	ILE	ILE	ILE	ILE	ILE
105	LEU	ILE	ILE	ILE							

Those in the V_H domain are listed in Table 6 below.

Table 1
Inward-Pointing, Buried Framework Residues in
the V_H of Fabs of Known Three-Dimensional Structure

POSITION	J123	McPC603	HyHEL-10	HyHEL-5	RL3.9	4-4-20	36-71	SI212	01.3	McW	ROL
2	VAL	VAL	VAL	LEU	LEU	LEU	VAL	VAL	VAL	LEU	VAL
4	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU	LEU
6	GLU	GLU	GLU	GLN	GLU	GLU	GLN	GLU	GLU	GLN	GLN
9				PRO			VAL	VAL	VAL	VAL	VAL
12	VAL	VAL	VAL	MET	VAL	MET	VAL	LEU	LEU	LEU	LEU
18	LEU	LEU	LEU	ILE	MET	LEU	MET	LEU	ILE	LEU	LEU
20	LEU	LEU	LEU	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS
22	CYS	CYS	CYS	ALA	ALA	ALA	ALA	ALA	VAL	VAL	SER
24	ALA	THR	VAL	ALA	ALA	ALA	ALA	ALA	THR	THR	THR
27	PHE	PHE	ASP	TYR	TYR	PHE	TYR	PHE	LEU	PHE	PHE
33	PHE	PHE	ILE	PHE	PHE	PHE	PHE	PHE	THR	THR	THR
36	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP	TRP
38	AAC	AAC	AAC	LYS	LYS	AAC	LYS	AAC	AAC	AAC	AAC
40						SER					
46		GLU	GLU	GLU			VAL	ILE	VAL	ILE	VAL
48	ILE	ILE	MET	ILE	ILE	ALA	ALA	AAC	AAC	AAC	ALA
49		ALA									
66	LYS	AAC	AAC	LYS	THR	PHE	THR	PHE	LEU	VAL	PHE
67	PHE	PHE	ILE	PHE	LEU	ILE	LEU	ILE	ILE	MET	ILE
69	ILE	VAL	ILE	PHE	VAL	VAL	VAL	AAC	LYS	VAL	AAC
71	AAC	AAC	AAC	ALA	VAL						
76						SER					
78	LEU	LEU	TYR	ALA	ALA	VAL	ALA	LEU	VAL	PHE	LEU
80	LEU	LEU	LEU	MET	MET	LEU	MET	LEU	LEU	LEU	MET
82	MET	MET	LEU	LEU	LEU	MET	LEU	MET	MET	LEU	MET
83C	VAL	LEU	VAL	LEU	LEU	LEU	LEU	LEU	VAL	VAL	LEU
84	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP	ASP
88	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA	ALA
90	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR	TYR
92	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS	CYS
94	AAC	AAC	ASP	HIS	AAC	THR	THR	THR	THR	SER	THR
107	THR	THR	VAL	LEU	VAL	THR	THR	THR	THR	VAL	VAL
109	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL
111	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL	VAL

5 From the above, it may be seen that

(1) There are many FR amino acids that either contact the CDRs or the opposite domain, or are found in the domain interior.

(2) These FR amino acids, which could influence the structure of the combining site, and thus the antigen-binding characteristics of an antibody, are
10 different from antibody to antibody.

It is obvious from these results that no one structure can serve as the perfect and sole basis of all "animalization", or in the present example "humanization", protocols. In fact, to "humanize" the 9 murine antibodies shown in Table 1 above by CDR-grafting with a view to preserving their ligand-
15 binding properties, the FR amino acids listed in Table 2 to 6 above would have to be retained.

A search through the tables of immunoglobulin sequences (Kabat et al., "Sequences of Proteins of Immunological Interest", 5th Ed. US Dept. of Health and Human Service, NIH Publication No.91-3242 (1991)), shows that human
20 variable domain sequences are known that already have most of the FR amino acids that need to be preserved as shown in Table 7 below.

Table 7:

Human Antibodies that are Most Similar in
Sequence to Murine Antibodies of Known
Three-Dimensional Structure

ANTIBODY DOMAIN		MOST SIMILAR HUMAN SEQUENCE
HYHEL-10	VH	58P2'CL (77/112)
	VH FRAMEWORK	15P1'CL, H11'CL (62/87)
	VH IMPT	58P2'CL, AD26'CL, C692'CL (28/38)
	VL	IARC/BL41'CL (73/107)
HYHEL-5	VH	743c'CL, X17115'CL (63/87)
	VH FRAMEWORK	21/28'CL, 51P1'CL, 783c'CL, 8E10'CL, AMD, KAS, NEI'CL,
	VH IMPT	X17115'CL (25/37)
	VL	HF2-1/17'CL, KAS (65/105)
	VL FRAMEWORK	HF2-1/17'CL (57/80)
R19.9	VH	21/28'CL (73/119)
	VH FRAMEWORK	21/28'CL, 51P1'CL, AMD, L52'CL, NEI'CL (60/87)
	VH IMPT	21/28'CL, 8E10'CL, L52'CL (28/38)
	VL	WALKER'CL (78/107)
4-4-20	VH	30P1'CL (77/116)
	VH FRAMEWORK	2P1'CL, 306'CL (65/87)
	VH IMPT	4B4'CL, H26'CL (36/41)
	VL	RPM1-6410'CL (91/112)
J539	VH	30P1'CL, Vh38Cl.10'CL (81/116)
	VH FRAMEWORK	18/2'CL, 30P1'CL, H43 (71/87)
	VH IMPT	38P1'CL, 56P1'CL, H72, H74 (36/40)
	VL	PA (62/105)
HEPC603	VH	LEN, MEA (53/80)
	VH FRAMEWORK	BI, DEN, KUE, REI, WALKER'CL, WIL(-) (26/35)
	VH IMPT	
	VL	H72 (81/120)
36-71	VH	4G12'CL, AD18'CL, H72 (70/87)
	VH FRAMEWORK	56P1'CL, H72, H74, RF-5J2'CL (36/42)
	VH IMPT	FK-001'CL, LEN (91/113)
	VL	LEN (70/80)
J539	VH	LEN (36/42)
	VH FRAMEWORK	21/28'CL (74/119)
	VH IMPT	21/28'CL, 51P1'CL, 783c'CL, AMD'CL, NEI'CL, X17115'CL,
	VL	(61/87)
J539	VH	21/28'CL, 8E10'CL (28/38)
	VH FRAMEWORK	AG (76/105)
	VH IMPT	REI (63/80)
	VL	REI, RZ, WALKER'CL (34/37)

Table

Human Antibodies that are Most Similar
Sequence to Murine Antibodies of Known
Three-Dimensional Structure (Cont'd)

B1312	VH	56P1*CL (83/119)
	VH FRAMEWORK	4B4*CL, 4G12*CL, H26*CL, H72, RF-SJ2*CL, Vh38C1.10*CL (68/87)
	VH IMPT	56P1*CL, H72, H74, RF-SJ2*CL (37/39)
	VL	RPH1-6410*CL (86/112)
D1.3	VL FRAMEWORK	GH-607*CL (69/80)
	VL IMPT	CUM, NIM (36/39)
	VH	C6B2*CL (72/116)
	VH FRAMEWORK	C6B2*CL (62/87)
	VH IMPT	H60*CL (32/37)
	VL	BR (75/107)
	VL FRAMEWORK	KF2-1/17*CL (64/80)
	VL IMPT	3D6*CL, BI, DEN, EU, KUE, PA, REI, WALKER*CL, WELI*

- 5 These human sequences are not necessarily those which are most similar to the murine antibodies, overall or in the framework regions only, but rather, those that possess the largest number of important amino acids in common, the latter sequences being included in Table 7 above.

- 10 The number of murine amino acids that still need to be retained in order to have all the important FR amino acids in the "humanized" or analogue versions of the murine antibodies, as shown in Table 7 above, ranges from 21 (for HyHEL-5:12 in V_H and 9 in V_L) to 5 (for B1312:2 in V_H and 3 in V_L). These are not very many amino acids, considering that the resulting "humanized" or analogue molecules will probably retain most or all their ligand-binding characteristics. It is possible that there exist other human sequences that are even more similar to these murine domains that are not included in the compilation of Kabat, et al. (1991), supra. When more sequences become available these may also be incorporated to improve the pool of basic data available.

- 10

10

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Table 9: Framework Residues in V_H That Probably Need to Be Preserved in Order to Reproduce the Ligand Properties of the Original Antibody

[illegible]

15

FR amino acids that are involved in the contact with the opposite domain as shown in Table 4 above, and many of those which are in contact with the CDRs as shown in Tables 2 and 3 above. Moreover, almost all of the FR amino acids that have been observed to participate in the binding to antigen (Amit, A.G., et al., Science 233:747-753 (1986); Sheriff, et al., P.N.A.S. (USA) 82:1104-1107 (1987); Padlan, E.A., et al., P.N.A.S. (USA) 86:5938-5942 (1989); Tulip, et al., Cold Spring Harbor Symp. Quant. Biol. 54:257-263 (1989); Bentley, et al., Nature (London) 348: 254-257 (1990)), are in these flanking regions. Thus, during "animalization" or "humanization" or formation of the analogue peptides, not just the CDRs are retained, but also some of the residues immediately adjacent to the CDRs. This provides a better chance of retaining more of the ligand-binding properties of the original antibody. The likelihood of retaining the antigen binding properties of the original antibody is even greater if the first few amino acids in the NH₂-termini of both chains are also retained, since some of them are found to be in contact with CDRs as shown in Tables 2 and 3 above. Further, Tables 8 and 9 above also show many other framework positions that are deemed structurally important in all the cases examined here. The xenogeneic residues at those positions should probably be retained as well.

Alternatively, it may possible to reduce immunogenicity, while preserving antigen-binding properties, by simply replacing those exposed residues in the framework regions which differ from those usually found in human antibodies (Padlan, E.A. (1991), supra). This would "humanize" the surface of the xenogeneic antibody while retaining the interior and contacting residues which influence its antigen-binding characteristics. The judicious replacement of exterior residues should have little, or no, effect on the interior of the domains, or on the interdomain contacts. For example, the solvent accessibility patterns of the F₂s of J539, a murine IgA (κ) and of KOL, a human IgG1 (λ) have been found to be very similar (Padlan, E.A. (1991), supra).

At present, more than 35 different Fab structures have been elucidated by X-ray diffraction analysis, although atomic coordinates for only 11 are currently in the Protein Data Bank as shown in Table 1 above. Most of the available structures have been analyzed to only medium resolution, some having been refined to only a limited extent. Eventually, atomic coordinates for more and better-refined structures will become available, so that the "important" FRs will be more easily assessed. This will improve the theoretical predictive record of the present method for determining the best mode for the analogue peptides.

As already indicated above, the specificity of an antibody depends on the CDR structures and sometimes, on some neighboring residues as well. These structures, in turn, depend on contacts with framework amino acids and on the

interaction of the V_L and V_H domains. Thus, to ensure the retention of binding affinity, not only the CDR residues must be preserved, but also those FRs that contact either the CDR's or the opposite domain, as well as all buried residues, which give shape to the variable domains.

5 This design of the humanized versions of murine antibodies is reached in stages as follows.

- 1- Choice of a xenogeneic model of known structure.
- 2- Choice of the target species FR.
- 3- Identification of xenogeneic/target species differences.
- 10 4- Identification of important xenogeneic amino acids.

(1) Choice of a xenogeneic model of known structure

The V_H and V_L domains of an antibody of desired specificity are classified according to Kabat et al.(1991), supra. Then, an antibody of the same species may be chosen, whose structure has been determined, and whose
15 variable regions belong to the same classes and subclasses. Modeling the xenogeneic antibody in question to such structure ensures maximal chance for success. This, however, is not absolutely necessary since the relative positions of the important amino acids do not vary considerably even in variable regions of different classes. Thus, with less than a perfect match this method may still
20 be applied to design the analogues of this invention. Once the xenogeneic model is chosen, it may be applied to identify the locations of important residues in the xenogeneic antibody to be animalized (humanized). Tables 2, 3, 4, 5, 6, 8 and 9 indicate the positions of the important amino acids in several antibodies whose structures have been determined to a high resolution level.

25 (2) Choice of the target species FR

The target species framework should, ideally, be a consensus framework. That is, one that has a maximum number of amino acids in common with all human frameworks of the same class. This is important, because, the goal of humanization is to avoid an immunological response against the engineered
30 analogue peptide.

The target species framework that is chosen is that which shares the greatest number of important amino acids with the original xenogeneic antibody. Thus, in choosing the target species (human) FR, the similarity between the important amino acids is more important than the overall similarity.

35 In practice, the sequences of the xenogeneic variable chains are aligned with the consensus sequences from all variable region classes of the target species and the number of differences in the amino acids that must be retained from the xenogeneic species are scored. The human consensus sequence(s) that

score(s) the lowest number of differences is (are) then chosen. These are the best analogue peptide candidates. Others with low numbers that are higher than the above may also be suitable, and are placed in a reserve pool, and so forth.

- If there are too many differences in the chosen framework (e.g., more than 16), then the same alignment procedure using all tabulated human sequences may be repeated in order to find a specific human framework whose similarity with the xenogeneic sequence is maximized at the positions of the important amino acids. Thus, most preferably, the target species FR should be a consensus sequence. Next preferable would be a framework of a common target species (human) antibody, and finally, the framework of any target species (human) antibody.

(3) Identification of xenogeneic target species differences

- The xenogeneic sequences are then aligned with the target species sequences and the positions of all amino acids that differ in the murine and in the human frameworks are tabulated. Such a table contains the maximum number of amino acids that can be changed toward the full "animalization" ("humanization") of the xenogeneic antibody (see, Tables 31 and 32 below). If all those changes were to be made, a so-called CDR-grafted antibody would be obtained. That is, only the original CDRs would be retained from the murine antibody. In some cases, possibly, such CDR-grafted antibody may maintain the original binding affinity. In most instances, however, the affinity of a CDR-grafted antibody would be considerably less than that of the original xenogeneic antibody. In order to maximize the chances for conserving the original affinity, the identities of all important amino acids must be preserved.

(4) Identification of important xenogeneic amino acids

- If the outlined approach to animalization (humanizing) an antibody is followed strictly, the amino acids that are correspondingly important in the model xenogeneic antibody chosen in step 1 are retained. In a more preferred approach, however, the amino acids that have been shown to occupy important positions in other antibodies of the same species or of the target species may also be retained and are therefore taken out from the group of candidates to be mutated. This preferred approach may be particularly appropriate when there is a chance that the amino acids in question could make contacts with the CDRs or with the opposite chains. Once the important xenogeneic amino acids are identified, the DNA sequence may be mutagenized to change all other amino acids, which for the most part occupy exposed positions.

The present method is exemplified for a murine antibody humanized with the intent of diminishing or avoiding a HAMA response upon its administration

to humans. Murine and human antibodies, whose three-dimensional structures have been deduced to a high degree of resolution, were utilized as guidance in the choice of the amino acids to be substituted in order to humanize the particular murine antibody utilized. The method, however, may be applied more generally to transform antibodies from one species into a less immunogenic form to be administered to a second species, provided that adequate three-dimensional models are available for antibodies from those species. Information on other murine antibodies from a Data Bank was used in the exemplary disclosure provided below to modify the BrE-3 and anti-KC-4 murine-human chimeric antibodies with human amino acids. Similarly, antibodies of other species besides murine may also be utilized, their CDRs and other amino acids preserved and those amino acids not considered "important" replaced with human amino acids. Similarly, the above approach may be applied to the preparation of "animalized" antibodies for any animal species. This may be attained by substituting amino acids of the antibody target species into an antibody of another species in accordance with this invention.

Various peptide structures, such as CDRs, and analogue antibodies, Fab, Fab', (Fab')₂, and variable fragments having a desired specificity, may be constructed and optionally bridged via a linker. In addition, one or more of the peptides may be attached to one or more effector agent(s) or bridged via a linker. Multiple antibody, variable regions, Fab, Fab', (Fab')₂, CDRs and the like, and combinations thereof, may also be constructed and bridged via linkers or attached to one or more effector agents such as are described below.

The cDNAs encoding the analogue variable regions of an antibody of a desired specificity may be cloned into a vector, optionally containing sequences encoding constant regions or fragments thereof, enzymes, neuropeptides, other peptide transmitters, toxins, hormones, operative conjugation regions, cytokines, lymphokines and the like, optionally under the same promoter. Although this is the cloning strategy utilized in the exemplary disclosure of this invention, other methods known in the art may also be utilized such as co-expression and the like. In the exemplary disclosure provided herein, the anti-BrE-3 and anti-KC-4 murine-human chimeric antibodies specifically binding to human mammary mucin and carcinoma cells was constructed by joining the DNAs of the anti-BrE-3 or anti-KC-4 murine variable domain to a human constant domain (an effector agent) cloned into a hybrid vector, and the product expressed by transfecting the vector into myeloma cells. The variable regions of the chimeric antibody were modified at the DNA level to obtain an analogue or "humanized" chimeric polypeptide. The modifications to the variable regions of the peptides may either be conducted by PCR amplification with primers that are custom tailored to

produce the desired mutations, or by gene synthesis.

The analogue "humanized" peptides prepared and exemplified below comprise the "humanized" variable regions of the anti-BrE-3 or anti-KC-4 murine antibodies (U.S. Patent Nos. 5,075,219 and 4,708,930) and the kappa and gamma 1 constant region of a human antibody. These humanized antibodies were characterized by their molecular weights and binding specificities, and shown to compete well with or better than the respective parent murine and chimeric antibodies for the antigen. The analogue "humanized" peptides were shown to bind weakly to normal breast, lung, colon and endometrium, and strongly to carcinoma tissue sections by the ABC immunoperoxidase method. The portions of the CDR and FR regions of the non-modified peptides (murine F₁ regions) and effector agents (human F₁ regions) were shown in both cases to be substantially identical to those of the non-human and human antibodies from which they were obtained. The analogue peptides and hybrid derivatives of this invention lacking any non-human constant region sequences possess less foreign antigenic epitopes than the whole xenogeneic or chimeric antibodies from which they are derived. Accordingly, they are expected to elicit a less complex immunogenic response in animals such as humans than the corresponding non-human whole antibodies or even than the chimeric antibodies. However, to what extent a portion of the non-human FR amino acids may be replaced without altering the binding characteristics of the CDRs could not have been predicted prior to this invention because of the substantial conformational alterations in the interior regions that affect the binding of the CDRs to the antigen that may occur upon modification of amino acid sequences.

Thus, the substantially pure, isolated analogue peptide of the invention specifically and selectively binds to an antigen present on the surface or in the cytoplasm of a carcinoma cell or that is released by the cell. The polypeptide consists essentially of at least one CDR or variable region of the light or heavy chains of an antibody of a first species having affinity and specificity for an antigen found on the surface or the cytoplasm of a carcinoma cell of another species or that is released by the cell, wherein when the framework regions (FRs) are present, about 1 to at least 46 amino acids in the FRs are substituted per chain with amino acids, e.g., present in equivalent positions in antibodies of other species or similar amino acids, or fragments thereof comprising 1 to 3 CDRs per chain, or 1 to 3 CDRs per chain plus flanking regions thereof, each of about 1 to 10 or more amino acids, alone or plus an N-terminal fragment of about 1 to 10 or more amino acids, combinations thereof wherein each analogue peptide is operatively linked to at least one other analogue peptide, combinations thereof and mixtures thereof.

A single unit of the analogue peptide of the invention may be as short as the shortest CDR and as long as the longest combination of variable regions, antibodies, and the like, including non-peptide polymers of up to about 10^6 molecular weight, and in some instances even larger. When several units are linked or other combinations provided, the size of the analogue peptide increases accordingly. The smaller molecular weight analogue peptides are particularly suited for greater penetration of cells, the brain-blood barrier, and tumors, among others, and have a shorter half life, whereas the higher molecular weight polypeptides are suited for either in vitro or in vivo applications such as therapy, imaging and diagnosis. The latter are generally cleared from the body over a longer period of time.

The analogue peptide of the invention may contain amino acid sequences derived from light and/or heavy chains of antibodies of a first or xenogeneic species raised against a variety of antigens and/or epitopes. For example, the murine antibodies disclosed in the examples were raised against human mammary fat globule mucin (BrE-3) and the "KC-4" antigen in human carcinoma cells (KC-4). Other antigens comprising a variety of epitopes may also be utilized to generate the xenogeneic antibodies as long as the antibody contributing the variable region displays low affinity and specificity for normal cells and higher affinity and specificity for human carcinomas that will permit their specific binding to carcinoma cells, preferably, in a variety of tissues. Similarly, the antibodies may be raised in animals of different xenogeneic species. The antibodies from which the polypeptide of the invention is derived may be a murine, rat, goat, birds including poultry, rabbit, guinea pig, equine, bovine, and primate including human and simian, antibodies, among others. The preparation of the antibody and fragments thereof encompassed by the invention is similar, whether the origin of the antibody is human or non-human. The original variable region mRNA may be obtained from cells of any desired xenogeneic species and the remainder of the work-up is similar, utilizing a model antibody of the same xenogeneic species and substituting amino acids from the target species.

The humanization procedure described here is designed to minimize potential losses in antigen binding affinity that may result from the introduced amino acids. In the case of the BrE-3 antibody exemplified below, eight amino acid changes were made in the variable region of the light chain and in the variable region of the heavy chain. In the case of the anti-KC-4 antibody exemplified below, seven amino acid changes were introduced in the variable region of the light chain and twelve amino acid changes were made in the variable region of the heavy chain. Furthermore, to minimize the immunological response to the humanized antibody, target human amino acid sequences were

used that comprise the consensus sequences of all appropriate human variable regions. Nevertheless, neither the exemplified amino acid changes nor the exemplified human target sequences are the only choices encompassed by this invention. Many other individual amino acid changes and permutations thereof
 5 may be made without the expectation of significantly affecting either the affinity of the resulting antibody or its human immunogenicity, as taught herein.

The following Tables 10 and 11 indicate other possible amino acid changes for the variable regions of the BrE-3 and KC-4 sequences of the invention. The amino acid positions (or numbers) are as conventionally accepted
 10 (Kabat et al., 1991, supra). The most preferred changes are indicated under the heading "Most Preferred Analogue". For antibodies other than the BrE-3 and anti-KC-4 antibodies, the amino acids shown in the BrE-3 and KC-4 columns become also part of the group of most preferred choices. Amino acid changes that are not part of the most preferred group but that are still part of this
 15 invention are indicated in the next column under the heading "Preferred Analogue". In some instances, the "Preferred Analogue" choices become too numerous and the least acceptable choices are provided instead for that position. Clearly, all amino acids other than those listed under "Not Preferred Analogue" may be substituted at that position.

Table 10: Alternative Amino Acids for V_L Chain

V _L	BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR1					
1	D	D			WIPLMCRT
2	V	V	I	WKCRH	YC
3	V	L	L(V)		
4	M	M	LVI	PTQ	
5	T	T			
6	Q	Q			
7	T	T	IADS		WYMCGRH
8	P	P	AE		WYFKCN
9	L	L	FP		WYMH
10	S	S	T		HDQ
11	L	L	NV		WYCRGH
12	P	P	S		WIKH
13	V	V			YKCRHN
14	S	T	T(S)		WMCE
15	L	P	PFIL		WYCGHDNEQ
16	G	G			
17	D	E	TEQ(D)		WYFPMCR
18	Q	P	PS(Q)		WYFCAHDN
19	A	A	V		WPKMCRTHNQ
20	S	S			WPLCHD
21	I	I			WPKCGDEQ
22	S	S			WVMCHE
23	C	C			

Table 10: Alternative Amino Acids for V_L Chain (Cont.)

BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR2				
35	W	W		
36	F	F	YL(F)	IVHN
37	L	L	Q	WLVKRTHDE
38	Q	Q		IFMCATSDN
39	K	K	R	WIPMCA
40	S	P	P(S)	FLKARTGO
41G				IYFPMCT
42	Q	Q		WYVCD
43	S	S	P	WYFKMHDNEQ
44	P	P		WYKCRGHQDQ
45	K	Q	EQR(K)	YPCGHD
46	L	L	RV	KCD
47	L	L	V	WILMTSN
48	I	I		FPLVMTS
49	Y	Y	S	PLVMA
FR3				
57	G	G		WVTSGDNEQ
58	V	V		IYFLVMATQ
59	P	P	S	
60	D	D	N	WFMCR
61	R	R	T	
62	F	F		
63	S	S	T	IYPLKARSG
64	G	G	D	
65	S	S		
66	G	G		YCHDQ
67	S	S	A	VKMCRHN
68	E	G	GD(E)	VMCARSQ
69	T	T		WPMCE
70	D	D		WIFPMCR
71	F	F		WKMTEQ
72	T	T		WLMCGHNEQ
73	L	L		
74	K	K	NLRE	WLMCHD
75	I	I	L	
76	S	S	IT	LVMC
77	R	R	S	WYFLKCHQ
78	V	V	ALI	WYFKCRHNE
79	E	E	KGQ	WVKM
80	A	A	P	WLKM
81	E	E		ILVKMAGDNE
82	D	D		
83	L	V	MV(L)	WCRH
84	G	G		LVARTSG
85	V	I	IM(V)	WFKCQ
86	Y	Y		
87	F	Y	YL(F)	IMSHE
88	C	C		

Table 10: Alternative Amino Acids for V_L Chain (Cont.)

BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR4				
98	F	F		
99	G	G		
100	G	G	ASQ	IPVKRTG
101	G	G		
102	T	T		
103	K	K	NR	IYMATGHDEQ
104	L	L	V	LG
105	E	E		ILVTSGHNEQ
106	I	I		YLVKMRTD
106	a	a		PLVTI
107	K	K	R	ILVMATSGNE

Table 11: Alternative Amino Acids for V_H Chain

VL	BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR1					
1	E	E	Q	PVLKARGHDEQ	WYPKCRHV
2	V	V	M		
3	K	Q	QR(K)	L	WYKCRSN
4	L	M			WYCG
5	E	V	VD(E)		WLYFMSH
6	E	E	QD		IMCRHD
7	S	S	T		
8	G	G	E		WIYKCRNQ
9	G	G			WIYFMCH
10	G	G	DA		WKCGHN
11	L	L	VF		WYPTHD
12	V	V	I		WYFCD
13	Q	Q	KE		WIYMC RDQ
14	P	P			IYCHN
15	G	G			WIYFMC
16	G	G	RSE		WIYCHDNE
17	S	S	PA		WYCD
18	M				WIYPCH
19	K	R	R(K)		WYFPKDNO
20	L	L	V		WMH
21	S	S			
22	C	C			WFLMCH
23	A	A	TSE		WKMC RDHNEQ
24	A	A	V		WIVMRHDEQ
25	S	S			
26	G	G			WPMCR
27	F	F			CHQ
28	T	A	AINS(T)		WYKCRHDNEQ
29	F	F			WLMCDQ
30	S	S			

Table 11: Alternative Amino Acids for V_H Chain (Cont'd.)

VL	BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
FR2					
36	W	W		WIFLVMATGQ	
37	V	V			YFS
38	R	R			FPVMCA
39	Q	Q			WY
40	S	A	VA(S)		
41	P	P	TS	PLVARTSHNEQ	
42	E	G	G(E)		YPLKMCTHN
43	K	K			WIYLVCS
44	G	G	SR		WIFMCH
45	L	L			
46	E	E	Q		FPMCRTD
47	W	W			PMARNQ
48	V	V	STG		YPKCARTHNO
49	A	A			WFPKCRHNO
FR3					
66	R	R			WYLVMCATSDE
67	F	F			WYPKMHNEQ
68	T	T	IS		WYPGE
69	I	I			WYKCHNQ
70	S	S	L		WVMCHDEQ
71	R	R			WYFCHDQ
72	D	D	N	VKRTSGHDE	
73	D	N	N(D)		WYFLC
74	S	S	A	FPLVTGDN	
75	K	K	EN		NPCGD
76	S	N	NTRK(S)		WFPLMCE
77	R	T	TNIVSM(R)		WFC
78	V	L	LA(V)		WKCNE
79	Y	Y	FH		WPG
80	L	L			
WYLYKARGHEQ					
81	Q	Q	E		WYFPC
82	M	M			YPCTGHDEQ
82a	I	N	SDN(II)		WMO
82b	S	S	IR		WYFPLMHQ
82c	L	L		PLVMAGE	
83	R	R	EKT		WILCH
84	A	A	SPVTI		WKCDEQ
85	E	E	D	WIYFPLMCRG	
86	D	D			
87	T	T	M		WLVMCNE
88	G	A	A(G)		
89	L	V	ITVM(L)		WYPKGEO
90	Y	Y	HF		
91	Y	Y			WIPLVKMAGQ
92	C	C			
93	T	A	ASHAV(T)		WIFR
94	G	R	R(G)	TSDQLAW	
FR4					
103	W	W	A		
104	G	G	YH		
105	Q	Q	THR		IYFLVMCDQ
106	G	G			
107	T	T	AQ		
108	L	L	STGM		WYCHDE

Table 11: Alternative Amino Acids for V_H Chain (Cont'd.)

VL	BrE3	KC4	Most Preferred Analogue	Preferred Analogue	Not Preferred Analogue
109	V	V	LT	IK	
110	T	T	SL		
111	V	V			
112	S	S	T		
113	A	S	S(A)	PLVATG	

Similar tables may be constructed for any and all amino acid sequences for other xenogeneic antibodies as taught herein.

In one particularly preferred embodiment of the invention, the humanized anti-carcinoma analogue peptide comprises the amino acid sequence ID No. 67 to 73 of Table 47, the sequence ID No. 75 to 81 of Table 48, the sequence ID No. 95 to 102 of Table 55 and/or the sequence ID No. 103 to 108 of Table 56, and the sequences wherein about 1 to 46 or more amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in human antibodies, or fragments thereof comprising 1 to 3 CDRs per chain or 1 to 3 CDRs per chain plus flanking regions thereof, each of about 1 to 10 or more amino acids, alone or plus an N-terminal fragment of about 1 to 10 or more amino acids, or combinations thereof wherein each analogue peptide is operatively linked to at least one other analogue peptide, and mixtures thereof.

The present analogue peptide is provided either as a naked peptide or in glycosylated form. When provided in glycosylated form, the analogue peptide may be operatively linked to a glycosyl residue(s) provided by the eukaryotic cell where it is expressed, or it may be cloned and expressed in a prokaryotic cell as the naked polypeptide and the glycosyl residue(s) added thereafter, for example by means of glycosyl transferases as is known in the art. Examples of glycosyl residue(s) that may be added to the analogue peptide of the invention are N-glycosylated and O-glycosylated residues, among others. The glycosyl residues added to the naked analogue peptide may have a molecular weight of about 20 to 50,000 daltons, and more preferably about 100 to 20,000 daltons or greater, depending on the size and molecular weight of the peptide to which they are attached. However, other types of polysaccharides and molecular weights may also be present. The glycosyl residues may also be attached to the naked analogue peptide of the invention by chemical means as is known in the art.

A single CDR is the smallest part of an antibody known to be capable of binding to an antigen. The sequences of the V_L and V_H CDRs of the BrE-3 exemplary analogue is shown in Tables 47 and 48 below. Thus, small peptides that have the sequence of a single CDR can bind antigen and are, therefore,

suitable for imaging tumors in vivo. A CDR attached to an effector peptide may be synthesized chemically or recombinantly encoded in a DNA segment. Such small molecules have great tumor penetration and extremely rapid clearing properties when compared to larger antibody fragments. In some cases, it is more convenient to produce these small molecules by chemical synthesis, as is known in the art, rather than by fermentation. In many cases, these small peptides are completely non-immunogenic and an immune response, such as the HAMA response, is altogether avoided. Also preferred are 2 and 3 CDR units per chain operatively linked to one another by 1 to 10 or more amino acids and up to the entire inter-CDR segment length as positioned in the variable regions.

Heavy and light chain analogue variable regions may be obtained individually or in V_H/V_L pairs, or attached to an effector peptide such as a constant region(s) or portions thereof, a drug, an enzyme, a toxin, a whole antibody, or any other molecule or radioisotope. The fragments of the analogue variable regions may be synthesized chemically as is known in the art or from the DNA segments encoding the non-human variable regions. This may be attained by PCR amplification of the DNA with primers synthesized to contain the desired mutation(s) as is known in the art. Similarly, the fragments encoding analogue variable regions may be synthesized chemically or obtained by established cloning methods of restriction digestion, ligation, mutagenesis, and the like, as is known in the art.

There are advantages to using the different molecular variants of the analogue peptide depending on the specific applications for which they are intended, some of which are listed below.

- a) Smaller molecules penetrate target tissues more efficiently and are cleared from the body much more rapidly than larger molecules.
- b) Single chain molecules can be manipulated and synthesized more efficiently than multiple chain molecules.
- c) Many of these variants can be synthesized efficiently and inexpensively in bacteria, including the non-glycosylated analogues.
- d) Bi-functional or multifunctional molecules may carry polypeptide effectors, such as enzymes, toxins, radioisotopes, drugs, and other molecules, to a target tissue.

The following list encompasses exemplary analogue peptides of the invention engineered with molecules derived from antibodies or antibody fragments. These analogue peptides, among others, are suitable for the practice of this invention. A more extensive list of polypeptide constructs may be found in O'Kennedy, R., and Roben, P. (O'Kennedy, R., and Roben, P., "Antibody Engineering: an Overview", *Essays Biochem. (England)* 26:59-75 (1991)).

The analogue peptides and hybrid peptides of this invention encompass CDRs and/or analogue variable regions, monoclonal antibodies, antibody

fragments such as Fab, Fab', (Fab')₂, and fragments thereof, CDRs, constant regions, single or multiple-domain and catalytic fragments, bi-functional or multifunctional combinations thereof, enzymes, peptide hormones, molecules such as drugs and linkers, transmitters, and toxins, among others. These are suitable for imaging, therapy, and diagnostics.

Single-Chain Antigen-Binding Polypeptides

A method for constructing single chain antigen-binding polypeptides has been described by Bird et al. (Bird, R.E., et al., Science 242:243-246 (1988); Bird, R.E., et al., Science 244:409 (1989)). Single Chain F₁ (scF₁ or sF₁) are single chain analogue peptides containing both V_L and V_H with a linker such as a peptide connecting the two chains (V_L-linker-V_H). The engineering may be done at the DNA level, in which case knowledge of the sequence is required. These analogue peptides have the conformational stability, folding, and ligand-binding affinity of single-chain variable region immunoglobulin fragments and may be expressed in E. coli. (Pantoliano, M.V., et al., Biochem. (US) 30:10117-25 (1991)). The peptide linker binding the two chains may be of variable length, for example, about 2 to 50 amino acid residues, and more preferably about 12 to 25 residues, and may be expressed in E. coli. (Pantoliano, M.V., et al. (1991), supra). An analogue peptide such as an scF₁ may be expressed and prepared from E. coli and used for tumor targeting. The clearance profiles for scF₁ in some situations fragments are advantageous relative to those of normal antibodies, Fab, Fab' or (Fab')₂ fragments. (Colcher, D., et al., J. Natl. Cancer Inst. 82:1191-7 (1990)). Another type of analogue peptide comprises a V_H-linker-V_L and may have about 230 to 260 amino acids. A synthetic gene using E. coli codons may be used for expression in E. coli. A leader peptide of about 20 amino acids, such that of Trp LE may be used to direct protein secretion into the periplasmic space or medium. If this leader peptide is not naturally cleaved, the sF₁ analogue peptide may be obtained by acid cleavage of the unique asp-pro peptide bond placed between the leader peptide and the sF₁-encoding region (Houston, J.S., et al., "Protein Engineering of Antibody Binding Sites: Recovery of Specific Activity in an Anti-Digoxin Single-Chain F₁ Analogue Produced in E. coli.", PNAS (USA) 85 (16):5879-83 (1988)). The construction, binding properties, metabolism and tumor targeting of the single-chain F₁ analogue peptides derived from monoclonal antibodies may be conducted as previously described (Milenic, D.E., et al., Cancer Res. (US) 51 (23 pt1):6363-71 (1991); Yokota, et al., "Rapid Tumor Penetration of a single-chain F₁ and Comparison with Other Immunoglobulin Forms", Cancer Res. (US) 52(12):3402-8 (1992)). This type of analogue peptide provides extremely rapid tumor penetration and

even distribution throughout tumor mass compared to IgG or Ig fragments Fab and F(ab')₂.

Bifunctional scF_v-Fxn or Fxn-scF_v

An example of this type of analogue peptide is a V_L-linker-V_H with an effector peptide such as a hormone, enzyme, transmitter, and the like. These hybrid analogue peptides may be prepared as described by McCarney et. al. (McCarney, J.E. et al., "Biosynthetic Antibody Binding Sites: Development of a Single-Chain F_v Model Based on Antidinitrophenol IgA Myeloma MOPC 315", J. Protein Chem. (US) 10 (6):669-83 (1991)). A bi-functional hybrid analogue peptide containing an F_v-binding fragment B of staph protein A amino terminal to a single-chain analogue F_v region of the present specificity is also encompassed and may be prepared as previously described. (Tai, M.S., et al., Biochem. 29 (35):8024-30 (1990)). In this example of a hybrid analogue peptide of this invention is a Staph. A fragment B (anti F_v) - scF_v polypeptide. The order is backward of normal cases. This FB-scF_v may be encoded in a single synthetic gene and expressed as peptide in E. coli. This analogue peptide is a good example of a useful multifunctional targetable single-chain polypeptide. A hybrid analogue peptide also comprising antibodies to a human carcinoma receptor and angiogenin is also part of this invention. Angiogenin is a human homologue of pancreatic RNase. This is an (Fab')₂-like antibody-enzyme peptide effector. Another hybrid analogue peptide comprising a V_H-CH1 heavy chain-RNase may be expressed in a cell that secretes a chimeric light chain of the same antibody. A secreted antibody of similar structure was shown to cause the inhibition of growth and of protein synthesis of K562 cells that express the human transferrin receptor (Rybak, S.M., et al., "Humanization of Immunotoxins", PNAS 89:3165-3169 (1992)).

Bi-specific Antibodies

A monoclonal antibody or antibody fragment may be incorporated into a bi-specific analogue peptide as described, for example, by Greenman et al. (Greenman, J., et al., Mol. Immunol. (England) 28 (11):1243-54 (1991)). In this example, a bi-specific F(ab')₂ comprising two (Fab')-(thioether-link)-Fab' was constructed. Bi-specific antibodies may also be obtained when two whole antibodies are attached. Another way to obtain bi-specific antibodies is by mixing chains from different antibodies or fragments thereof. In this manner the "left" branch of the bi-specific antibody has one function while the "right" branch has another.

Phage-Display Libraries

The analogue peptides in accordance with this invention may be screened with a filamentous phage system. This system may also be used for expressing any genes of antibodies or fragments thereof as well as for screening for
5 mutagenized antibody variants as described by Marks et al. (Marks, J.D., et al., "Molecular Evolution of Proteins on Filamentous Phage. Mimicking the Strategy of the Immune System", J.Mol. Biol. (England) 267 (23):1607-10 (1992)). A library of V_H and V_L genes or analogue thereof may be cloned and displayed on the surface of a phage. Antibody fragments binding specifically to several
10 antigens may be isolated as reported by Marks (Marks, J.D., "By-Passing Immunization. Human Antibodies from V-gene Libraries Displayed on Phage", J. Mol. Biol. (England) 222 (3):581-97 (1991)).

Covalent Oligosaccharide Modifications

The present analogue peptides alone or as hybrid peptides comprising
15 antibodies and fragments thereof may be, e.g., covalently modified utilizing oxidized oligosaccharide moieties. The hybrid analogue peptides may be modified at the oligosaccharide residue with either a peptide labeled with a radioisotope such as ¹²⁵I or with a chelate such as a diethylenetriaminepentaacetic acid chelate with ¹¹¹In. The use of
20 oligosaccharides provides a more efficient localization to a target than that obtained with antibodies radiolabeled either at the amino acid chain lysines or tyrosines (Rodwell, J.D. et al., "Site-Specific Covalent Modification of Monoclonal Antibodies: In Vitro and In Vivo Evaluations", PNAS (USA) 83:2632-6 (1986)).

25 Of the analogue peptides of this invention, preferred are those having the sequences ID Nos. 67 through 73, 75 through 81, 95, 96, and analogues thereof wherein about 1 to 42 amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the species for which the analogue is intended, such as human, or fragments thereof
30 comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 amino acids, or up to the complete N-terminal region, or combinations thereof. Examples of possible substitute amino acids in all positions, including the most important positions are shown in Tables 10 and 11 above, and are
35 indicated in the columns titled BrE-3 and KC-4, "Most Preferred Analogue and Preferred Analogue" amino acids substituents. Others are also suitable as may be deduced by the method described herein. These amino acid sequences may be bound by a peptide or non-peptide linker such as is known in the art.

Examples of peptide linkers are polylysines, leucine zippers, EGKSSGSGSEJI, and (GGGGS)x3, and non-peptide polymers, among others. Effector agents: such as peptides and non-peptides may also be attached to the analogue peptides of the invention. These include non-peptide polymers, monomers, atoms, etc.,

5 which are discussed below.

Another preferred embodiment comprises a bi-functional analogue peptide having a pair of light and heavy chains of the same specificity attached to one another by a linker, such as those provided above. In another preferred embodiment, a bi-functional analogue peptide comprises one set of light and heavy chains comprising at least one xenogeneic CDR or variable region, e.g., 10 the amino acid sequence ID No. 11 or 13 of Tables 15 and 16, sequence ID No. 95 or 96 from Tables 55 and 56 below, or the respective CDRs alone or separated by 1 to 10 amino acids and up to the entire amino acid segments of their flanking sequences, and optionally with 1 to 10 amino acids of each N-terminal region, sequence ID No. 68, 70 and/or 72, alone or with flanking 15 sequences of varying length, or 76, 78 and/or 80, alone or with flanking sequences of varying length, with the modifications shown above, wherein about 1 to 42 amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the target species, or 20 fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more amino acids, and one set of light and heavy chains comprising at least one xenogeneic CDR or analogue variable region, e.g., amino acid sequence ID No. 11, 13, 95, or 96 having a different set of substitute amino 25 acids, wherein about 1 to 46 or more amino acids in the FR are substituted per chain with amino acids such as those present in equivalent positions in antibodies of the target species, or fragments comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more amino acids, or 30 fragments or combination thereof. Multi-functional hybrid analogue peptides may comprise several identical units or combinations of the above bi-functional analogue peptides of the same or different specificities or xenogeneic species. Preferred analogue peptides are those comprising murine CDRs and other regions substituted with human amino acids.

35 In another aspect, this invention provides a hybrid analogue polymer that comprises at least one anti-tumor analogue peptide and at least one effector agent operatively linked to the peptide, combinations thereof and mixtures thereof. The effector agent utilized in this invention comprises peptide polymers other than the constant region of an antibody of the same species as the CDRs.

non-peptide polymers, monomers, and atoms such as metals. In one particularly preferred embodiment, the effector agent may comprise an atom such as a radioisotope, an enzyme or a fluorescent label. These effector peptides are suited for in vivo and in vitro assays because they permit the identification of complexes formed by the peptide of the invention. Radioisotopes are particularly preferred for in vivo imaging. Polypeptide labeling is known in the art (Greenwood, F.C., et al., Biochem. J. 89:114-123 (1963)). When a glycosylated polypeptide is utilized, the radiolabel may be attached to the glycosyl residue as is known in the art (Hay, G.W. et al, in Methods in Carbohydrate Chemistry, Vol 5:357, Whistler, R.L. Ed., Academic Press, NY and London (1965)). Effector agents comprising a monomer may be therapeutic, immunogenic or diagnostic agents, radioisotopes, DNA, or RNA monomers, chemical linkers, chemical chelators, transmitter molecules, combinations thereof, or combinations thereof with peptide and non-peptide polymers or copolymers and atoms. Examples of therapeutic agents are anti-neoplastic drugs such as vincristine, intercalation drugs, adriamycin, enzymes, toxins and hormones, among others. Examples of immunogenic agents are other vaccines against tumors such as r carcinomas or for others purposes. Examples of diagnostic agents are radioisotopes and enzymes, among others. Examples of therapeutic, immunogenic and diagnostic agents are toxins, vaccines, and radioisotopes, among others. Examples of radioisotopes are ^{111}In , ^{35}S , ^{90}Y , ^{188}Re , ^{225}Ac , ^{125}I and $^{99\text{m}}\text{Tc}$, among others. Examples of DNA and RNA monomers are A, T, U, G, C, among others. Examples of chemical linkers are dithiobis(succinimidyl)propionate and bis-(sulfosuccinimidyl)suberate, among others. Examples of transmitter molecules are cAMP and cGMP, among others. Examples of toxins are ricin A-chain and abrin A-chain, among others.

When the effector agent is a non-peptide polymer linked to the analogue peptide of the invention it may comprise an ester, ether, vinyl, amido, imido, alkylene, arylalkylene, cyanate, urethane, or isoprene polymers, DNA polymers, RNA polymers, copolymers thereof and copolymers thereof with peptide polymers or monomers, or have labeled atoms attached thereto. Examples of these are polyesters, polyethers, polyethyleneglycols, polyvinyls, polyamido and polyimido resins, polyethylenes, polytetrafluoroethylene, poly(ethyleneterephthalate), polypropylene, silicone rubber, isoprenes and copolymers thereof, copolymers of silicone and carbonated polylactic or polyglycolic acid or collagen, and the like. Particularly preferred are biodegradable and bioresorbable or bioabsorbable materials, which if detached from the polypeptide and left in the systemic circulation will not damage endogenous tissues. The effector agent being a peptide may comprise

antibodies such as IgA, IgG, IgM, IgE or IgG, the constant region of antibodies of a species different from the variable region or fragments thereof, and the CDRs, variable regions, Fab, Fab', (Fab')₂, fragments of antibodies of the classes described above, hormones, enzymes, peptide transmitters and whole antibodies.

5 combinations thereof, and combinations thereof with non-peptide polymers, copolymers, monomers and atoms such as radioisotopes. Examples of other antibodies, Fab, Fab', (Fab')₂, CDRs and variable regions thereof are those that specifically bind tumor epitopes such as those of carcinomas, including the murine BrE-3 and anti-KC-4 antibodies and others having specificities for different

10 tumor epitopes such as the BrE-1 (ATCC No. HB 9738), BrE-2 (ATCC No. HB 9795), and Mc5 antibodies, among others, and fragments thereof. All of the antibodies exemplified above selectively bind to the human mammary mucin, and more particularly to the human mammary fat globule (HMFG). However, antibodies with different specificities for antigens of the target species are also

15 encompassed. Examples of peptide transmitters and hormones suitable for use herein are insulin, growth hormone, FSH, LH, endorphins, and TNF, among others. Examples of enzymes are peroxidase, LDH, alkaline phosphatase and galactosidase, among others.

In a particularly preferred embodiment of the hybrid analogue, the

20 analogue peptide polymer of the invention comprises non-human CDRs and variable region sequences, and the effector peptide comprises the constant region of the light or heavy chains of a human antibody or fragments thereof capable of being bound by immunoglobulins of a different species selectively binding to the constant regions of antibodies, protein G or protein A, or

25 fragments having this binding capability. Also preferred is a half humanized/half chimeric or murine antibody (e.g., humanized light chain and murine or chimeric heavy chain and vice versa). In one of the most preferred embodiments, the analogue peptide(s) comprise(s) all CDRs, flanking sequences of 1 to 10 amino acids connecting them, and an N-terminal region of at least up to 10 amino

30 acids. One preferred embodiment comprises the fully humanized BrE-3HZ or HuBrE3V2, ATCC No. HB 11200 and the fully humanized HuKC-4HZ or HuKC4V2, ATCC No. HB 11455, both of which were deposited under the Budapest Treaty as an example of a best mode of this invention on November 13, 1992 and September 23, 1993, respectively. Other most preferred

35 embodiments are those having chimeric heavy chains and humanized light chains such as the HuBrE3V1, ATCC No. HB _____, deposited on November 10, 1993, and the HuKC4V1, ATCC No. HB 11454 deposited on September 23, 1993, and those having chimeric light chains and humanized heavy chains such as the HuBrE3V3, ATCC No. HB _____ deposited on November 10, 1993, and

HuKC4V3, ATCC No. HB 14456 deposited on September 23, 1993, all of which deposited under the Budapest Treaty as an example of a best mode of this invention).

The hybrid analogue polymer may comprise two heavy and two light chains, each light and heavy chain comprising at least one CDR or analogue variable region polypeptide or fragments thereof of one species and the constant region and the substitute amino acids of an antibody of a different species such as human, at least one other CDR, analogue variable region, chimeric Fab, Fab' or (Fab')₂, fragments thereof, combinations thereof, and mixtures thereof. Still more preferred is a hybrid analogue peptide comprising at least two "humanized" murine-human or chimeric antibody fragments thereof, Fab, Fab' or (Fab')₂, fragments thereof operatively linked to one another. The peptide fragments may be covalently attached to one another as is known in the art (Marchis-Mouren G., et al., "HT 29, a Model Cell Line: Stimulation by the Vasoactive Intestinal Peptide (VIP); VIP Receptor Structure and Metabolism", *Bioch.* 70 (5):663-71 (1988)), or they may be synthesized by methods known in the art (Allen, G., et al., "Production of Epidermal Growth Factor in Escherichia Coli from a Synthetic Gene", *J. Cell Sci. Suppl.* 3:29-38 (1985)).

The hybrid analogue polymer of the invention described above may have two heavy and two light analogue chains operatively linked to one another, where each pair of heavy and light chains, has specificity for a different epitope. One example of this analogue peptide is a pair of "humanized" variable region heavy and light chains of a BrE-3 analogue peptide and a pair of "humanized" variable region light and heavy chains of a KC-4 analogue peptide that are covalently attached to one another by a peptide or non-peptide polymer or a disulfide bridge, or non-covalently by means of a leucine zipper or two helical structures, and the like. Non-peptide polymers may be covalently attached to peptides by methods known in the art (Duronio, V., et al., "Two Polypeptides Identified by Interleukin 3 Cross-Linking Represent Distinct Components of the Interleukin 3 Receptor", *Exp. Hematol.* 20 (4):505-11 (1992)). In another embodiment, the invention provides a hybrid analogue peptide comprising at least one CDR or analogue variable region of the heavy chain of an antibody of a first species or fragments thereof, operatively linked to a first effector agent, and at least one CDR or analogue variable region of the light chain of an antibody of a second species or fragments thereof operatively linked to a second effector agent and combinations thereof, wherein each pair of light and heavy chains has a predetermined specificity, combinations thereof, and mixtures thereof. In another preferred embodiment of the hybrid analogue peptide, at least one CDR or analogue variable region of the heavy chain of a murine antibody or fragments

thereof and at least one CDR or variable region of the light chain of a murine antibody or fragments thereof are linked to one another by a non-peptide polymer such as an isoprene polymer or monomer. In still another preferred embodiment, the hybrid analogue peptide of the invention is one wherein at least one pair of
5 light and heavy chains comprising at least one murine CDR or analogue variable region or fragment thereof is linked to at least one other pair of light and heavy chains comprising at least one murine CDR or analogue variable region or fragment thereof. In another embodiment the two or more F₁ regions are covalently attached to one another by a peptide or non-peptide polymer or a
10 disulfide bridge, or non-covalently by means of a leucine zipper or two helical structures, and the like. In a most preferred embodiment, the analogue peptides and hybrid polypeptides of the invention have affinity and specificity for an epitope located in the most hydrophilic region of a 20 amino acid tandem repeat that makes up a large part of the polypeptide core of mammary mucin, to
15 hexamer fragments of the sequence APDTRPAPG or trimer TRP fragments shown to afford the strong binding of all five different monoclonal antibodies raised against the human mammary fat globule (Mc1, Mc5, BrE-1, BrE-2 and BrE-3). The monoclonal antibodies were shown to bind to different but overlapping polypeptide epitopes but to have different tissue and tumor specificities, to
20 quantitatively differ in their binding to tumor cells such as breast carcinoma cell lines when observed by flow cytometry and have different competition patterns for binding to the native antigen on breast carcinoma cells. Also preferred amongst antibodies utilized for the preparation of the present analogue peptide and hybrid polypeptide are those that exhibit strong binding to the hexamer
25 peptides described above or to fragments comprising a TRP trimer to tandem repeats thereof. In one most preferred embodiment, the analogue peptide comprises the humanized mutein antibody expressed by the hybridoma cell line having the ATCC Accession No. HB 11200 (BrE-3 HZ). This cell was deposited with the ATCC under the Budapest Treaty on November 13, 1992. In another
30 most preferred embodiment, the analogue peptide comprises the humanized mutein antibody expressed by the hybridoma cell line having the ATCC Accession No. HB 11455 (HuKC-4V2), deposited under the Budapest Treaty on September 23, 1993. These were deposited as the best mode of the invention known to the inventors.

35 The anti-tumor analogue peptide and/or hybrid analogue polymer of the invention are also provided as an anti-tumor composition along with a carrier or diluent, preferably a pharmaceutically-acceptable carrier or diluent. The anti-tumor analogue peptide and the hybrid polymer provided herein may be present in the composition in an amount of about 0.001 to 99.99 wt%, more preferably

about 0.01 to 20 wt%, and still more preferably about 1 to 5 wt%. However, other amounts are also suitable. Carriers generally, and pharmaceutically-acceptable carriers in particular are known in the art and need not be further described herein. The carrier may be provided in a separate sterile container or in admixture with the polypeptide. Typically, saline, aqueous alcoholic solutions, albumin-saline solutions, and propylene glycol solutions are suitable. However, others may also be utilized. When utilized for therapeutic purposes the proteic material must be of a purity suitable for human administration, and the composition may contain other ingredients as is known in the art. Examples of these are other anti-neoplastic drugs such as adriamycin and mitomycin, cytoxan, PALA and/or methotrexate, among others. However, other therapeutic drugs, carriers or diluents, immunological adjuvants and the like may be also be added. When the composition described above is utilized for in vivo imaging, it may comprise about 0.001 to 99.9 wt% analogue peptide, and more preferably about 0.01 to 25 wt% analogue peptide. Typically, when the composition is utilized for therapeutic purposes it may contain about 0.001 to 99.9 wt% analogue peptide, and more preferably about 0.01 to 30 wt% analogue peptide. When utilized for the ex vivo purging of neoplastic cells from bodily fluids such as spinal fluid, the composition may comprise about 0.0001 to 50wt%, and preferably about 0.01 to 20wt% analogue peptide. When applied to the in vitro diagnosis of tumors such as carcinomas the composition of the invention may comprise about 0.001 to 35 wt% analogue peptide, and more preferably about 0.01 to 10 wt% analogue peptide. Other amounts, however, are also suitable.

Such products find one utility in the treatment of tumors such as carcinomas, such as breast, lung, ovary, endometrial, pancreas, prostate and colon cancers, among others. The "humanized", "half humanized" and "partially humanized" analogue peptides may be used for the in vivo treatment or diagnosis of humans. The "animalized", "half animalized" and "partially animalized" analogue peptides of the invention may be utilized for the treatment of non-human species such as were described above insofar as the amino acids of the species are substituted for those of the xenogeneic amino acids present at a specific location, and any constant region present in the analogue. The present analogue peptides are particularly suitable for repeated administration to a subject and for long term therapy, such as is the case of metastases and/or the reoccurrence of tumors. Of all analogues described and encompassed herein, the ones most suitable for in vivo applications are those that exhibit low or no binding to serum antigens and to normal cells. Suitable for in vitro or ex vivo uses are those that exhibit good binding to tumor cell antigens such as the carcinoma cell antigen and weak or no binding to normal cells. Even though a

patient may have in circulation an interfering amount of a molecule that can bind the analogue peptide, the peptide may still be administered after removal of such serum molecule either by ex-vivo procedures or by administration of flush doses of the analogue peptide or fragments thereof.

5 A kit for the diagnosis of tumors such as carcinomas may comprise a composition comprising the anti-tumor analogue peptide of the invention, a solid support, anti-tumor antibody (positive control), immunoglobulins of a different species selectively binding the constant regions of the antibody, protein G or protein A, and instructions for its use. This diagnostic kit may be utilized by
10 covalently attaching the antigen or the analogue peptide of the invention or a fusion protein thereof to the solid support by means of a linker as is known in the art. In a particularly preferred embodiment, the support is coated with a polypeptide such as methylated albumin as described in US Patent No. 4,572,901, the relevant text of which is incorporated herein by reference. When
15 a biological sample is added to a well, the analogue peptide of the invention will bind any tumor antigen, such as a carcinoma antigen, present in the biological sample. If a competitive assay is utilized, to the solid supported antigen or hybrid peptide thereof are added a known amount of the analogue peptide and the sample. Thereafter, γ -globulin, protein G or protein A in labeled form may
20 be added for detection. Anti-tumor antibodies of a first species may be obtained by challenging a subject of another species with tumor cells such as carcinoma cells, the human milk fat globule mucin and the like, as is known in the art (Peterson, J.A., et al., Hybridoma 9:221 (1990); US Patent No. 4,708,930). Monoclonal antibodies may be prepared as described by Kohler and Milstein
25 (Kohler, G. and Milstein, C., "Continuous Culture of Fused Cell Secreting Antibody of Predefined Specificity", Nature 256:495-497 (1975)). Suitable for use in this invention are antibodies such as IgG, IgM, IgE, IgA, and IgD. Protein A, protein G and γ -globulin may be obtained commercially.

A diagnostic kit for detecting tumors such as carcinomas, and more
30 particularly human carcinomas is provided herein that comprises an anti-tumor composition comprising a hybrid analogue peptide and an effector agent comprising an enzyme, a radioisotope, a fluorescent label and/or a peptide comprising the constant region of an antibody of the species for which use it is intended, or fragments thereof capable of binding anti-constant region
35 immunoglobulins, protein G or A, anti-tumor antibody, anti-constant region immunoglobulins, protein G or protein A, a solid support having operatively linked thereto an antigen which specifically binds to the anti-tumor hybrid analogue peptide of the invention and the antibody, and instructions for its use. When the effector agent comprises a peptide, such as the constant region of an antibody

of the target species, the solid support may have operatively linked thereto an antibody which specifically binds to a portion of a fusion protein other than the antigen of the invention. This permits the binding of the anti-tumor analogue peptide to the antigen molecule now attached to the solid support. Any complex
5 formed between the hybrid analogue peptide of the invention and the supported tumor antigen will, thus, remain attached to the solid substrate. A competitive assay may then be conducted by addition to the solid supported antigen of a known amount of the hybrid antigen peptide and the sample. The amount of antigen present in the sample may be obtained from a dilution curve by addition
10 of anti-constant region immunoglobulins, protein G, protein A or other antibody binding molecules, e.g., labeled, to bind the hybrid analogue peptide that is now attached to the support. This kit may be used in a competitive assay where the supported antigen molecule competes with antigen in the sample for a known amount of the analogue peptide of the invention. The assay was described by
15 Ceriani, R.L., et al. (Ceriani, R.L., et al., Anal. Biochem. 201:178-184 (1992)), the relevant text thereof being incorporated herein by reference.

A tumor such as a carcinoma may be imaged in vivo and/or diagnosed by administering to a subject suspected of carrying a carcinoma the anti-tumor analogue peptide of the invention in radiolabeled form, in an amount effective to
20 reach the tumor cells and bind thereto, and detecting any localized binding of the labeled analogue peptide to the tumor. Typically, the analogue peptide of the invention may be administered in an amount of about 0.001 to 5000 mg/kg weight per treatment, more preferably about 0.01 to 5000 μ g/kg weight per treatment, and more preferably about 0.1 to 500 μ g/kg weight per treatment.
25 However, other amounts may also be utilized. Radiolabels that may be utilized are ^{111}In , ^{125}I , $^{99\text{m}}\text{Tc}$, and ^{131}I , among others. These radioisotopes may be detected with a PET scanner, NMR imaging, and radioactivity counting apparatus that are in wide use by the medical community.

The presence of a tumor such as a carcinoma may also be diagnosed in
30 vitro by contacting a biological sample with the anti-tumor analogue peptide or hybrid polypeptide of the invention to form an anti-tumor analogue peptide-antigen complex with any tumor antigen present in the sample, and detecting any complex formed. The biological sample is typically obtained from a subject such as a human suspected of being afflicted with the tumor. Suitable biological
35 samples are serum, blood, sputum, feces, lymph fluid, spinal fluid, lung secretions, and urine, among others. Clearly, any source of fluid, tissue and the like may be prepared for use in this method as is known in the art.

Neither the hybrid half chimeric/half humanized BrE-3 analogue peptide nor the chimeric BrE-3 polypeptide of the invention or the murine BrE-3 antibody

show substantially strong binding to normal tissue. The hybrid BrE-3 analogue peptide shows a binding pattern similar to the chimeric BrE-3 polypeptide and the murine BrE-3 antibody. The murine BrE-3 antibody was shown to bind with specificity to various tumors such as carcinomas of the breast, lung, ovary, bladder, and the endometrium, mesothelioma, colon, kidney, liver, pancreas, salivary glands, sarcomas and thyroid, among others. Weak binding was only shown to normal breast tissue, lung tissue, distal convoluted tubes of the kidney, acini of the pancreas and stomach mucosa (Peterson, J.A., et al. (1990), supra). The KC-4 hybrid murine peptide has tissue specificity similar to that of the murine KC-4 antibody. The KC-4 monoclonal antibody was shown to bind specifically and strongly to solid tumor tissue in the lung, colon, kidney, breast, stomach, prostate, pancreas, lymph node duct and lymphoma, and non-specifically and weakly to normal breast, kidney, and stomach tissue. KC-4 also showed some weak binding to normal tissue including spinal cord, uterus, thyroid, tongue, prostate, spleen, adrenal, lung, gall bladder, heart, lymph nodes, colon, liver, brain, testes, thymus, and placenta (U.S. Patent No. 4,708,930). In one preferred embodiment of the in vitro diagnostic method, the anti-carcinoma analogue peptide added to the biological sample comprises a labeled hybrid analogue peptide. Suitable labeling materials were described above. This method may be practiced, with the solid support containing kit described above, as a competitive assay as disclosed by Ceriani, R.L., et al. (Ceriani, R.L., et al. (1992), supra).

The present analogue peptides are also applicable to the purging of neoplastic cells, such as carcinoma cells, from biological samples, be it fluid or tissue samples. The purging of neoplastic cells from a fluid sample is part of the invention and may be practiced by contacting a biological fluid suspected of comprising neoplastic cells with the analogue peptide of the invention, which is capable of selectively binding to an antigen of the neoplastic cells and allowing the peptide to bind to the antigen, and separating the analogue peptide-cell complex from the remainder of the fluid.

This method may be utilized for purging unwanted cells ex vivo by extracting a biological sample from a patient, eliminating the neoplastic cells therefrom by separation of the analogue peptide-cell complexes or by further addition of an effector such as complement or a toxin or a radioactive label that can act upon the cell and then replenishing the purged sample to the patient. This is typically suitable for use with spinal taps where spinal fluid is rid of neoplastic cells such as carcinoma cells prior to reinjection. Other fluids may also be treated in this manner.

The present analogue peptides may also be applied to the histochemical

assessment of the presence of neoplastic cells such as carcinoma cells in a tissue obtained from a subject suspected of being afflicted by a carcinoma by methods that are standard in the art, like the preparation of tissue slices and fixation on a solid substrate to permit the application of the peptide and then the
5 assessment of any binding to neoplastic cells in the sample as indicated by the formation of complexes between the analogue peptide and antigens on or in the cells.

The growth or the size of a primary or metastasized tumor or neoplasia such as a carcinoma may be inhibited or reduced by administering to a subject
10 in a need of the treatment an effective amount of the anti-tumor hybrid analogue peptide of the invention. Typically, the hybrid analogue peptide may be administered in an amount of about 0.001 to 2000 $\mu\text{g/kg}$ body weight per dose, and more preferably about 0.01 to 500 mg/kg body weight per dose. Repeated
15 doses may be administered as prescribed by the treating physician. However, other amounts are also suitable. Generally, the administration of the hybrid analogue peptide is conducted by infusion so that the amount of radiolabel, toxin or other effector agent present that may produce a detrimental effect may be kept under control by varying the rate of administration. Typically, the infusion of one dose may last a few hours. However, also contemplated herein is the
20 constant infusion of a dose for therapeutic purposes that will permit the maintenance of a constant level of the hybrid polypeptide in serum. The infusion of the hybrid analogue peptide of the invention may be conducted as follows. Intravenous (I.V.) tubing may be pretreated, e.g., with 0.9 % NaCl and 5 % human serum albumin and placed for intravenous administration. The prescribed
25 dose of the analogue peptide may be infused as follows. Unlabeled analogue peptide may be infused initially. 30 minutes after completion of the unlabeled antibody infusion, ^{111}In -labeled and ^{90}Y labeled antibody may be co-infused. The I.V. infusion may comprise a total volume of 250 ml of 0.9 % NaCl and 5 % human serum albumin and be infused over a period of about 2 hours depending
30 on any rate-dependent side effects observed. Vital signs should be taken every, e.g., 15 minutes during the infusion and every one hour post infusion until stable. A thorough cardiopulmonary physical examination may be done prior to, and at the conclusion, of the infusion. Medications including acetaminophen, diphenhydramine, epinephrine, and corticosteroids may be kept at hand for
35 treatment of allergic reactions should they occur. The administration of the hybrid analogue peptide of the invention may be repeated as seen desirable by a practitioner. Typically, once a first dose has been administered and imaging indicates that there could be a reduction in the size of the tumor, whether primary or metastasized, repeated treatments may be administered every about

1 to 100, and more preferably about 2 to 60 days. These repeated treatments may be continued for a period of up to about 2 years, and in some circumstances even for longer periods of time or until complete disappearance of the tumor(s). The administration of the hybrid analogue peptides of this invention is typically
5 more useful for therapeutic purposes when a primary tumor has, for example, been excised. Thus, it is primarily, for mopping up after surgical intervention or in cases of cancerous metastases that the present method is of most use.

A pure, isolated analogue polydeoxyribonucleotide that comprises an analogue oligodeoxyribonucleotide encoding the analogue peptide or hybrid
10 peptide of this invention, including all redundant sequences may be applied to the preparation of the peptide. In one preferred embodiment, the analogue polydeoxyribonucleotide of the invention comprises a DNA sequence selected from the group consisting of DNA sequence ID No. 64 of Table 45, or DNA sequence ID No. 65 of Table 46, or DNA segments encoding the CDR fragments
15 sequence ID No. 68, 70 and/or 72, or 76, 78 and/or 80 flanked by 1 to 10 or more amino acids, and the N-terminal fragment of 1 to 10 or more amino acids of human origin. The above DNA sequences may be cloned for expression under the same promoter. Similarly preferred are also the DNAs having the sequence ID No: 93 and/or 94 of Tables 52 and 53 and the segments encoding their CDR
20 fragments alone or separated by DNA segments encoding 1 to 10 or more flanking amino acids and/or terminated by DNA segments encoding 1 to 10 or more amino acids of the N-terminus.

Also provided herein is a hybrid vector that comprises a vector having the analogue polydeoxyribonucleotide of this invention operatively linked thereto.
25 Typically, vectors capable of replication both in eukaryotic and prokaryotic cells are suitable. When the preparation of a glycosylated analogue polypeptide is desired the vector should be suitable for transfection of eukaryotic host cells. In one preferred embodiment, the hybrid vector comprises the analogue polydeoxyribonucleotide and a polydeoxyribonucleotide comprising an
30 oligodeoxyribonucleotide encoding an effector peptide, the effector peptide-encoding polydeoxyribonucleotide being operatively linked to the vector. As already indicated, the various DNA sequences may be cloned for expression under the same promoter. In addition, the polydeoxyribonucleotide encoding the effector polypeptide may also be cloned for expression under the same promoter.

35 This invention also encompasses a host cell that has been transfected with the hybrid vector described above. Suitable hosts are prokaryotic and eukaryotic hosts such as bacteria, yeast, and mammalian cells such as insect cells and non-producing hybridoma cells, among others. Suitable vectors and/or plasmids for the transfection of each one of these types of hosts are known in

the art and need not be further described herein. Also known in the art are methods for cloning DNA sequences into each one of these types of vectors and for transfecting the different types of host cells. Particularly preferred is the cell line having the ATCC Accession No. HB 11200 (BrE-3 HZ).

5 Polyribonucleotides may be obtained by transcription of the polydeoxyribonucleotides described above as is known in the art. Provided herein are analogue polyribonucleotides comprising analogue oligoribonucleotides encoding at least one CDR or an analogue variable region or fragments thereof, combinations thereof, and combinations thereof with an effector peptide may be
10 prepared by cloning the desired DNA segments and then transcribing the thus obtained hybrid polydeoxyribonucleotide into the corresponding RNA sequences.

The analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a neoplastic cell, or is released by the cell, may be produced by a method that comprises cloning the analogue
15 polydeoxyribonucleotide of the invention into a vector to form a hybrid vector, transfecting a host cell with the hybrid vector and allowing the expression of the anti-tumor analogue peptide, and isolating the anti-tumor polypeptide or mixtures thereof. The DNA segment encoding the analogue polypeptide may be obtained by chemical synthesis or by site-specific modification of the sequence encoding
20 the variable region of the xenogeneic species by PCR amplification with specifically designed primers as is known in the art. The fragment DNAs may also be prepared by PCR with primers that introduce a stop codon at a desired position as is known in the art. Preferably, the cloning and transfection steps are conducted by cloning polydeoxyribonucleotides encoding the analogue peptides
25 selected from the group comprising at least one CDR or analogue variable region of the heavy or light chains of the xenogeneic species, antibodies thereof, or fragments thereof. The method may further comprise allowing the expressed analogue peptides to interact with one another to form double chain analogue peptides, one or both analogue peptide chain comprising at least one xenogeneic
30 CDR or variable region of the light or heavy chain of the antibody or fragment thereof modified as described above. Still part of this invention is a method of producing a hybrid analogue peptide comprising an effector peptide and an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a tumor cell such as a carcinoma cell or that is released by the cell,
35 the method comprising transfecting a host cell with the hybrid vector of this invention carrying a DNA sequence encoding the hybrid analogue peptide, allowing the expression of the hybrid analogue peptide, and isolating the hybrid analogue peptide or mixtures thereof. The techniques for obtaining mRNA, conducting reverse transcription and PCR amplification of DNA, chemical

synthesis of primers, cloning DNA sequences into a vector, transfecting a host cell, and purifying polypeptides from a culture medium are known in the art and need not be further described herein.

Another aspect of this invention relates to an anti-idiotypic peptide that

5 comprises polyclonal antibodies raised against anti-tumor antibodies, the analogue peptide of the invention, monoclonal antibodies thereof, fragments thereof selected from the group consisting of CDRs, Fab, Fab', (Fab')₂, and variable region fragments and fragments thereof, analogues thereof selected from the group consisting of Fab, Fab', (Fab')₂, and variable regions thereof,

10 wherein about 1 to at least 46 amino acids in the FRs are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in human antibodies, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to 10 or more amino acids, alone or with an N-terminal fragment of about 1 to 10 or more

15 amino acids. The technique for obtaining anti-idiotypic polypeptides is known in the art and need not be further described herein (Nisonoff, A. and Lamoyi, "Implication of the Presence of an Internal Image of an Antigen in Anti-Idiotypic Antibodies: Possible Applications to Vaccine Production", Clin. Immunol. Immunopathol. 21:397-406 (1981)). Moreover, the technique for producing

20 hybridomas secreting monoclonal antibodies of a certain specificity is also known in the art (Kohler, G. and Milstein, C. (1975), supra). Techniques for obtaining different antibody fragments were described above or are known in the art and need not be further described herein (Wilbanks, T., et al., "Localization of Mammary Tumors In Vivo with ¹²⁵I-Labeled Fab Fragments of Antibodies Against

25 Murine Mammary Epithelial (MME) Antigens", Cancer 48:1768-1775 (1981)). The techniques for modifying peptides to obtain the analogue peptides of this invention have been described above or are known in the art.

The hybrid anti-idiotypic polymer may further comprise an effector agent operatively linked to the anti-idiotypic polypeptide. Effector agents suitable for

30 use herein were described above for the anti-tumor analogue polypeptide of the invention are also suitable for use with the anti-idiotypic polypeptide. Preferred hybrid polymers are polyclonal antibodies raised against the anti-tumor monoclonal antibodies or the analogue peptide of the invention, and a monoclonal antibody obtained by fusion of a B-cell producing an antibody having

35 specificity for the analogue peptide of the invention and an immortalized cell line. Also preferred are fragments of the monoclonal antibody such as Fab, Fab', (Fab')₂, and variable region fragments, analogues and fragments thereof as described above, and CDRs. Also, as described above for the anti-tumor polypeptide analogue, preferred are combinations of the above fragments and

analogues and combination of the fragments with whole antibodies and analogues thereof. In another preferred embodiment, the anti-idiotypic polymer comprises an analogue variable region of a monoclonal antibody linked to a peptide comprising the hexamers or trimers described above or tandem repeats

5 thereof.

DNA and RNA segments encoding the anti-idiotypic polymer and hybrid polymer, a hybrid vector having the DNA operatively linked thereto and a host cell transfected with the hybrid vector are also contemplated herein.

Also provided herein is an anti-tumor vaccine that comprises the anti-
10 idiotypic polypeptide of the invention, and a pharmaceutically-acceptable carrier. Typically, the anti-idiotypic polypeptide is present in the composition in an amount of about 0.001 to 99.99 wt%, and more preferably about 0.01 to 50 wt% of the composition. However, other amounts are also suitable. Pharmaceutically-acceptable carriers are known in the art and need not be further
15 described herein. The vaccine provided herein may further comprise other ingredients such as adjuvants, and the like. Examples of adjuvants are SAF-1 and Freund's, among others. Suitably, other ingredients typically used for the preparation of vaccines may also be utilized herein. In one embodiment, the vaccine of the invention may be provided in unit form as a powder or in a
20 diluent. In another embodiment, it may be provided in powder form in a sterile container comprising a plurality of doses for preparation prior to utilization. Diluents that are suitable for the preparation of a formulation that may be administered to a patient by injection are known in the art. Examples were provided above.

25 An anti-tumor vaccination kit is also provided by this invention that comprises the vaccine described above and a diluent in separate sterile containers, and instructions for its use.

An effective amount of the anti-idiotypic polypeptide or hybrid polypeptide described above may be administered to vaccinate a human. Typical
30 amounts administered to vaccinate a human are about 0.001 to 5000 mg/kg body weight/dose, more preferably about 0.1 to 5000 μ g/kg body weight/dose. The anti-idiotypic vaccine of the invention may be administered repeatedly in order to boost the active immunization produced by the first dose. An anti-idiotypic antibody very likely resembles the epitope on the neoplastic cell to
35 which the anti-tumor antibody binds. Thus, it may be utilized for the production of an immunological response by a subject such as a human or other mammals against its own neoplastic cells.

When an anti-idiotypic polypeptide of, e.g., non-human origin is administered to a, e.g., human, it may produce a somewhat detrimental

response. Accordingly, in theory, the smaller the non-human amino acid sequence the anti-idiotypic polypeptide contains, the lesser the immunogenic response to its xenogeneic sequences it will elicit in a human. Accordingly, preferred anti-idiotypic polypeptides are those containing at least one CDR or
5 variable region of a non-human antibody binding specifically to the anti-tumor polypeptide described herein, optionally as a hybrid polypeptide. Also preferred are human anti-idiotypic antibodies, CDR and variable fragments thereof, and fragments thereof that are operatively linked to an effector agent comprising a human polypeptide that may include the constant region of a human antibody
10 and fragments thereof, non-peptide polymers, monomers and atoms that may be radiolabeled as described above. Other types of constructs are also possible, several of which were described above.

Peptides comprising the sequence APDTRPAPG or fragments thereof comprising hexamers with the trimer TRP or TRP by itself or tandem
15 repeats thereof may also be utilized for the preparation of the fusion protein, particularly as part of the antigenic peptide. The peptide comprising the hexapeptide or tripeptide sequences may be utilized as a tandem repeat comprising up to about 10,000 repeats of the basic unit, and in some instances up to about 500,000 repeats. In another embodiment, peptides comprising one
20 or more hexapeptides or tripeptides may be operatively linked to other polypeptide sequences of related or unrelated function, which sequences provide bulk that aids the clearance through the liver and/or kidneys of the immunological complex formed between the circulating unbound or residual antibody or polypeptides utilized for the therapy of neoplasias such as carcinomas and the
25 hexapeptide. The peptides comprising the hexapeptide or tripeptide may also be provided as a hybrid analogue peptide with other analogue peptides described above. In the absence of such treatment, the therapeutic antibody, which may carry a radioisotope, a toxin or other therapeutic molecules, may remain in the circulation for several days and in some instances weeks. This, in the case of a
30 radioactively labeled antibody or analogue of the invention may produce extensive damage, which is highly detrimental to the health of the patient, and in some instances lethal.

Thus, the serum concentration of a circulating antibody or polypeptide that binds to an antigen on the surface or in the cytoplasm of tumor cells such
35 as carcinoma cells or released by the cells may be lowered by administering to a subject the anti-idiotypic polypeptide described above, in an amount effective to bind the circulating polypeptide, to thereby accelerate its clearance. Typically, the anti-idiotypic polypeptide or hybrid polymer are administered in an amount of about 0.001 to 5000.00 mg/kg body weight/dose, more preferably about 0.01

to 500 mg/kg body weight/dose. However, other amounts may also be utilized. The administration of the anti-idiotypic polypeptide may be infusion as described above.

The growth or the size of a primary or metastasized tumor may be inhibited or reduced by administering to a subject in need of the treatment an effective amount of an antibody or an anti-tumor hybrid analogue peptide comprising an effector agent selected from the group consisting of radioisotopes, therapeutic drugs and vaccines, and an anti-tumor polypeptide which specifically binds to an antigen on the surface or in the cytoplasm of a tumor cell or released by the cell, allowing the hybrid polypeptide to reach the tumor and the polypeptide to bind thereto, and administering to the subject an amount of the anti-idiotypic polypeptide of the invention effective to bind residual or unbound circulating hybrid analogue peptide to thereby accelerate the clearance of the hybrid polypeptide.

Having now generally described this invention, the same will be better understood by reference to certain specific examples, which are included herein for purposes of illustration only and are not intended to be limiting of the invention or any embodiment thereof, unless so specified.

EXAMPLES

Example 1: Methods Utilized

The procedures utilized herein for the reverse-transcription (RT) of RNAs encoding the variable regions and the subsequent amplification of the cDNAs by the polymerase chain reaction (PCR) have been described (Orlandi, R., et al., "Cloning Immunoglobulin Variable Domains for Expression by the Polymerase Chain Reaction", PNAS (USA) 86:3833-3837 (1989); Coloma, M.J., et al., "Primer Design for the Cloning of Immunoglobulin Heavy-Chain Leader-Fvs from Murine Hybridoma Cells Using the PCR", Bio.Techniques 11:152-156 (1991); Gavilondo-Cowley, J.V., et al., "Specific Amplification of Rearranged Immunoglobulin Fv Genes from Murine Hybridoma Cells", Hybridoma 9:407-417 (1990)).

Total RNA is an adequate substrate for RT-PCR. Polyadenylated RNA was utilized herein, however, because it contains only minor levels of contaminating ribosomal RNA and practically no DNA. The polyadenylated RNA was isolated with a Fast Track mRNA isolation kit (Invitrogen Corporation, San Diego, CA).

The oligonucleotides were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City, CA). A PCR murine Ig

primer set was purchased from Novagen (Madison, WI), and complementary DNA (cDNA) was prepared with an RNA PCR kit (Partin Elmer-Cetus, Norwalk, CT).

5 PCR DNA fragments were cloned directly into pCR1000, using a TA cloning kit (Invitrogen Corporation, San Diego, CA). Plasmid DNA was isolated with a kit purchased from Qiagen (Tchapsworth, CA), and DNA sequencing was conducted with a Sequenase 2.0 DNA sequencing kit (United States Biochemical, Cleveland, Ohio) using aqueous 5'-³²SdATP at 600 mCi/mmol (Amersham Corporation, Arlington Heights, Ill).

10 Sequence analyses were performed on a Macintosh computer using the program GeneWorks (IntelliGenetics, Inc, Mountain View, CA).

Example 2: Tissue Culture Media

15 SP2/O-Ag14 cells (Shulman, M., et al. (1978), below) were cultured either in Dulbecco's modified Eagle's medium (DME): fetal bovine serum (FBS), 90:10 (v/v) or in a mixture of DME:RPMI:FBS, 45:45:10 (v/v/v) or RPMI:FBS, 90:10 (v/v). Penicillin and streptomycin were added to prevent bacterial growth. When serum-free medium was utilized, it contained an HL-1 supplement as directed by the manufacturer (Ventrex Labs., Portland, ME). The freezing medium was 10% DMSO in bovine serum.

Example 3: PCR Primers

20 Primers and primer mixtures MulgxV_L5'-C, MulgIV_L3'-1, MulgV_H5'-C, MulgV_H5'-F, and MulgV_H3'-2 were part of a primer set purchased from Novagen. Their sequences may be obtained from Novagen. Other primers were synthesized by the inventors. These sequences are shown in Table 12 below.

Table 12: Synthetic Primers

25	JO2- T GAA GCT TGC TCA CTG GAT GGT GGG AA (Seq. ID No: 1);
	JO3- AGA TGG GGG TGT CGT TTT GG (Seq. ID No: 2);
	JO4- GCT TGA ATT CCA GGG GCC AGT GGA TAG A (Seq. ID No: 3);
30	V _H 1BACK (*) - AG GT(CTG) (CA)A(GA) CTG CAG (CG)AG TC(ITA) GG (Seq. ID No: 4);
	JO14- ATG TAC TTG GGA CTG AAC TAT GTC TT (Seq. ID No: 5).

35 * Oriandi, R., et al. (Oriandi, R., et al. "Cloning Immunoglobulin Variable Domains for Expression by the Polymerase Chain Reaction", PNAS (USA) 86: 3833-3837(1989)).

Example 4: Cloning of Chimeric BrE-3 Antibody Polydeoxyribonucleotide

Two expression vectors pAG4622 and pAH4604 were utilized herein (Coloma, M.J., et al., "Novel Vectors for the Expression of Antibody Molecules Using Variable Regions Generated by PCR", J. Immunol. Methods 152:89-104 (1992)). These were kindly provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). The construction and expression of chimeric genes was performed as described by Coloma et al. (Coloma, M.J., et al. (1992), supra).

Oligonucleotides were synthesized and used in a PCR mixture to produce variable heavy (V_H) and variable light (V_L) fragments with the correct ends for insertion into the pAG4622 and pAH4604 expression vectors. These sequences are shown in Table 13 below.

Table 13: Synthesized Oligonucleotides

JO16 (sense V_H leader) - GGG GATATC CACC ATG TAC TTG GGA CTG AAC TAT GTC TTC A (Seq. ID No: 6);	
JO17 (sense V_L leader) - GGG GATATC CACC ATG AAG TTG CCT GTT AGG CTG TTG GT (Seq. ID No: 7);	
JO18 (anti-sense JH3) - GGG GCTAGC TGC AGA GAC AGT GAC CAG AGT CC (Seq. ID No: 8);	
JO19 (anti-sense J κ 1) - GGG GTCGACTTAC G TTT GAT TTC CAG CTT GGT GCC TCC A (Seq. ID No: 9).	

The original pCR1000 clones were utilized as the starting templates for the PCR. The new PCR products were cloned back into pCR1000 and their sequence confirmed. Correctly modified and amplified fragments were excised with either EcoR V and Sal I (for V_L) or with EcoR V and Nhe I (for V_H). These fragments were then ligated into the respective vectors, which had been cut open with the appropriate restriction enzymes. Both the vectors and the inserts were purified from an agarose gel prior to ligation, using the Bio101 GeneClean kit (glass beads) (La Jolla, CA).

Example 5: Expression of Murine-Human Chimeric BrE-3 Antibody

The V_H and V_L regions in the final murine-human chimeric antibody were sequenced once again to verify that their sequences were correct.

The non-producer myeloma cell line SP2/O-Ag14, (ATCC: CRL 1581, Shulman, M., et al., "A Better Cell Line for Making Hybridomas Secreting Specific Antibodies", Nature 276:269-270, (1978)) was transfected, and a chimeric antibody were isolated as described by Coloma, M.J. et al. (1992), with the following modification. The selection was only undertaken for the uptake of hisD by adding 5mM histidinol to the medium and readjusting the pH to 7.4 with

NaOH.

Example 6: Production of Transfected Hosts

After ten days, the stable transfectant colonies were clearly established at a frequency of approximately 10^{-6} . The colonies were transferred to a normal medium (without histidinol) and the supernatants from stable transfectants were assayed for the presence of the murine-human chimeric BrE-3 antibody. This was done by capturing the secreted murine-human chimeric BrE-3 antibody with a plate-bound goat anti-human- κ antibody and developing with goat anti-human- γ antibody as described by Coloma et al. with the following modification. The secondary antibody utilized herein was radiolabeled with ^{125}I .

Example 7: Confirmation of Murine-Human Chimeric BrE-3 Antibody Expression

The supernatants were assayed for binding to human milk fat globule (HMFG) as described by Ceriani et al. (Ceriani R.L., et al., "Diagnostic Ability of Different Human Milk Fat Globule Antigens in Breast Cancer", *Breast Cancer Res. Treat.* 15:161-174 (1990)). HMFG was bound to the microtiter plates as described previously (Ceriani R.L., "Solid Phase Identification and Molecular Weight Determination of Cell Membrane Antigens with Monoclonal Antibodies", in: *Monoclonal antibodies and functional cell lines. Progress and application*, Bechtol, K.B., McKern, T.J., and Kennett, R., Eds., Plenum Press, New York, pp 398-402 (1984)).

Most colony supernatants were positive by both assays. The colonies that secreted the highest level of chimeric antibody into the supernatants, as determined by these assays, were subcloned and subsequently adapted to serum-free medium for the purification of antibody.

Example 8: Determination of Affinity Constants

The antibody-antigen affinity constants for the murine-human chimeric antibody anti-human milk mucin and the whole murine antibody were determined by obtaining the reciprocal value of the concentration of competing unlabeled monoclonal antibody giving 50% binding as described by Sheldon et al. (Sheldon, K., et al., "Characterization of Binding of Four Monoclonal Antibodies to the Human Ovarian Adenocarcinoma Cell Line HEY", *Biochem. Cell Biol.*, 65: 423-428, (1987)). The protocol for the assay was as follows.

Microtiter plates (Dynatech, Chantilly, VA) were prepared using successive layers of methylated BSA, glutaraldehyde, anti- β -galactosidase and the bacterial fusion protein 11-2 (a hybrid of β -galactosidase and human mammary mucin) as described in Ceriani et al. (Ceriani, R.L., et al., "A Novel

Serum Assay for Breast Cancer Epithelial Antigen Using a Fusion Protein". Anal. Biochem. 201:178-184 (1992). Each well contained 388ng of the 11-2 fusion protein. To each well were added 25 μ l 125 I -BrE-3 (ATCC No. HB 10028) in RIA buffer (10% bovine calf serum, 0.3% triton X-100, 0.05% sodium azide pH7.4, in phosphate buffered saline), and competed with 25 μ l of either unlabeled murine antibody or murine-human chimeric antibody in RIA buffer at final concentrations in the nanomolar range.

Iodinations were performed with 125 I (17 Ci/mg, Nordion International Inc., Kanata, Ontario, Canada). 50 micrograms of monoclonal antibody BrE-3 (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described by Ceriani, R.L. and Blank, E.W., (Ceriani, R.L., and Blank, E.W., "Experimental Therapy of Human Breast Tumors with 131 I-Labeled Monoclonal Antibodies Prepared Against the Human Milk Fat Globule", Cancer Res. 48:4664-4672 (1988)). When the counts of bound radiolabeled anti-BrE-3 murine antibody were plotted on the Y axis and the logarithm of the nanomolar (nM) concentration of competing unlabeled anti-BrE-3 murine antibody or murine-human chimeric antibody were plotted in the X axis, both curves overlapped within 5% error (Figure not shown). This proves that the variable region's affinity characteristics have been preserved.

Example 9: Affinity Binding Constants for BrE-3 Murine and Murine-Human Chimeric Antibody

The purified murine-human chimeric BrE-3 antibody and purified murine BrE-3 antibody gave similar competition curves when tested against 125 I-labeled murine BrE-3 binding to its antigen. The affinity binding constants of the murine antibody and the murine-human chimeric antibody were determined in independent competition assays as described in Example 8 above. The values of the constants are $2.68 \times 10^9 \text{ M}^{-1}$ and $3.75 \times 10^9 \text{ M}^{-1}$ for the hybrid chimeric BrE-3 polypeptides and for the murine BrE-3 antibody, respectively. These values are not distinguishable at a 95% confidence interval.

Example 10: Amplification of cDNAs Encoding BrE-3 Antibody Variable Regions

The cDNAs that encode the anti-BrE-3 murine immunoglobulin variable domains (V_H and V_L) were prepared by reverse transcription and PCR amplification (RT-PCR) from polyadenylated RNA isolated from 10^6 BrE-3 hybridoma cells by the following procedure. The JO2, JO3, JO4, JO14 and V_H 1BACK primers were synthesized, and their sequences shown in Example 3 above. Other primers were purchased from Novagen. With the exception of V_H 1BACK, which is a framework-specific primer, all sense primers are specific

for the leader peptide region. All anti-sense primers are specific for the constant regions. The degenerate λ chain of the specific primer MulgVL3'-1 (from Novagen), was used to isolate the κ chain cDNA clones because of the similarity of the λ and κ . An identical κ chain clone was isolated with primer JO2 which is specific for the κ chain constant domain. The V_H region cDNA could not be isolated with the available leader peptide primers. Thus, the V_H 1BACK primer was used, which yielded the V_H cDNA γ 72. The leader-peptide primer JO14 was then designed by extrapolating from the framework sequence of γ 72, using cataloged nucleotide sequences (Kabat, E.A., et al., "Sequences of Proteins of Immunological Interest", U.S. Dept. Health and Human Services, NIH publication No. 91-3242, 5th Edition (1991). After sequential PCR reactions, this new primer yielded the complete V_H framework cDNA. This information is summarized in Table 14 below.

Table 14: Primer Combinations for PCR Amplification

	Clone No.	Sense Primers	Antisense Primers
15	V_L 152	MulgVL5'-C	JO2
	164	MulgVL5'-C	MulgVL3'-1
20	V_H γ 72	V_H 1BACK	(JO3 or JO4)
	1012	JO14(1 st PCR)	JO3
	1043	JO14(2 nd PCR)	JO4
25		JO14(1 st PCR)	JO3
		(MulgVH5'-C + MulgVH5'-F) (2 nd PCR)	MulgVH3'-2

Example 11: Isolation of Amplified BrE-3 V_L and V_H cDNA and Sequences

The PCR products were cloned without prior purification into pCR1000 (Invitrogen) and sequenced in both directions. Clones 152, 164, 1012, and 1043 were isolated independently during different RT-PCR runs. The sequences of V_L clones 152 and 164 were found to be identical, as were the sequences of the V_H clones 1012, 1043. The V_H and V_L DNA sequences and their derived protein sequences are shown in Tables 15 and 16 below.

Table 15: BrE-3 V_L Nucleotide and Derived Protein SequencesDNA Sequence

ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG TTG TTC TGG ATT CCT GCT TCC ATC AGT GAT GTT GTG ATG ACC CAA ACT CCA CTC TCC CTG
 CCT GTC AGT CTT GGA GAT CAA GCT TCC ATC TCT TGC AGA TCT AGT CAG AAC CTT GTA CAC AAC AAT GGA AAC ACC TAT TTA TAT TGG TTC
 CTG CAG AAG TCA GGC CAG TCT CCA AAG CTC CTG ATT TAT AAG GCT TCC ATC CCA TTT TCT GGG GTC CCA GAC AAG TTC AGT GGC AGT GGA
 TCA GAG ACA GAT TTC ACA CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT CTG GGA GTT TAT TTC TGC TTT CAA GGT ACA CAT GTT CCG TGG
 ACG TTC CGT GGA GGC ACC AAG CTG GAA ATC AAA C (Seq. ID No: 10)

Amino Acid Sequence

M R I P V F I I V I L F W I P A S I S D I V V M T Q T P L S L
 P V S L G D Q A S I S C R S S O N L V H N H G N T Y L Y W F
 L Q K S G Q S P K L L I Y R A S I R F S G V P D R F S G S G
 S E T D F T L K I S R V E A E D L Q V Y F C F O O T H V P W
 I F Q Q G T E L E I E (Seq. No: 11)

Table 16: BrE-3 V_H Nucleotide and Derived Protein SequencesDNA Sequence

ATG TAC TTG GGA CTG AAC TAT GTC TTC ATA GTT TTT CTC TTA AAA GGT GTC CAG AGT GAA GTG AAG CTT GAG GAG TCT GGA GGA GGC TTG
 GTG CAA CCT GGA GGA TCC ATG AAA CTC TCT TGT GCT GCT TCT GGA TTC ACT TTT AGT GAT GCC TGG ATG GAC TGG GTC CCG CAG TCT CCA
 GAG AAG GGG CTT GAG TGG GTT GCT GAA ATT AGA AAC AAA GCC AAT AAT CAT GCA ACA TAT TAT GAT GAG TGT GTG AAA GGG AGG TTCACC
 ATC TCA AGA GAT GAT TCC AAA AGT AGA GTG TAC CTG CAA ATG ATA AGC TTA AGA GCT GAA GAC ACT GGC CTT TAT TAC TGT ACT GGG GAG
 TTT GCT AAG TGG GGC CAG GGG ACT CTG GTC ACT GTC TCT GCA G (Seq. ID No: 12)

Amino Acid Sequence

M Y I G I N Y V F I V F L L K G V Q S E I V K L E E S G G G L
 V Q P G G S M K L S C A A S G F T F S D A W M D W V R Q S P
 E K G L E W V A E I R N K A N N H A T Y Y D E S V K G R F T
 I S R D D S K S R V Y L Q M I S L R A E D T G L Y Y C T G E
 F A N W G Q G T L V T V S A (Seq. ID No: 13)

The sequences were interpreted as described by Kabat et al. (1991). The residues that are shown in lower case correspond to PCR primers. The mature chains begin at D1 (V_L) and E1 (V_H), respectively. The amino-acids that are underlined are those corresponding to the CDRs. The underlined nucleotides indicate joining segments.

The framework and CDR polypeptide segments were identified according to Kabat et al. (1991), supra. V_L is a group IIx chain. Part of the CDR 3 and all of framework 4 (FR4) are encoded by J_{L1}. V_H belongs to group IIIc. CDR 3 and FR4 are encoded by J_{H3}. Little or nothing remains from an unidentified D minigene.

Thus, the CDR 3 is only 4 amino-acids long.

Example 12: Comparison of cDNA deduced Amino Acid Sequence with Directly Determined N-Terminal Fragment Sequence

5 Table 17 below shows a comparison between the cDNA-derived polypeptide sequence and the polypeptide sequence determined directly from purified murine BrE-3 monoclonal antibody.

Table 17: Comparison of cDNA-Deduced Protein Sequence with Directly Determined N-terminal Protein Sequence

10	VL	cDNA-deduced	DVVMTQTPLSLPVSLGDAQSISCRS
	VL	Protein sequence	GVVMTQTPLSLPVVLGDAQSIIXRX
15	VH	cDNA-deduced	EVKLEESGGGLVQPGGSMKLSAAS
	VH	Protein sequence	EVKLEESGGVLVQPGGSMKLSSAAS

The murine BrE-3 antibody was reduced with 5% mercaptoethanol, separated on a 10% SDS polyacrylamide gel, and electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA). Amino acid sequencing was performed directly on the immobilized bands by the Biotechnology Instrumentation Facility, University of California, Riverside. The protein sequence given here is the sequencer's best guess.

Once the variable region cDNAs were cloned, it was confirmed that, in fact, they encoded the variable regions of the murine BrE-3 antibody and not those of another antibody by comparing the cDNA-derived amino acid sequences of the cloned murine BrE-3 antibody variable region with the N-terminal sequence of the purified anti-BrE-3 murine antibody directly determined by a single run of protein sequencing. The cDNA sequences were shown to be accurate by comparison with two independently reverse transcribed clones.

30 The general agreement between the predicted and the determined amino-acid sequences shows that the cloned cDNAs encode polypeptides of the same class and subclass as the variable regions of the murine BrE-3 antibody. This indicates that the cDNAs encode authentic variable regions. The authenticity of the variable region polypeptide and, therefore, that of the murine-human chimeric BrE-3 antibody is unquestionable given that the variable regions and the chimeric antibody affinity constant are indistinguishable from those of the murine BrE-3 antibody.

Example 13: Construction of Murine-Human Chimeric Antibody Genes

The vectors used were developed by Coloma et al. (Coloma, M.J., et al. (1992), supra) and kindly provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). Both vectors were derived from pSV2 (Mulligan, R.C., and Berg, P., "Expression of a Bacterial Gene in Mammalian Cells", Science 209:1422-1427 (1980)), and contain genomic fragments encoding either the heavy or the light chain constant domains. The vectors accept cDNAs that encode the F₁ regions. To ligate the F₁ cDNAs to the vectors, restriction ends were added to the cDNAs in a set of PCR reactions, using the JO16, JO17, JO18 and JO19 primers.

The pAG4622 light chain vector contains the human κ chain constant region, including the J-C intron. It encodes xanthine-guanine phosphoribosyltransferase or gpt (Mulligan, R.C., and Berg, P., "Selection for Animal Cells that Express the Escherichia Coli Gene Coding for Xanthine-Guanine Phosphoribosyltransferase", PNAS (USA) 78:2072-2076 (1981)) as a dominant selectable marker. It accepts the murine VL cDNA between the ribosome binding site (Kozak, M., "Compilation and Analysis of Sequences Upstream from the Translational Start Site in Eukaryotic mRNAs", Nucleic Acids Res. 12:857-872 (1984)), which is preceded by the VH promoter from the anti-dansyl murine monoclonal antibody 27.44 (Coloma, M.J., (1992), supra), and the J-C intron. The J-C intron contains the κ chain enhancer (Potter, H., et al., "Enhancer-Dependent Expression of Human κ Immunoglobulin Genes Introduced into Murine Prep-B Lymphocytes by Electroporation", PNAS (USA) 81:7161-7165 (1984); Emorine, L., et al., "A Conserved Sequence in the Immunoglobulin J Kappa-C Kappa Intron: Possible Enhancer Element", Nature 304: 447-449 (1983)).

The pAH4604 heavy chain vector contains the gene for the heavy chain $\gamma 1$ constant region, but no J-C intron. It encodes histidinol-dehydrogenase or hisD (Hartman, S.C. and Mulligan, R.C. Two Dominant-Acting Selectable Markers for Gene Transfer Studies in Mammalian Cells", PNAS (USA) 85:8047-8051 (1988)) as a dominant selectable marker. It accepts the murine V_H cDNA between the dansyl promoter-ribosome binding site and the constant $\gamma 1$ gene. The vector also contains an insert that encodes the heavy chain enhancer (Rabbitts, T.H., et al, "Transcription Enhancer Identified Near the Human C mu Immunoglobulin Heavy Chain Gene is Unavailable to the Translocated c-myc Gene in a Burkitt Lymphoma", Nature 306:806-809 (1983)).

The new V_H and V_L DNA fragments with appropriate restriction ends were integrated into pAH4604 and pAG4622 as described in Example 4 above. The

vectors were then electroporated (together) into SP2/0-Ag14 myeloma cells as described by Coloma et al. (1992), *supra*.

Example 14: Characterization of Murine-Human Chimeric BrE-3 Antibody

5 The supernatants from stable transfectants were assayed for the presence of the murine-human chimeric antibody as described in Examples 6 and 7 above. High producing transfectants were subcloned and subsequently adapted to grow in serum-free medium. The murine-human chimeric antibody produced by the myeloma cell line was then purified from the culture supernatant using a
10 Sepharose 4B-protein A column (Bio-Rad, Richmond, CA) as described in Ey, P.L., et al. (Ey, P.L., et al., "Isolation of Pure IgG1, IgG2a and IgG2b Immunoglobulins from Murine Serum Using Protein A-Sepharose", *Immunochemistry* 15:429-436 (1978)). Antibody disulfide bonds were reduced to separate the light and heavy chains by heating for 10 min at 65° in Laemmli
15 loading buffer containing 5% beta-mercaptoethanol. The separated chains were then chromatographed on a SDS polyacrylamide gel (10%). The reduced murine-human chimeric BrE-3 antibody and murine antibodies were electrophoresed in separate lanes next to 97.4, 66.2, 45.0, 31.0 and 2.5 Kdalton protein markers. Table 18 below shows the apparent molecular weights of the two bands
20 obtained for both.

Table 18: Apparent Molecular Weights of Light and Heavy Chains of Murine and Chimeric BrE-3 Antibodies

	Chimeric Antibody		Murine Antibody	
	Heavy Chain (Kd)	Light Chain (Kd)	Heavy Chain (Kd)	Light Chain (Kd)
25	50	30	49	29

The heavy and light chains of the anti-BrE-3 chimeric antibody separate as expected when electrophoresed on a polyacrylamide gel.

Example 15: Tissue Binding Studies with Chimeric BrE-3 Antibody

30 Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was conducted with the murine-human chimeric BrE-3 antibody. A control was stained with the anti-human secondary antibody only. Positive staining resulted from the use of the murine-human chimeric BrE-3 antibody, followed by the anti-human antibody
35 specific binding. (Pictures not shown).

The breast carcinoma tissue sections were stained with the supernatant

of the transfected cells using the Vectastain ABC method (Vector Labs, Burlingame, CA). The tissue stained with the goat anti-human Ig secondary antibody only shows background or non-specific staining of necrotic areas of the tissue section.

- 5 The tissue stained with murine-human chimeric BrE-3 antibody, followed by the secondary antibody, shows specific staining of the breast carcinoma cells in the breast tissue sections.

Example 16: Chimeric BrE-3 Antibody Imaging Studies

- 10 The anti-BrE-3 murine monoclonal antibody has been shown to be highly effective for imaging and for the radioimmunotherapy of breast cancers. For example, in a pharmacokinetic study of 15 breast cancer patients conducted with an ¹¹¹In-MXDTPA-BrE-3 radio-immunoconjugate (anti-BrE-3 antibody), the serum levels were low in most patients, the blood clearance correlated with the circulating antigen and the imaging results showed that about 86% of all sites
15 could be imaged (Liebes, L., et al., "Pharmacokinetics of ¹¹¹In-BrE-3 Monoclonal Antibody in Patients with Breast Carcinoma", Proc. Am. Assoc. Cancer Res. 33:216(Abs No. 1292) (1992)). A ⁹⁰Y-BrE-3 radioimmunoconjugate having similar pharmacokinetic characteristics and extrapolating the ¹¹¹In-BrE-3 dosimetry results provide a superior therapeutic agent, as well.

- 20 As with many other monoclonal antibodies, however, the clinical applications of the anti-BrE-3 murine antibody, a whole murine antibody, are limited by the HAMA response. A chimeric monoclonal antibody should give a more restricted HAMA response.

Example 17: Chimeric BrE-3 Antibody Immunogenicity

- 25 The BrE-3 variable region murine polypeptides have been cloned without the constant regions to produce less immunogenic polypeptides than the parent murine antibody. It has, moreover, been shown herein that the murine-human chimeric BrE-3 antibody lacking its original murine constant region preserves its antigen binding characteristics.

- 30 An BrE-3 antibody variable region alone or as a murine-human chimeric antibody also containing a constant human region or fragment thereof would be significantly less immunogenic to humans than the parent murine antibody. The hybrid polypeptide comprising the variable region of the BrE-3 antibody and the constant region of a human antibody was shown to preserve the original binding
35 affinity of the murine antibody. In this hybrid polypeptide, approximately 2/3 of its contiguous non-human immunogenic targets (C₁ and C₂ regions) were entirely replaced by human constant domains.

Example 18: PCR Primers used in First Isolation of Anti-KC-4 cDNAs

The PCR primers were purchased from Novagen (Madison, WI). Their sequences, reproduced from the booklet provided by Novagen, are shown in

5 Table 19 below.

Table 19: PCR Primer Sequences

-
- Mul κ V_L5'-C: sense primer mix for kappa leader.
 10 ACTAGTCGACATGAAGTTGCCTGTTAGGCTGTTGGTGCTG (Seq. ID No: 14)
 ACTAGTCGACATGGAGWCAGACACACTCCTGYTATGGGT (Seq. ID No: 15)
 ACTAGTCGACATGGATTTWCAGGTGCAGATTWTCAGCTTC (Seq. ID No: 16)
- Mul κ V_L3'-1: antisense kappa constant region.
 CCCAAGCTTACTGGATGGTGGGAAGATGGA (Seq. ID No: 17)
- 15 Mul γ V_H5'-F: sense primer mix for heavy chain leader.
 ACTAGTCGACATGRACTTTGGGYTCAGCTTGRTT (Seq. ID No: 18)
 ACTAGTCGACATGAGAGTGCTGATTCTTTTGTG (Seq. ID No: 19)
 ACTAGTCGACATGGATTTTGGGCTGATTTTTTTATTG (Seq. ID No: 20)
- Mul γ V_H3'-2: antisense gamma constant region.
 20 CCCAAGCTTCCAGGGRCCARKGGATARACIGRTGG (Seq. ID No: 21)
-

Example 19: Cloning of Murine-Human Chimeric anti-KC-4 Antibody Ribonucleotide

- The two expression vectors pAG4622 and pAH4604 described in
- 25 Example 4 were utilized. Oligonucleotides synthesized and used in a PCR to produce V_H and V_L fragments with the correct ends for insertion into the pAG4622 and pAH4604 expression vectors are shown in Table 20 below.

Table 20: PCR Primers Sequences

-
- 30 JO20 - sense kappa leader
 GGG GATATC CACC ATG AAG TTG CCT GTT AGG CTG TTG (Seq. ID No: 22)
- JO21 - antisense JK2
 CCC GTCGACTTAC G TTT TAT TTC CAG CTT GGT CCC CCC T (Seq. ID No: 23)
- JO22 - sense V_H leader
 35 GGG GATATC CACC ATG GAC TTT GGG CTC AGC TTG GTT TT (Seq. ID No: 24)
- JO24 - antisense JH3
 CCC GCTAGC TGC AGA GAC AGA GAC CAG AGT CC (Seq. ID No: 25)
-

- The original pCR1000 clones were the starting templates for the PCR and the
- 40 rest of the procedures as described in Example 4 above.

Example 20: Expression of the anti-KC-4 Chimeric Antibody Gene

15 The V_H and V_L regions in the anti-KC-4 murine-human chimeric antibody were sequenced once again to verify that their sequences were correct. The transfection of the non-producer myeloma cell line SP2/O-Ag14, (ATCC: CRL 1581) and isolation of polypeptide was conducted as described in Example 5 above.

Example 21: Production of Transfected Hosts

20 After ten days, stable transfectant colonies were clearly established at a frequency of approximately 1/10,000. The colonies were transferred to normal medium and the assays conducted as described in Example 6 above.

Example 22: Confirmation of anti-KC-4 Murine-Human Chimeric Antibody Expression

25 The supernatants were assayed for binding to human milk fat globule (HMFG) and the breast epithelial mucin (BEM) as described previously in Example 7 above. HMFG and BEM were bound to the microtiter plates as described previously by Ceriani, R.L. (1990), supra. In this radioassay the bound anti-KC-4
30 chimeric antibody (HMFG and BEM) was detected by anti-human gamma chain conjugated to ¹²⁵I. Most colony supernatants were positive by both assays. The colonies that secreted the highest level of chimeric antibody in the supernatants, as determined by these assays, were subcloned.

Example 23: Western Blot

35 75 µl of the culture supernatant was added to 20 µl of 4x Laemmli buffer and 5 µl β-mercaptoethanol and the mixture was heated at 65 °C for 15 min., in order to reduce antibody disulfide bonds and, thus, separate heavy from light chains. 20 µl of the treated sample was chromatographed in duplicate lanes on a 10% SDS polyacrylamide gel together with other antibodies that were treated
40 similarly and that were loaded for comparison. Pre-stained size markers (BioRad, Richmond, CA) were also loaded.

5 The chromatographed proteins were electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA) in 90% 30 mM CAPS pH11, 10% methanol, for 1 hour at 25 V and at 4°C. The membrane was cut into 2 parts containing identical antibody samples. The 2 membranes were immersed in 20% bovine calf serum in PBS and shaken slowly at room temperature for
10 1 hour 35 min. ¹²⁵I-labeled goat anti-human κ chain antibody was added to one membrane and ¹²⁵I labeled goat anti-human γ chain antibody to the other

membrane. Antibodies were labeled at a specific activity of approximately 10 mCi/mg using the chloramine-T method as described by Ceriani, R.L. and Blank, E.W. (1988), the labeled antibodies were diluted to 4,000 cpm/ μ l in RIA buffer.

After incubating 3 hours at room temperature the blots were washed
5 twice in TBS for 10 min each time, once in TBST (50 mM TRIS pH7.5, 3 mM EDTA 25 mM NaCl) 10 min and once more in TBS (TBS with 0.05% Tween 20) for 10 min. The membranes were dried and exposed to Kodak XAR film.

Western blot analysis of culture supernatants revealed that three
antibody chains were expressed that corresponded to the three antibody chains
10 seen in the original anti-KC-4 murine antibody. These were a heavy chain that stained with goat anti-human γ chain 125 I-labeled antibody, and two light chains that stained with goat anti-human κ chain 125 I-labeled antibody (Figure not shown).

The treatment of the original anti-KC-4 murine antibody with
15 N-glycosidase F (Boehringer Mannheim GmbH Germany) following the recommendations of the manufacturer, produced a noticeable decrease in the intensity of the "top" light chain and a concomitant increase in the intensity of the bottom light chain (Figure not shown).

The explanation for the existence of an extra light chain is that this chain
20 is glycosylated. Three lines of evidence substantiate this. First, the detection of an asparagine-linked glycosylation site in the amino acid sequence of the light chain. That is the triad NIS (Asn-Ile-Ser) in framework 3. Second, the decrease of the intensity in the putative glycosylated band after treatment with N-glycosidase F, while concomitantly the intensity of the non-glycosylated band
25 was increased. Finally, 2 corresponding light chain bands are seen in the chimeric antibody version. The extra light chain in the chimeric version cannot be a contaminant since it was specifically stained by goat anti-human κ chain antibody. It can only be a product expressed by pAG4622. Thus both light chains must have the same V_L amino acid sequence and the same human
30 constant region. These observations show that approximately half of the light chains of both the anti-KC-4 murine and chimeric antibodies are glycosylated at the asparagine-linked glycosylation site.

**Example 24: Amplification of cDNAs Encoding
anti-KC-4 Antibody F_L Regions**

35 The cDNAs that encode the anti-KC-4 murine immunoglobulin V_H and V_L were prepared as described in Example 9 above from polyadenylated RNA isolated from 100 million KC-4 hybridoma cells. All clones were obtained from independent PCRs. The sequences of the primers are given in Example 19 and

20 above. All primers are specific for either the leader peptide region or for the constant regions. The primer combinations utilized herein are shown in Table 21 below.

Table 21: Primer Combination for PCR Amplifications

	Clone No.	Primer combinations
5		
V_L	96	Mulgx V_L 5'-C + Mulgx V_L 3'-1
	107	Mulgx V_L 5'-C + Mulgx V_L 3'-1
	K1	JO20 + JO21
10	66	Mulgx V_H 5'-F + Mulgx V_H 3'-2
	209	Mulgx V_H 5'-F + Mulgx V_H 3'-2
	H3	JO22 + JO24
	H7	JO22 + JO24

Example 25: Isolation of Amplified anti-KC-4 F_H (V_L) and F_H (V_H) cDNA and Sequences

15 The PCR products were cloned, without prior purification, into pCR1000 (Invitrogen) and sequenced in both directions. The V_H and V_L DNA sequences and their derived protein sequences are shown in Tables 22 and 23 below.

**Table 22: V_L Nucleotide Sequences
anti-KC-4 V_L (k11-Jk2)**

20	
	ATG AAG TTG CCT GTT AGG CTG TTG GTG CTG ATG TTC TGG ATT CCT
	GCT TCC AGC AGT GAT GTT TTG ATG ACC CAA ACT CCT CTC TCC CTG
	CCT GTC AGT CTT GGA GAT CAA GCT TCC ATC TCT TGC AGA TCT AGT
25	CAG AGC ATT GTA CAT AGT AAT GGA AAC ACC TAT TTA GAA TGG TAC
	CTG CAG AAA CCA GGC CAG TCT CCA AAG CTC CTG ATC TAC AAA GTT
	TCC ATC CGA TTT TCT GGG GTC CCA GAC AAG TTC AGT GGC AGT GCA
	TCA GGG ACA GAT TTC ACA CTC AAT ATC AGC AGA GTG GAG GCT GAG
	GAT CTG GGA ATT TAT TAC TGC TTT CAA GGT TCA CAT GTT CCG TAC
30	AGC TTC GGA GGG GGG ACC AAG CTG GAA ATA AAA C (Seq. ID No: 26)

**Table 23: V_H Nucleotide Sequences
anti-KC-4 V_H (IIID-D9-JH)**

35	ATG GAC TTT GGG CTC AGC TTG GTT TTC CTT GTC CTT ATT TTA AAA
	GCT GTC CAG TGT GAA GTG CAG ATG GTG GAG TCT GGG GGA GTG AAG
	CCT GGA GGG TCC CTG AAA CTC TCC TGT GCA GCC TCT GGA TTC GCT
	TTC AGT AGC TAT GCT ATG TCT TGG GTT GGC CAG GAG AAG AAG CTG
	GAG TGG GTC GCA GAA ATT AGT AGT GGT GGT AAT TAC GGC TAC TAT
40	CAA GAC ACT GTG AGG GGC CGA TTC ACC AGA GAC AAT GCT AAG AAC
	ACC CTG TAC CTG GAA ATG AGC AGT CTG AGG TCT GAG GAC ACG GGC
	ATG TAT TAC TGT GCA AGG GAG GGT ATC CCG GCG TGG TTT GCT TAC
	TGG GGC CAA GGG ACT CTG GTC TCT GTC TCT GCA G (Seq. ID No: 27)

Example 26: Amino Acid Sequences of anti-KC-4
Chimeric Antibody Fv Regions

5 After the anti-KC-4 F_v region cDNAs were cloned, and sequenced, and their cDNA-derived amino acid sequence was compared with the N-terminus sequence directly determined by a single run of amino acid sequencing on purified anti-KC-4 antibody. The cDNA sequences were shown to be accurate since in both cases they were identical for clones that were prepared from
10 independent reverse transcription reactions. This confirms that the cloned cDNAs are authentic anti-KC-4 F_v regions. The sequences are shown in Tables 24 and 25 below.

Table 24: V_L Amino Acid Sequences
anti-KC-4V_L(k11-Jk2)

15

MKLPVRLLVLMFWIPASSS (Seq. ID No: 28)
FR1 DVLMTQTPLSLPVSLGDQASISC (Seq. ID No: 29)
CDR1 RSSQSIVHSNGNTYLE (Seq. ID No: 30)
FR2 WYLQKPGQSPKLLIY (Seq. ID No: 31)
20 CDR2 KVSIRFS (Seq. ID No: 32)
FR3 GVPDRFSGSGSGTDFTLNISRVEAEDLGIYYC (Seq. ID No: 33)
CDR3 FQGSHVPYT (Seq. ID No: 34)
FR4 FGGGTKLEIK (Seq. ID No: 35)

Table 25: V_H Amino Acid Sequences anti-KC-4V_H (IIIID-D9-JH3)

25 MDFGLSLVFLVLILKGVQC (Seq. ID No: 36)
FR1 EVQMVESGGGLVKPGGSLKLSAASGFAFS (Seq. ID No: 37)
CDR1 SYAMS (Seq. ID No: 38)
FR2 WVRQSPEKRLEWVA (Seq. ID No: 39)
CDR2 EISSGGNYAYYQDTVTG (Seq. ID No: 40)
30 FR3 RFTISRDNKNTLYLEMSSLRSEDAMYYCAR (Seq. ID No: 41)
CDR3 EGIPAWFAY (Seq. ID No: 42)
FR4 WGQGTLSVSA (Seq. ID No: 43)

The sequences were interpreted as described by Kabat et al. (1991), supra. The residues that are underlined correspond to PCR primers. The mature V_L and
35 V_H chains begin at amino-acids D and E of framework 1 (FR1), respectively.

Framework and CDR protein segments were identified according to Kabat et al. (1991), supra. V_L is a group II κ chain. Part of the CDR 3 and all of the

framework 4 (FR4) are encoded by Jk2. V_H belongs to group IIIId. CDR 3 and FR4 resulted from a genomic recombination involving minigenes D9 and JH3. There is an asparagine glycosylation site in the light chain in FR3. The site reads NIS (Asn Ile Ser).

5 **Example 27:** Comparison of cDNA-deduced Amino Acid Sequence with Directly Determined N-Terminal Fragment Sequence

A comparison between the cDNA-derived polypeptide sequence and the amino acid sequence determined directly on the purified anti-KC-4 monoclonal antibody was undertaken. The results are shown in Table 26 below.

Table 26: Comparison of cDNA-deduced with Directly Determined N-Terminal Amino Acid Sequences

	FIRST BAND TOP
V_H , cDNA-deduced	EVQMVESGGGLVKPGGSLKLS (Seq. ID No: 44)
V_H , Protein sequence	EVQMVESGGGLVKPGGXLKLS (Seq. ID No: 45)
	SECOND BAND
V_L , cDNA-deduced	DVLMQTQPLSLPVSLGDOASI (Seq. ID No: 46)
V_L , Protein sequence	DVLMQTQPLSLPVXXGDOASI (Seq. ID No: 47)
	THIRD BAND
V_L , cDNA-deduced	DVLMQTQPLSLPVSLGDOASI (Seq. ID No: 48)
V_L , Protein sequence	DVLMQTQPLSLPVSLGDOASI (Seq. ID No: 49)
X uncertain or alternative calls.	

A sample of anti-KC-4 chimeric antibody (approximately 190 μ g) was reduced with 5% beta-mercaptoethanol (65°C for 15 min.), separated on three lanes of a 10% SDS polyacrylamide gel, and electroblotted onto a ProBlott membrane (Applied Biosystems, Foster City, CA) in 90% 30 mM CAPS pH11, 10% methanol, for 1 hour at 25 V and at 4°C. The transferred protein species were stained with Commassie Brilliant Blue. 3 bands were seen in each lane, of which 2 migrated as expected for a heavy and light chain. The third band migrated above the light chain. Amino acid sequencing was performed directly on the immobilized bands by the Biotechnology Instrumentation Facility, University of California, Riverside. The amino acid sequence given here is the sequencer's best guess.

Example 28: Construction of anti-KC-4 Murine-Human Chimeric Antibody Genes

The vector used were described in Example 1 above. Restriction ends were added to the cDNAs in a set of PCR reactions, using primers JO20, 21, 22, and 24.

The pAG4622 light chain vector and the pAH4604 heavy chain vector were described in Example 12 above.

The new V_H and V_L DNA fragments with appropriate restriction ends were integrated into pAH4604 and pAG4622 as described in Example 12 above.

The vectors were then electroporated (together) also as described in Example 12 above.

Example 29: Tissue Binding Studies

The supernatants from stable transfectants were assayed for the presence of the anti-KC-4 murine-human chimeric antibody as described in Example 13 above. The chimeric antibody secreted in the supernatant bound both HMFG and BEM very strongly. In addition, the supernatants containing anti-KC-4 murine-human chimeric antibody were used to stain human breast carcinoma tissue sections by using the immunoperoxidase immunohistochemical staining technique. The intensity of the staining was comparable to that obtained with the original murine monoclonal antibody. The anti-KC-4 monoclonal antibody is known to bind the human milk fat globule and the breast epithelial mucin. This binding specificity of the anti-KC-4 murine monoclonal antibody was maintained even after the recombinant procedure. The anti-KC-4 chimeric antibody bound very strongly to HMFG and BEM as determined by a radioassay (Ceriani, et al., Breast Cancer Res. Trent. 15:161 (1990)). In addition, the anti-KC-4 chimeric antibody bound several human breast tumors in histopathological sections in a manner comparable to the anti-KC-4 murine monoclonal antibody, as detected by immunostaining described in Example 15 above. This specificity of binding demonstrated the retained binding reactivity of the variable regions of anti-KC-4 murine antibody by the polypeptide of the invention when attached to the human F_c fragment.

Example 30: Materials and Assays for Epitope Mapping

The specific details of the preparation of materials, cell lines, and techniques employed were disclosed by Peterson et al. (Peterson, J.A., et al., "Molecular Analysis of Epitope Heterogeneity of the Breast Mucin", Breast Epithelial Antigens, Ed. Ceriani, R.L., Plenum Press, NY (1991)), the relevant text of which is incorporated herein by reference.

Overlapping peptide hexamers were synthesized onto the ends of polyethylene pins using an Epitope Spinning Kit (Cambridge Research Biochemicals, Cambridge, UK), which is based on a method originally described by Geysen et al. (Geysen, H.L., et al., "Use of Peptide Synthesis to Probe Vital Antigens for Epitopes to a Resolution of a Single Amino Acid", PNAS (USA) 81:3998-4002 (1984)). The polyethylene pins were arranged in a 8x12 configuration that fits into a 96 well microtiter dish. The pins are supplied with an alanine attached to the ends to which the amino acids are added consecutively using pentafluorophenyl active esters of fluorophenyl-methyloxycarbonyl (Fmoc)-L-amino acids. Each consecutive overlapping hexamer differs from the previous one by a single amino acid and enough were synthesized to span the entire sequence of the peptide to be tested so that every combination of hexamer was present. Each monoclonal antibody was tested for binding to the synthetic peptides using an ELISA method with horseradish peroxidase-conjugated goat anti-murine IgG (Promega, Madison, WI) and color development with 2,2-azino-bis (3-ethylbenzothiazoline- 6-sulfonic acid (Sigma, St. Louis, MO).

The hexapeptides starting with A, P, D, and T bind well to the antibodies (Hexamers 1 to 3 and 20), whereas the hexamers starting between these positions did not. The hexamers that were prepared are shown in Table 27 below. The linear amino acid sequence essential for its binding to the antigen may be deduced from the hexamer that each monoclonal antibody binds. For example, the anti-BrE-3 antibody required the sequence TRP within the hexamer. Other monoclonal antibodies required other amino acid sequences (e.g., anti-Mc5, TRPAP; anti-Mc1, DTR; anti-BrE-1, DTRP). BrE-2 also required TRP but its different specificity for normal and tumor tissue indicates that its epitope on the native antigen is different from BrE-3.

Table 27: Epitope Mapping of Repeat Peptide Breast Mucin

Hexamer	GV TSA	PD TR P A P G S T A P P A H G V T S	A P D T R P
1		PD TR P A	
2		D TR P A P	
3		TR P A P G	
4		R P A P G S	
5		P A P G S T	
6		A P G S T A	
7		P G S T A P	
8		G S T A P P	
9		S T A P P A	
10		T A P P A H	
11		A P P A H G	
12		P P A H G V	
13		P A H G V T	
14		A H G V T S	
15		H G V T S A	
16		G V T S A P	
17		V T S A P D	
18		T S A P D T	
19		S A P D T R	
20		A P D T R P	

Example 31: Epitope Mapping

Five different monoclonal antibodies (Mc1, Mc5, BrE1, BrE2 and BrE3) were prepared using HMFG for immunization. All identified epitopes on the highly glycosylated large molecular weight breast mucin. By immunohistochemistry they appeared to recognize different epitopes since each had different tissue and tumor specificities (Peterson, J.A., et al., "Biochemical and Histological Characterization of Antigens Preferentially Expressed on the Surface and Cytoplasm of Breast Carcinomas Cells Identified by Monoclonal Antibodies Against the Human Milk Fat Globule", Hybridoma 9:221-235 (1990)). Each monoclonal antibody bound to a different spectrum of normal tissues and their specificities for different carcinomas were different. Anti-BrE2 and anti-BrE3, however, were quite similar. In addition, by screening breast *Agtl* cDNA expression libraries with some of these monoclonal antibodies, cDNA clones were isolated that produced fusion proteins that bound all of them, while other cDNA clones bound just some (Larocca, D., et al., "High Level Expression in *E. Coli* of an Alternate Reading Frame of pS2 mRNA that Encodes a Mimotope of Human Breast Epithelial Mucin Tandem Repeat" Hybridoma 11(2):191-201 (1992)). This binding to the fusion proteins indicated that the epitopes for these five monoclonal antibodies included the polypeptide portion of this glycoprotein. To confirm this the binding of these monoclonal antibodies to two synthetic polypeptide 20-mers (PDTRPAPGSTAPP AHGV TSA and APPAHGVTSAPDTRPAGST) that spanned the tandem repeat consensus sequence was tested (Gendler, S.J., et al., "Cloning of Partial cDNA Encoding

Differentiation and Tumor-Associated Mucin Glycoproteins Expressed by Human Mammary Epithelium", PNAS (USA) 84:6060-6064 (1987); Siddiqui, J., et al., "Isolation and Sequencing of a cDNA Coding for the Human DF3 Breast Carcinoma-Associated Antigen", PNAS (USA) 85:2320-2323 (1988).

5 One was started at the beginning of the published 20 amino acid repeat (Gendler, S.J., et al. (1987), *supra*) unit, and the other was started in the middle. All five monoclonal antibodies bound to both synthetic peptides, as did DF3, a monoclonal antibody against breast carcinoma cells produced by others (Hull, S.R., et al., "Oligosaccharide Differences in the DF3 Sialomucin Antigen from
10 Normal Human Milk and the BT-20 Human Breast Carcinoma Cell Line", *Cancer Comm.* 1:261-267 (1989)). Three other monoclonal antibodies against other components of the HMFG that do not cross-react with the breast mucin, Mc13, against a 70 KDa glycoprotein, and Mc3 and Mc8, against a 46 KDa glycoprotein do not bind to these synthetic peptides (data not shown) (Ceriani, R.L., et al., "Characterization of Cell Surface Antigens of Human Mammary
15 Epithelial Cells with Monoclonal Antibodies Prepared Against Human Milk Fat Globule", *Somat. Cell Genet.* 9:416-427 (1982); Peterson, J.A., et al., "Biochemical and Histological Characterization of Antigens Preferentially Expressed on the Surface and Cytoplasm of Breast Carcinoma Cells Identified by
20 Monoclonal Antibodies Against the Human Milk Fat Globule", *Hybridoma* 9:221-235 (1990)).

Example 32: Approach for Humanization of Antibodies

The present humanization approach is based on Padlan, E.A., "Choosing the Best Framework to Use in the Humanization of an Antibody by CDR-Grafting:
25 Suggestions from 3-D Structural Data", *Antibody Engineering* 2nd. Annual Conf. San Diego, CA (Dec 16-17, 1991).

The fine specificity may be preserved in a "humanized" antibody only if the CDR structures, their interaction with each other, and their interaction with the rest of the variable domains can be maintained. (Padlan, E.A. (1991), *supra*).
30 This requires the preservation of residues of the FR amino acids which contact the CDRs, those which are involved in the V_L - V_H contact, and those which are buried and could influence the overall domain structure and the structure of the combining site.

By examination of murine Fab structures, for which atomic coordinates are available, the FR amino acids that are probably "important" in maintaining the
35 structure of the combining site may be determined (Padlan, E.A., 8th International Congress of Immunol., Budapest, Hungary, Abstracts p. 19 (August 2-28, 1992)).

The specificity of an antibody depends on the CDR structures and sometimes, on some of its neighboring residues as well. These CDR structures, in turn, depend on contacts with framework amino acids and on the interaction of the V_L and V_H domains. Thus, to ensure the retention of binding affinity, not only the CDR residues must be preserved, but also those FR residues that contact either the CDRs or their opposite domains, as well as all buried residues, which give shape to the variable domains. The buried amino acids are placed in exactly the same positions in human and in murine frameworks (Padlan, E.A., "A Possible Procedure for Reducing the Immunogenicity of Antibody Variable Domains While Preserving Their Ligand-Binding Properties", *Molecular Immunology* 28:489-498 (1991)).

This approach was applied to design humanized analogues of the variable regions of the murine antibodies of the invention. The humanization or design of the exemplary analogue peptide provided herein was undertaken as follows. The identification of the residues, which are most probably "important" in preserving the combining site structure, permits the selection of the best human FR sequences to use in the "humanization" of each chimeric antibody of known structure or analogues peptides of the invention. The results of the analysis can be used also to predict which FR amino acids should probably be retained in those cases where no three-dimensional structural data are available.

The present procedure was designed to reduce the immunogenicity of the xenogeneic antibodies by preparation of their chimeric derivatives or fragments thereof while preserving their antigen-binding properties. In general, the antigen binding properties of an antibody are primarily determined by its CDRs. The CDRs of the murine antibody were "grafted" herein onto a human framework. In addition, the FR amino acids in the antibody, that are judged as probably important in maintaining the combining-site structure, may also be retained in the humanized molecule.

Example 33: Choice of a Murine Model of Known Structure for the Humanization of the BrE-3 Antibody

The classification of the V_H and V_L domains of an antibody such as the anti-BrE-3 antibody was done according to Kabat, E.A., et al. (Kabat, E.A., et al., "Sequences of Proteins of Immunological Interest" NIH (1991)). The anti-BrE-3 kappa chain V_L domain belongs to group II and the V_H domain belongs to group IIIc. A murine antibody was then found, whose structure had been determined, and whose variable regions belong to the same classes. The anti-fluorescein murine antibody 4-4-20 shown in Table 1 above (Herron, J.N., et al., "Three-Dimensional Structure of a Fluorescein-Fab Complex Crystallized in 2-Methyl-2,4-

Pentanediol", Proteins, 5:1-280 (1989) fits these requirements since, like BrE-3, it has V_L and V_H domains belonging to groups II and III C. Thus, the three-dimensional structures of antibodies BrE-3 and 4-4-20 should be similar, and BrE-3 may be modeled after 4-4-20.

5 **Example 34: Choice of Target Human Framework for Humanization of BrE-3 Antibody**

10 The choice of the target human framework was not based on the similarity of the amino acid sequence of the entire framework, but strictly on the similarity at the residues that were judged to be structurally important according to the 4-4-20 model. That is, only amino acids that could be involved in contacts with CDR of the opposite chain, or amino acids whose side-chains were predicted to be inwardly pointed. The positions of these amino acids are shown in Tables 8 and 9 and also in Tables 2, 3, 4, 5 and 6 above. These positions are as follows.

15 For the light chain variable region framework: 1, 2, 3, 4, 5, 6, 11, 13, 19, 21, 23, 35, 36, 37, 38, 44, 45, 46, 47, 48, 49, 58, 60, 61, 62, 69, 71, 73, 75, 78, 82, 86, 88, 98, 102, and 104.
20 For the heavy chain variable region framework: 4, 6, 12, 18, 20, 22, 24, 27, 28, 29, 30, 36, 37, 38, 39, 40, 45, 46, 47, 48, 49, 66, 67, 68, 69, 71, 73, 76, 78, 80, 82c, 86, 88, 90, 91, 92, 93, 94, 103, 107, 109 and 111.

25 The numbering system is conventionally accepted (Kabat, et al. (1991), supra) and shown in Tables 10 and 11 above. In this case, the consensus sequences of all human F_v regions were selected as the target human framework to minimize the immunogenicity of the product.

30 First, the sequences of the murine variable chains were aligned with consensus sequences from all known human variable region classes (Herron, J.N., (1989), supra) and the number of differences in the amino-acids that must be retained from the murine were scored. The positions of these amino acids were obtained from those of murine monoclonal antibody 4-4-20, which was chosen to model the anti-BrE-3 antibody as shown in Tables 28 and 29 below.

Table 28: Choice of BrE-3 V_L Target Human Framework

BrE-3 V _L	
CDR1	CDR2
.....
BRE3K	DVVMTOTPLSLPVS LGDQASISC RSSQNLVHN-NGNTYLY WFLQKSGQSPKLLTY RASIRFS
HuKi-a	.IQ...S...SA.V...RVT.T...A...S...XSS-ISN...A...YQ...P.KA.....A...SLE.
HuKi-ai	.I....S.....TP.EP.....S.L.SXD.....N...Y...P...Q....LV.N.A.
HuKi-ii	EI.L...S.GT.SL.P.ER.TL...A...SVSSS...—A...YQ...P...A.R....G...S.A.T
HuKi-iv	.I....S.D...A.....ER.T.N.K...SVLYSS.NKN...A...YQ...P...P.....W.T.E.
CDR3	
BRE3K	GVPDRFSGSGSETDFTLKISRVEAEDLGVYFC FQGTHVPW-T FGGGTKLEIK
HuKi-a	...S.....G.....T...SLQP...FAT.Y. Q.YNSL.EW...Q...V...
HuKi-aiG.....V...Y. M.ALQX.RX...Q...V...
HuKi-ii	.I.....G.....T...LP...FA...Y. Q.YGSS.PL...Q...V...
HuKi-ivG.....T...SLQ...VA...Y. Q.YYST-X...Q...V...

. identity with the murine sequence

* the murine residues that are structurally important.

Table 29: Choice of BrE-3 V_H Target Human Framework

BrE-3 V _H	
CDR1	CDR2
.....
BRE1VH	EVKLEESGGGLVQPGGSMKLSCAASGFTFS DAWMD- WVRQSPEKGLEWVA
HuHI	Q.Q.VQ...AEVKK...A.V.V...K...Y...T SYAIS- ...A.GQ...MG
HuHII	Q.Q.Q...P...K.SQTL.S.T.TV...GSV. SYXWSWN .I.P.G.....IG
HuHIII	..Q.V.....LR..... SYAS- ...A.G.....S
BRE1VH	EIRNKANNHATYYDESVKG RFTISRDDSKSRVYLQMISLR AEDTGLYYCTG
HuHI	W.N-PYG.GD.N.AQKPK...V...TA.T.T.TA.MELS...S...AV...AR
HuHII	R.YYR.YSGS.X.NP.L.S...V...V.T...NQPS.KLS.VT.A...AV...AR
HuHIII	V.SG.TDGG...AD... ..N.NTL...N.....AV...AR
CDR3	
BRE1VH	EF—AN WGQGTLVTVSA
HuHI	APGYGSGGGCYRGDYXFDYS
HuHII	ELPGGYXGDDYYXXGFDVS
HuHIII	GRXGXSLSGXYYYHYFDYS

. identity with the murine sequence

* the murine residues that are structurally important.

Based on these scores, the human frameworks belonging to groups V_LII and V_HIII were chosen to receive the anti-BrE-3 CDRs plus other important amino acids.

Example 35: Identification of BrE-3 Murine-Human Differences

5 The original murine sequences (BrE-3 V_L or V_H) were aligned with their closest human (Human KII or HIII) relatives that were chosen after comparing their sequences in Table 28 above. The alignment of these two sequences is shown in Table 30 below. The information in this table is also contained in Table

28 above, but is reproduced here for clarity. The CDRs are not shown, since their sequences were not changed during the humanization process. Thus, Table 30 shows the maximum number of amino acids that can be changed toward the humanization of the anti-BrE-3 antibody, based on the consensus human sequences obtained from the current databases (Kabat, E.A., et al. (1991), supra). If all these positions were to be replaced with the corresponding human amino acids, the corresponding CDR grafted antibody variable regions would be attained.

Table 30: Corresponding Amino Acid Sequences of VK BrE-3 and Human KII

	CDR1	CDR2
BrE-3 VK	DVVMTQTPLSLPVSLGDQASISC	WFLQKSGQSPKLLTY
Human KII	.I...S.....TP.EP....	.Y...P...Q....
	CDR3	
BrE-3 VK	GVPDRFSGSGSETDFTLKISRVEAEDLGVYFC	FGGGTKLEIK
Human KIIG.....V...Y.Q...V...	
	CDR1	
BrE-3 VII	EVKLEESGGGLVOPGGSMKLSCAASGFTFS	WVRQSPEKGLEWVA
Human III	..Q.V.....LR.....	...A.G.....S
	CDR2	
BrE-3 VII	RFTISRDDSKSRVYLQMISLR	AEDTGLYYCTG
Human IIIN...NTL....N.....AV...AR	
	CDR3	
BrE-3 VII	WGQGTLVTVSA	
Human IIIS	

Table 31 and 32 below contain the same information as Table 30 above in a different format. They show the numbers of the residues that would have to be changed in order to completely convert the original murine framework completely into a human consensus framework.

Table 31: BrE-3 V_L Amino Acid Candidates for Change into Human Consensus Sequences

V _L			
VI2	FY36	EG68	GQ100
TS7	SP40	LV83	LV104
ST14	KQ45	FY87	LP15
DE17			
QP18			

Table 32: BrE-3 V_H Amino Acid Candidates for Change into Human Consensus Sequences

	V _H		
	KQ3	SA40	DN73
5	AS113	EV5	EG42
	SN76	ML18	AS49
	RT77	KR19	VL78
	IN82a	GA88	LV89
			TA93
10			GR94

Example 36: Identification of Important Murine BrE-3 Amino Acids

The "important" murine amino acids that should be preserved were chosen based on the contacts of a particular amino acid with the CDRs, and with the opposite chains and/or whether their side chains are pointing inwardly or outwardly. The positions of these "important" amino acids were determined based on the examination of the known structures of other antibodies. This information is provided in Tables 33 and 34 below.

Table 33: Important V_L Amino Acid Positions to be Preserved

20	V _L		
	V2	CDR contact	Buried
	F36	CDR contact	Contact with VH
	K45	CDR contact	
25	F87	Possible contact with VH	
	G100	Possible contact with VH	
	L104	Buried	

Table 34: Important V_H Amino Acid Positions to be Preserved

30	V _H		
	M18	Buried	
	S40	Buried	
	A49	CDR contact	Buried
35	D73	CDR contact	
	S76		Buried
	V78	CDR contact	Buried
	L89	Might affect interaction with V _L	
40	T93	CDR contact	
	G94	CDR contact	Buried

Most of the "important" amino acids were selected on the basis of the structure of antibody 4-4-20 and according to Tables 2, 3, 4, 5, 6, 8 and 9 above. Two important amino acids out of each chain, however, were selected based on more general structural analysis, using other antibody structures. This was done to maximize the chances of conserving ligand binding properties. In particular the preservation of the Leu at 89-H was suggested in order to ensure the maintenance of, the $V_L:V_H$ contact. Although the residue at 89-H is usually not in contact with V_L and is only partly buried, it nonetheless contributes to the interface. A Val for Leu replacement at this position could very well create a "cavity" which could affect the contact. An Ile for Leu replacement would probably be fine since these amino acids have essentially the same side chain volume.

Finally, by comparing the position of all amino acids that are candidates for mutation, shown in Tables 31 and 32 above, with those that are "important" and should be preserved, shown in Tables 33 and 34, the final selection of amino acid positions for actual mutation was attained. Any "important" amino acid position was eliminated from the list of candidates. Table 35 below shows the amino acids that were selected for change from murine to human identities to obtain the present humanized analogue.

Table 35: Selected Amino Acids for Mutation

	V_L	V_H
	TS7	KQ3
25	ST14	EV5
	LP15	KR19
	DE17	EG42
	QP18	RT77
	SP40	IN82a
30	EG68	GA88
	LV83	AS113

Example 37: Introduction of Changes in BrE-3 Amino Acid Sequence

The changes were done at the DNA level in sequential manner. All but one of the codon mutations were performed using enzymatic inverse PCR (EIPCR), a mutagenesis technique developed by Stemmer and Morris (Stemmer, W.P.C and Morris, S.K., "Enzymatic Inverse Polymerase Chain Reaction: a Restriction Site Independent, Single Fragment Method for High Efficiency Site-Directed Mutagenesis", BioTechniques 13:146-220 (1992)).

First, the entire plasmid, containing the target cDNA was amplified by inverse PCR using terminal mutagenic oligonucleotides. Second, BsaI was used to cut the ends of the incorporated primers. This enzyme cuts at a site that is displaced from its recognition site. Thus, after digestion of the open amplified plasmid with BsaI, the BsaI recognition sequence was removed from the ends of the DNA. The DNA was left with complementary sticky ends and could be closed into a functional plasmid that contains the mutagenized region. The amino acid and DNA sequences of the non-mutated (wild-type) variable light and heavy chains of the anti-BrE-3 antibody are shown in Tables 15 and 16 above. The amino acid sequences of the anti-BrE-3 antibody frameworks and the mutations that were performed for the humanization, the oligonucleotide that was used for the mutagenesis, and the method of mutagenesis are shown in Tables 36 to 44 below.

Table 36: F_V FR1 Mutation Sites

Position	FR1	Analogue 1 (8 Changes)	DNA Codon (FR1) (Analogue)	Method for Mutagenesis	Primers
Leader Peptide Not Shown					
	D				
	V				
	V				
	M				
	T				
7	Q	S	ACT TCT	EIPCR*	(J037,J038)
	T				
	P				
	L				
	S				
	L				
	P				
	V				
14	S	T	AGT ACT	EIPCR	(J037,J038)
15	L	P	CTT CCT	EIPCR	(J037,J038)
	G				
17	D	E	GAT GAG	EIPCR	(J037,J038)
18	Q	P	CAA CCA	EIPCR	(J037,J038)
	A				
	S				
	I				
	S				
	C				

* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 37: F_{VL} FR2 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
CDR1 Not Shown					
40	W F L Q K S G Q S P K L L I Y	P	TCA CCA	EIPCR*	(JO39,JO40)

* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 38: F_{VL} FR3 Mutation Sites

Position	FR3	Analogue 1 (8 Changes)	DNA Codon (FR3) (Analogue)	Method for Mutagenesis	Primers
CDR2 Not Shown					
68	G V P D R F S G S G S E T D F T L K I S R V E A E D L G V Y F C	G	GAG GGG	EIPCR*	(JO41,JO42)
83		V	CTG GTG	EIPCR	(JO41,JO42)

* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 39: F_{VL} FR4 Mutation Sites

Position	FR4	Analogue 1 (10 Changes)	Analogue 2 (6 Changes)	DNA Codon (FR4) (Analogue)	Method for Mutagenesis	Primers
CDR3 Not Shown						
	F		F			
	G		G			
	G		G			
	G		G			
	T		T			
	K		K			
	L		L			
	E		E			
	I		I			
	K		K			

Table 40: F_{VH} FR1 Mutation Sites

Position	FR1	Analogue 1 (8 Changes)	DNA Codon (FR1) (Analogue)	Method for Mutagenesis	Primers
Leader Peptide Not Shown					
	E				
	V				
3	K	Q	AAG CAG	EIPCR*	(J057,J058)
5	L	V	GAG GTG	EIPCR	(J057,J058)
	E				
	S				
	G				
	G				
	L				
	V				
	Q				
	P				
	G				
	S				
19	M	R	AAA AGA	EIPCR	(J057,J058)
	K				
	L				
	S				
	C				
	A				
	A				
	S				
	G				
	F				
	T				
	F				
	S				

* EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 41: F_{VM} FR2 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
CDR1 Not Shown					
42	W V R S P E K G L E W V A	Q	G	GAG GGG	EIPCR [*] (JO55,JO56)

^{*} EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 42: F_{VM} FR3 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
CDR2 Not Shown					
77	R F T I S R D D S K S R V Y L Q M	T	AGA ACT	EIPCR [*]	(JO53,JO54)
82	I S L R A E D	N	ATA AAT	EIPCR	(JO53,JO54)
88	T G L Y C T G	A	GGC GCC	EIPCR	(JO53,JO54)

^{*} EIPCR: Enzymatic Inverse Polymerase Chain Reaction

Table 43: F_W FR4 Mutation Sites

Position	FR2	Analogue 1 (8 Changes)	DNA Codon (FR2) (Analogue)	Method for Mutagenesis	Primers
CDR3 Not Shown					
	W				
	G				
	Q				
	G				
	T				
	L				
	V				
	T				
	V				
	S				
113	A	S	GCA TCT	PCR*	JO51, JO52)*

*PCR: Polymerase Chain Reaction

Table 44: Sequences of Mutagenic Oligonucleotides

JO37	TCC CTG GGT CTC ACT CCT GGA GAG CCA GCT TCC ATC TCT TGC AGA TCT AGT (Seq. ID No. 50)
JO38	AGC TTG GGT CTC AGG AGT GAC AGG CAG GGA GAG TGG AGA TTG GGT CAT CAC AAC (Seq. ID No. 51)
JO39	G TTC CTG GTC TCG CCA GGC CAG TCT CCA AAG CTC CTG (Seq. ID No. 52)
JO40	T TGG AGG TCT CCC TGG CTT CTG CAG GAA CCA ATA TAA AT (Seq. ID No. 53)
JO41	TTC ACA GGT CTC ATC AGC AGA GTG GAG GCT GAG GAT GTG GGA GTT TAT TT (Seq. ID No. 54)
JO42	AGC CTC GGT CTC GCT GAT CTT GAG TGT GAA ATC TGT CCC TGA TCC ACT GC (Seq. ID No. 55)
JO51	CCT GGA GGA TCC ATG AGA CTC TCT TGT GCT (Seq. ID No. 56)
JO52	GTT GGG GCT AGC AGA AGA GAC AGT GAC CAG AGT (Seq. ID No. 57)
JO53	TAC CTG GGT CTC AAT AGC TTA AGA GCT GAA GAC ACT GCC CTT TAT TAC TGT (Seq. ID No. 58)
JO54	TTC AGC GGT CTC GCT ATT CAT TTG CAG GTA CAC AGT ACT TTT GGA ATC ATC (Seq. ID No. 59)
JO55	GTC CGC GGT CTC CCA GGG AAG GGG CTT GAG TGG GTT GCT GAA ATT AGA AA (Seq. ID No. 60)
JO56	CTC AAG GGT CTC CCC TGG AGA CTG GCG GAC CCA GTC CAT CCA GGC ATC A (Seq. ID No. 61)
JO57	T GAG GAG GTC TCA GGA GGC TTG GTG CAA CCT GGA GGA TCC ATG AGA CTC TCT (Seq. ID No. 62)
JO58	GG TTG CGG TCT CCC TCC TCC AGA CTC CAC AAG CTG CAC TTC ACT CTG GAC A (Seq. ID No. 63)

* note: Primer set JO51 and JO52 was originally intended for mutagenizing K19 to R (by JO51) and A113 to S (by JO52), as described below. Primer JO51, however, was somehow defective (it did not run as a single band on a polyacrylamide gel) and thus only the A113 to S mutagenesis was successful. Mutation K19 to R was accomplished at a later time with EIPCR, using primers JO57 and JO58.

Example 38: Synthesis of Primers

All primers except JO51 and JO52 were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City CA). Primers JO51 and JO52 were purchased from Keystone Laboratories, Inc. Menlo Park, CA.

Both, the PCR method used with JO51 and JO52 and the EIPCR method used with all other primer sets, are described below.

the plasmid DNA template was extracted with a kit purchased from QIAGEN (Tchapsworth, CA) and diluted to 1 ng/ μ l in 10 mM TRIS 1 mM EDTA pH 7.5 - 8. This plasmid is composed of vector pCR1000 (Invitrogen Corporation, San Diego, CA) into which the cDNA encoding the variable region to be humanized was inserted.

A mixture of PCR primers was made where each primer was present at a concentration of 10 pmole/ μ l in water.

The PCR amplification conditions were as follows. All reagents as well as the GeneAmp PCR system 9600 were purchased from Perkin Elmer Cetus. Optimal PCR conditions were determined empirically for each pair of mutagenic primers. A matrix of conditions varying the concentration of $MgCl_2$, mutagenic primers, and template plasmid DNA was set up. The annealing and extension temperatures during PCR may be varied.

Plasmid template (500 pg/ μ l), 0.5 μ M each mutagenic oligonucleotide, 1 mM $MgCl_2$, 10 mM TRIS pH 8.3, 50 mM KCl, 0.2 mM each nucleotide triphosphate (dGTP, dATP, TTP, dCTP), and Taq polymerase 1 unit/ 20 μ l reaction mixture.

Example 39: Hot Start PCR

All the components of the PCR mixture, with the exception of Taq polymerase, were mixed in a 95 μ l volume. The mixture was then dispensed in 19 μ l aliquots into 5 PCR tubes. The reason for performing five independent reactions was to decrease the odds that unwanted mutations be isolated as a result of nucleotide misincorporation during PCR. The tubes were heated to 95°C for 5 minutes and then cooled to 72°C. While at that temperature 1 μ l of an appropriate Taq polymerase dilution in 10 mM TRIS, pH 8.3, 50 mM KCl was added to the reaction tubes. The temperature cycling then proceeded as follows.

94°C, 3 min

[(94°C, 1 min) (50°C, 1 min) (72°C, 3.5 min)] 3 cycles

[(94°C, 1 min) (55°C, 1 min) (72°C, 3.5 min)] 25 cycles

72°C, 10 min

Example 40: Extra Final Extension

After cycling, the contents of the five tubes were mixed and an extra final extension reaction was carried out. Extra nucleotide triphosphates were added (to 125 μ M), 5 units of Taq polymerase were also added and the mixture was heated to 72°C for 10 minutes.

Example 41: Purification of PCR Products

The PCR products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5 μ g/ml Ethidium Bromide. The correct DNA band was visualized with UV light (366 nm), excised from the gel and extracted with the GeneClean kit purchased from Bio 101, La Jolla, CA.

Example 42: Restriction Digestion

The DNA was then digested with BsaI for two hours at 60 degrees celsius in 25 μ l (20.5 μ l of DNA, 2.5 μ l 10 X buffer 4 (NEB) and 2 μ l BsaI (NEB). BsaI sites were designed near the 5' end of the PCR primers. The primers included 6 extra nucleotides 5' of the BsaI sites to facilitate digestion by BsaI. There were no BsaI sites elsewhere in the plasmid. Other restriction enzymes may be used as advised by Stemmer and Morris (Stemmer and Morris (1992), supra). This special class of restriction enzyme cuts at a site that is different from its recognition site but, nevertheless, at a precise distance from it. Using this method, there was no need for having restriction sites in the sequence in order to perform the mutagenesis.

Example 43: Second Purification

The restricted products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5 μ g/ml Ethidium Bromide. The correct DNA band was visualized with UV light (366 nm), excised from the gel and extracted with the GeneClean kit purchased from Bio 101, La Jolla, CA.

Example 44: Ligation (Reclosure of Plasmid)

The ligation mixtures consisted of 5 μ l extracted DNA, 2 μ l 10x ligation buffer (NEB) 1 μ l T4 DNA polymerase (NEB), 12 μ l water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation was carried out at room temperature for 2 hrs., or alternatively at 14°C overnight.

Example 45: Transformation and Sequencing

The reclosed plasmids were then transformed into E. coli. We used Inv alpha F' competent cells purchased from Invitrogen Corporation, San Diego CA.

Plasmid DNA was then prepared from a few transformants and sequenced to verify that mutagenesis was successful.

Example 46: Mutagenesis of BrE-3 Antibody using JO51 and JO52

These oligonucleotides were designed to mutagenize K19 to R (by JO51)

and A113 to S (by JO52), not by using EIPCR but using the normal PCR (with primers pointing to each other). Primer JO51 carried a BamHI site and primer JO52 carried an NheI site. After mutagenic PCR amplification the resulting amplified DNA cassette was inserted into the plasmid in lieu of the corresponding wild-type DNA fragment. There was no compelling reason to use this method over the EIPCR method except that conveniently placed restriction sites (BamHI and NheI) were available. This method, however, yielded only the A113 to S mutation. A subsequent analysis showed that the JO51 primer, which carried the K19 to R mutation ran aberrantly on a polyacrylamide gel.

The protocol for the mutagenic amplification step was as follows. Plasmid template (500 pg/ μ l), 0.75 μ M each mutagenic oligonucleotide, 2 mM $MgCl_2$, 10 mM TRIS pH 8.3, 50 mM KCl, 0.2 mM each nucleotide triphosphate (dGTP, dATP, TTP, dCTP), and Taq polymerase 1 unit/20 μ l reaction mixture..

The PCR was not started as described for EIPCR above. The temperature cycling conditions used were as follows.

94°C, 3 min

[(94°C, 1 min) (44°C, 1 min) (72°C, 1 min)] 3 cycles

[(94°C, 5 sec) (65°C, 1 min) (72°C, 1 min)] 32 cycles

72°C, 10 min

The extra final extension and purification of PCR products were conducted as described for EIPCR above.

The restriction digestion of the vector and the insert were conducted as follows. The PCR product was digested with BamHI and NheI for 1 hr. 50 min. at 37°C (19 μ l of DNA, 2.5 μ l 10Xbuffer 3 (NEB), 1.5 μ l BamHI (NEB), 1.5 μ l NheI (NEB). The vector, which is the starting plasmid described above, was digested under similar conditions (1 μ l plasmid (1 μ g), 2 μ l 10Xbuffer 3 (NEB), 1.5 μ l BamHI (NEB), 1.5 μ l NheI (NEB) 14 μ l water, 2 1/2 hrs. at 37°C).

The restricted products, vector and insert, were then purified once more as described above for EIPCR.

The ligation of the fragments was conducted as follows. The ligation mixtures consisted of 5 μ l vector, 5 μ l insert, 2 μ l 10x ligation buffer (NEB) 1 μ l T4 DNA polymerase (NEB), 7 μ l water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation is carried out at 14°C overnight. A control ligation with vector only was carried out in parallel. The transformation of the host cells was conducted as described for EIPCR.

Example 47: Plasmid Preparation and Sequencing of Humanized BrE-3 V_H

Plasmid DNA was then prepared from several independent transformants. A few of the plasmids that were shown by restriction analysis to contain the insert were then sequenced to verify that the mutagenesis was successful. When one of the sequenced DNAs contained the desired mutation it was utilized for the next mutation cycle. The fully mutated humanized analogue BrE-3 DNA sequences for the V_H and V_L segments are shown in Tables 45 and 46 below.

Table 45: BrE-3 Antibody V_L Humanized Analogues DNA Sequences
BrE-3 V_L FR-HZ

ATG AAG TTC CCT GTT AAG CTG TTG GTG CTG TTG TTC TGG ATT CCT GCT TCC ATC AGT GAT GTT GTG
ATG ACC CAA TCT CCA CTC TCC CTG CCT GTC ACT GCT GGA GAG CCA GCT TCC ATC TCT TGC AGA TCT
AGT CAG AAC GTT GTA CAC AAC AAT GGA AAC ACC TAT TTA TAT TGG TTC CTG CAG AAG CCA GGC CAG
TCT CCA AAG CTC CTG ATT TAT AAG GCT TCC ATC CGA TTT TCT GGG GTC CCA GAC AAG TTC AGT GGC
AGT GGA TCA GGC ACA GAT TTC ACA CTC AAG ATC AGC AGA GTG GAG GCT GAG GAT GTG GGA GTT TAT
TTC TGC TTT CAA GGT ACA CAT GTT CCG TGG ACC TTC GGT GGA GGC ACC AAG CTG GAA ATC AAA C
(Seq. ID No. 64)

Table 46: BrE-3 Antibody V_H Humanized Analogue DNA Sequences

BrE-3 V_H FR-HZ
ATG TAC TTG GGA CTG AAC TAT GTC TTC ATA GTT TTT CTC TTA AAA GGT GTC CAG AGT GAA GTG GAG
CTT GTG GAG TCT GGA GGA GGC TTG GTG CAA CCT GGA GGA TCC ATG AGA CTC TCT TGT GCT GCT TCT
GGA TTC ACT TTT AGT GAT GGC TGG ATG GAC TGG GTC GGC CAG TCT CCA GGC AAG GGC CTT GAG TGG
GTT GCT GAA ATT AGA AAC AAA GGC AAT AAT CAT GCA ACA TAT TAT GAT GAG TCT GTG AAA GGC AAG
TTC ACC ATC TCA AGA GAT GAT TCC AAA AGT ACT GTG TAC CTC CAA ATG AAT AGC TTA AGA GCT GAA
GAC ACT GGC CTT TAT TAC TGT ACT GGG GAG TTT GCT AAC TGG GGC CAG GGC ACT CTG GTC ACT GTC
TCT TCT (Seq. ID No.: 65)

Example 48: Humanized BrE-3 Antibody Expression

Two expression vectors pAG4622 and pAH4604 (Coloma, M.J., et al. (1992), supra) were used that were developed and provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). Any cDNA encoding a signal peptide and either the variable heavy chain or the variable light chain can, in principle, be inserted into these vectors resulting in a construction that encodes an IgG1, K, antibody with human constant regions. Correctly modified cDNAs were excised from pCR1000 with EcoRV and Sal I and inserted into pAG4622. These encode the modified light chain. The wild-type heavy chain

was similarly excised from pCR1000 by digestion with EcoRV and NheI and inserted into pAH4604. The restriction and ligation reactions necessary to accomplish these operations were performed under the conditions stipulated by the enzyme manufacturers (New England Biolabs, Beverly, MA). Both the vectors and the inserts were purified from an agarose gel prior to ligation, using the Bio101 (La Jolla, CA) GeneClean kit (glass beads). The V_H and V_L regions in the final constructions were sequenced once again to verify that they are correct. The non-producer myeloma cell line SP2/O-Ag14, ATCC: CRL 1581, (Shulman M., et al. (1978), supra) was transfected with both plasmid constructions, and antibody producers were isolated following the recommendations outlined in (Coloma, M.J. et al. (1992), supra) except that selection was done only for the uptake of hisD (by adding 5 mM histidinol to the medium and readjusting the pH to 7.4 with NaOH). Usually after ten days, stable transfectant colonies were established at a frequency of approximately 10⁻⁶ to 10⁻⁴. Colonies were then transferred to normal medium (without histidinol). The culture media were either Dulbecco's modified Eagle's medium (DME): fetal bovine serum (FBS), 90:10, v/v, or a mixture of DME:RPMI:FBS, 45:45:10, v/v/v. Penicillin and streptomycin were added to prevent bacterial growth.

The supernatants from stable transfectants were assayed for the presence of the antibodies. This was done by capturing the secreted chimeric antibody with a plate-bound goat anti-human-kappa chain antibody and developing with goat anti-human-gamma chain antibody, essentially as described previously (Coloma, M.J. (1992), supra) except that the secondary antibody was radiolabeled with ¹²⁵I. The supernatants were also assayed for binding to human milk fat globule (HMFG) as described previously (Ceriani R.L., et al., "Diagnostic Ability of Different Human Milk Fat Globule Antigens in Breast Cancer", Breast Cancer Res. Treat., 15:161-174 (1990)). HMFG is bound to the microtiter plates as described previously (Ceriani, R.I. (1984), supra). Usually most colony supernatants were positive by both assays. Colonies that secrete the highest level of antibody in the supernatants, as determined by these assays, were subcloned and subsequently adapted to serum-free medium for the purification of antibody. Serum free medium contains HL-1 supplement as directed by the manufacturer (Ventrex Labs., Portland, ME).

35 Example 49: Half Humanized-Half Chimeric BrE-3 Antibody

A BrE-3 humanized light chain was paired with an anti-BrE-3 non-humanized chimeric heavy chain by co-transfection of SP2/O myeloma cells with hybrid plasmids carrying the respective DNA sequences and those of a human

F.

Example 50: Determination of Affinity Constants for Half Humanized and Fully Humanized BrE-3 Antibodies

The secreted half humanized-half chimeric and fully humanized antibodies were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), supra). Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), supra) using successive layers of methylated BSA, glutaraldehyde, anti- β -galactosidase and the bacterial fusion protein 11-2 (a hybrid of β -galactosidase and human mammary mucin). Each well contained 388 ng of the 11-2 fusion protein. To each well were added 25 μ l 125 I-BrE-3 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % sodium azide pH7.4, in phosphate buffer saline) and compete with 25 μ l of either unlabeled murine or chimeric antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM. Iodinations were performed with 125 I (17Ci/mg, Nordion International). Fifty micrograms of monoclonal antibody BrE-3 (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), supra)

Antibody-antigen affinity constants were determined by taking the reciprocal of the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. (1987), supra). The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to antigen against the same radiolabeled antibody. Both, the half humanized-half chimeric antibody and the fully humanized antibody competed about as well or better with the anti-BrE-3 murine antibody for the antigen.

Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

Example 51: Histochemical Specificity of Half and Fully Humanized BrE-3 Antibodies

Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain useful affinity for the carcinoma antigens. Breast carcinoma tissue sections were stained with the supernatant of the half humanized/half chimeric and fully humanized transfected cells using

the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies showed strong staining patterns.

Example 52: Gel Chromatography of Half Humanized-Half Chimeric and Fully Humanized BrE-3 Antibodies

Antibody disulfide bonds were reduced by heating for 10 min at 65°C in Laemmli loading buffer containing 5% beta-mercaptoethanol. The separated chains were then chromatographed on a SDS polyacrylamide gel (10%). Two bands were observed for both antibodies of similar migration pattern as the murine antibody. These data were also confirmed by Western blotting.

Example 53: Deduced Amino Acid Sequences of Humanized BrE-3 Variable Light and Heavy Chains

The amino acid sequences of the light and heavy chains of the analogue humanized antibody are shown in Tables 47 and 48 below. These amino acid sequences may be improved either to increase affinity for the antigen or to decrease immunogenicity in humans. Numerous variants of this sequence may be engineered in accordance with the invention.

Table 47: Humanized BrE-3 V_L Analogue Amino Acid Sequence
BrE-3 V_L FR-HZ

20	Leader	mklpvrlilFWIPASIS (Seq. ID No: 66)
	FR1	DVVMTQSPLSLPVTGPGEASISC (Seq. ID No: 67)
	CDR1	RSSQNLVHNNNGNTLY (Seq. ID No: 68)
	FR2	WFLOKPGQSPKLLIY (Seq. ID No: 69)
	CDR2	RASIRFS (Seq. ID No: 70)
25	FR3	GVPDRFSGSGSGTDFLTKISRVEAEDVG VYFC (Seq. ID No: 71)
	CDR3	FGTHVPWT (Seq. ID No: 72)
	FR4	FGGGTKLEIK (Seq. ID No: 73)

Table 48: BrE-3 V_H Analogue Amino Acid Sequence
BrE-3 V_H FR-HZ

30	Leader	mylglnyvtlVFLKGVQS (Seq. ID No: 74)
	FR1	EVQLVESGGGLVOPGGSMRLSCAASGFTFS (Seq. ID No: 75)
35	CDR1	DAWMD (Seq. ID No: 76)
	FR2	WVRQSPGKGLEWVA (Seq. ID No: 77)
	CDR2	EIRNKANNHATYYDESYKG (Seq. ID No: 78)
	FR3	RFTISRDDSKSTVY LQMNSLRAEDTALYYCTG (Seq. ID No: 79)
	CDR3	EFAN (Seq. ID No: 80)
40	FR4	WGQGLTVTVSS (Seq. ID No: 81)

**Example 54: Choice of Murine Model of Known Structure
for Humanization of anti-KC-4 Antibody**

The classification of the V_H and V_L domains of an antibody such as the anti-KC-4 antibody was done according to Kabat et al. (Kabat, E.A., et al., "Sequences of Proteins of Immunological Interest" NIH (1991)). The KC-4G3 kappa chain V_L domain belongs to group II and the V_H domain belongs to group IIId. A murine antibody was then found, whose structure had been determined, and whose variable regions belong to the same classes. The anti-myohemerythrin peptide antibody B1312 fits these requirements since, like the anti-KC-4 murine antibody, it has V_L and V_H domains belonging to groups II and IIId (Stanfield, R.L., et al., "Crystal Structures of an Antibody to a Peptide and its complex with Peptide Antigen at 2.8 Å", Science 248:712-719 (1990)). Thus, the three-dimensional structures of antibodies the anti-KC-4 and B1312 antibodies should be similar, and the humanization of the anti-KC-4 antibody may be modeled after B1312.

**Example 55: Choice of Target Human Framework for
Humanization of Chimeric anti-KC-4 Antibody**

The choice of the target human framework was based strictly on the similarity at the residues that were judged to be structurally important according to the B1312 model. That is, only amino acids that could be involved in contacts with CDRs of the opposite chain, or amino acids whose side-chains were predicted to be inwardly pointed. The positions of these amino acids are shown in Tables 49 below.

Table 49: Important Amino Acid Positions for anti-KC-4 Antibody

Light Chain Variable Region Framework

2, 3, 4, 6, 7, 11, 13, 19, 21, 22, 23, 35, 36, 37, 38, 43, 44, 46, 47, 48, 49, 58, 60, 61, 62, 69, 71, 73, 75, 78, 82, 85, 86, 87, 88, 98, 102, 104 and 106.

Heavy Chain Variable Region Framework

2, 4, 6, 12, 18, 20, 22, 24, 27, 28, 29, 36, 37, 38, 39, 43, 45, 46, 47, 48, 49, 66, 67, 69, 71, 78, 80, 82, 82c, 86, 88, 90, 91, 92, 93, 94, 103, 107, 109 and 111.

The numbering system is conventionally accepted (Kabat, et al. (1991), supra) and is shown in Tables 10 and 11 above. In this case, the consensus sequences of all human F_v regions were selected as the target human framework to minimize the immunogenicity of the product.

First, the sequences of the murine variable chains were aligned with consensus sequences from all known human variable region classes (Herron, J.N., (1989), supra) and the number of differences in the amino-acids that must be retained from the murine were scored. The positions of these amino acids were obtained from those of the B1312 murine monoclonal antibody, which was chosen to model the humanization of the anti-KC-4 antibody.

Based on these scores, the consensus sequences human frameworks belonging to groups V_HII and V_HIII were chosen to receive the anti-KC-4 murine antibody CDRs plus other important amino acids.

10 **Example 56:** Identification of Murine-Human
anti-KC-4 Antibody Differences

The original murine sequences (anti-KC-4 V_L or V_H) were aligned with their closest human (Human KII or HIII) relatives that were chosen after comparing their sequences as described in Example 34 above. In the present example, it was intended to be substituted as many amino acids as possible in going from the murine to the humanized variable consensus sequences, leaving the important amino acids intact as described in Examples 55 and 57. The amino acids chosen to be preserved were a subset of those listed above. These were selected by analogy to the B1312 sequence. The single exception was the glycine (100) residue of the original framework of the variable region of the murine kappa chain, which was retained despite not being encompassed in Table 49 above since it was thought that it might contact the variable domain of the heavy chain. Such contacts were observed in at least three Fab that lack a gly at this position.

25 **Example 57:** Identification of Important Murine
anti-K-4 Antibody Amino Acids

The "important" murine amino acids were chosen for preservation based on the contacts of a particular amino acid with the CDRs, and with the opposite chains and/or whether their side chains are pointing inwardly or outwardly. The positions of these "important" amino acids were determined based on the examination of the known structures of other antibodies.

Most of the "important" amino acids were selected on the basis of the structure of antibody B1312 and according to Tables 2, 3, 4, 5, 6, 8 and 9 above. The final selection of amino acid positions for actual mutation was attained by comparing the position of all amino acids that are candidates for mutation with those that are "important" and should be preserved. Any "important" amino acid position was eliminated from the list of candidates. Table 50 below shows the amino acids that were selected for change in the

murine sequence to attain the humanized sequence in the present exemplary analogue.

**Table 50: Anti-KC-4 Murine Antibody Variable Region
Amino Acids Selected for Mutation**

Position	KC-4G3 Murine Identity	→	Consensus Human Identity
Light Chain Variable Region			
10	14 15 16 17 18 19 20 21 22 23	→	14 15 16 17 18 19 20 21 22 23
Heavy Chain Variable Region			
15	13 14 15 16 17 18 19 20 21 22 23 24 25	→	13 14 15 16 17 18 19 20 21 22 23 24 25

The change N → K at position 74 in the variable light chain knowingly eliminated an N-linked glycosylation site, which was present in the original murine monoclonal antibody.

**Example 58: Introduction of Changes in Amino Acid Sequence
for Humanization of anti-KC-4 Antibody**

The introduction of the changes in the amino acid sequence was not done as described in Example 37 above. Instead the DNA encoding each humanized variable region was synthesized in a single polymerase chain reaction (PCR) using overlapping oligonucleotides in accordance with the method described by Ye et al. (Ye, Q-Z, Johnson, L.L., and Baragi, V., "Gene Synthesis and Expression in *E. coli* for PUMP, a Human Matrix Metalloproteinase", BBRC 186(1):143-149 (1992)). The sequences of the mutagenic are shown in Table below.

**Table 51: Primers for Humanization of anti-KC-4
Murine Antibody Variable Regions**

	J059	CCCCGATCC TTTAAAGGT GTCCAGTGTG AAGTCAGAT GGTGGAG TGTG (SEQ. ID No.: 82)
5	J060	GAATTCGGGGC TACCACTAGA GACAGTGACC AGAGTCCCTT GGGCCGAG (SEQ. ID No.: 83)
	J061	AGTCGAGATG GTGCACTGTG GGGGAGGCTT AGTCGAGCCT GGAGGGTCCC TGAGACTCTC
		CTGTGCAACC TGTGCACTCC CTTTCAGTAG CTATGCCATG T (SEQ. ID No.: 84)
	J062	CTTGATAGTA GCGTAATTA CCACCACTAC TAATTCTGC GACCACTCC AGCCCTCTCC
		CTGGAGCCTG GCGAAGCTCA GACATGGCAT AGCTACTGAA A (SEQ. ID. No.: 85)
10	J063	TAATTAGGCC TACTATCAAG ACACTGTGAC GGGCGAATC ACCATCTCCA GAGACAATTC
		CAAGAACACC CTGTACTTGC AAATGAACAG TGTGAGGCT G (SEQ. ID. No.: 86)
	J064	CCAGACTCCC TTGGCCGCGAG TAAGCAAAACC AGGCGGGAT ACCGTAGTCC TCCCTTGCAC
		AGTAATACAC GGGCTGTCTC TCAGCCCTCA CACTCTTCAT T (SEQ. ID. No.: 87)
	J073	GGGAAGCTTG ATATCCACCA TGAAGTTGCC TGTAGGCTG TTGGTCTGTA TTTCTGGAT
15		TCCTGC (SEQ. ID No.: 88)
	J074	AAAGATTCC TCGACTTACG TTTTATTTCC AGCTTGGTCC CCGCTCC GAA CCGTAAGGA
		ACATGT (SEQ. ID. No.: 89)
	J075	CTGATTTCT GGAATCTCTC TTCAGCAAT GATGTTTGA TGACCC AAAC TCTCTCTCC
		CTGGCTGTCA CTCAGGAGA GGCAGCTTCC ATCTCTTCA (SEQ. ID. No.: 90)
20	J076	CTGTGAGAC TGGCTGCTT TCTGCAGTA CCATCTAAA TAGGTG TTTC CATTACTATG
		TACAATGCTC TGACTAGATC TCGAAGAGAT CGAGGCTGCC (SEQ. ID. No.: 91)
	J078	CGAAGCTGA CGAACATGT GAGCTTGAA AGCAATAATA AATTCC CACA TCTTCAGGCT
		CCACTCTGCT GATCTTGAAT GTCAAAATCG TCTCTGATCC (SEQ. ID. No.: 92)

**25 Example 59: Synthesis of Primers for Humanization
of anti-KC-4 Antibody**

All primers were synthesized on a PCR-Mate EP DNA synthesizer model 391 (Applied Biosystems, Foster City CA) using 40nmole columns, cycle 1:63, with Trityl off. None were purified before use. The EIPCR method was used
30 for preparing all primer sets. Their sequences are shown in Table 51 above.

The plasmid DNA template was extracted with a kit purchased from QIAGEN (Tchapsworth, CA) and diluted to 1ng/ μ l in 10mM TRIS 1mM EDTA pH 7.5 - 8. This plasmid is composed of vector pCR1000 (Invitrogen Corporation, San Diego, CA) into which the cDNA encoding the variable region to be
35 humanized was inserted.

**Example 60: Synthesis of anti-KC-4 Humanized
Heavy Chain Variable Regions**

A mixture of PCR primers was made, where each primer was present at a concentration of 10 pmole/ μ l in water.

40 Four 101 'mer oligonucleotides (J061, J062, J063 and J064), one 50' mer (J059), and one 49' mer (J060), were used for the synthesis of the humanized variable heavy chain. The oligonucleotides concentrations were

estimated using the formula

$$c = [(A_{260})/30] \mu\text{g}/\mu\text{l}$$

The PCR amplification conditions were as follows. All reagents as well as the GeneAmp PCR system 9600 were purchased from Perkin Elmer Cetus.

- 5 Optimal PCR conditions were determined empirically for each pair of mutagenic primers. A matrix of conditions varying the concentration of MgCl_2 , mutagenic primers, and template plasmid DNA were set up as follows. However, the annealing and extension temperatures during PCR may be varied.

- 10 2 μM primer JO59 150nM each of primers JO61, 62, 63 and 64.
 2 μM primer JO60 200 μM each of dGTP, dATP, TTP, and dCTP.
 10mM KCl 20mM Tris-HCl pH 8.8
 10mM $(\text{NH}_4)_2\text{SO}_4$ 2 units per 100 μl reaction Vent DNA
 0.1% Triton X-100 polymerase (New England Biolabs)
 6mM MgSO_4

15 **Example 61: Hot Start PCR for Humanization of anti-KC-4 Antibody**

- All the components of the PCR mixture, with the exception of Vent DNA polymerase, were mixed. The mixture was then dispensed in 19 μl aliquots into 5 PCR tubes. The reason for performing five independent reactions was to decrease the odds that unwanted mutations be isolated as a result of nucleotide misincorporation during PCR. The tubes were heated to 95°C for 5 minutes and then cooled to 72°C. While at that temperature 1 μl of an appropriate Vent DNA polymerase dilution in 1 x buffer was added to the reaction mixture (hot start). The temperature cycling then proceeds as follows.

- 25 [(96°C, 6 sec) (55°C, 10 sec) (72°C, 30 sec)] 3 cycles
 [(96°C, 5 sec) (60°C, 10 sec) (72°C, 30 sec)] 29 cycles
 72°C, 10 min

Example 62: Extra Final Extension for Humanized anti-KC-4 Antibody DNA

- After cycling, one extra final extension reaction was carried out. Extra deoxyribonucleotide triphosphates (to 125 μM) and 1 unit of Vent DNA polymerase were added, and the mixture was heated to 72°C for 10 minutes.

The resulting synthetic DNA fragment was digested with DraI and NheI and inserted into the same restriction sites a plasmid construct encoding the corresponding murine heavy chain variable region.

35 **Example 63: Synthesis of anti-KC-4 Humanized Light Chain Variable Regions**

The light chain variable region (V_L) genes were synthesized in a similar way as described in Examples 60 to 62 above for the heavy chain variable regions.

Example 64: Purification of Humanized anti-KC-4 PCR Products

The PCR products were then separated on a 0.8% agarose gel in 1XTAE buffer and 0.5 µg/ml ethidium bromide. The correct DNA band were visualized with UV light (366 nm), excised from the gels and extracted with the GeneClean kit (Bio 101, La Jolla, CA).

Example 65: Ligation of Humanized anti-KC-4 DNA to Plasmid (Reclosure of Plasmid)

The ligation mixtures consisted of 5 µl extracted DNA, 2 µl 10x ligation buffer (NEB) 1 µl T4 DNA polymerase (NEB), 12 µl water. The amount of plasmid DNA may be varied depending of the intensity of the band extracted from the Gel. Ligation into a pBluescript KS⁺ plasmid (Stratagene) was carried out at room temperature for 2 hrs., or alternatively at 14°C overnight.

Example 66: Transformation and Sequencing of Humanized anti-KC-4 DNA

The reclosed plasmids were then transformed into E. coli utilizing Inv alpha F' competent cells purchased from Invitrogen Corporation, San Diego CA. Plasmid DNA was then prepared from a few transformants and sequenced to verify that mutagenesis was successful.

Example 67: Hybrid Plasmid Preparation and Sequencing

Plasmid DNA was then prepared and sequenced to verify that the mutagenesis was successful. The mutated anti-KC-4 humanized analogue DNA sequences for the V_H and V_L segments are shown in Tables 52 and 53 below.

Table 52: Humanized anti-KC-4 Antibody V_L Analogue DNA Sequences

		anti-KC-4 V _L FR-HZ																			
		ATG	AAG	TTG	CCT	GTT	AGG	CTG	TTG	GTG	CTG	ATG	TTC	TGG	ATT	CCT	GCT				
		TCC	AGC	AGT	GAT	GTT	TTG	ATG	ACC	CAA	ACT	CCT	CTC	TCC	CTG	CCT	GTC				
30		ACT	CCA	GGA	GAG	CCA	GCC	TCC	ATC	TCT	TGC	AGA	TCT	AGT	CAG	AGC	ATT				
		GTA	CAT	AGT	AAT	GGA	AAC	ACC	TAT	TTA	GAA	TGG	TAC	CTG	CAG	AAA	CCA				
		GGC	CAG	TCT	CCA	CAG	CTC	CTG	ATC	TAC	AAA	GTT	TCC	ATC	CGA	TTT	TCT				
		GGG	GTC	CCA	GAC	AGG	TTC	AGT	GGC	AGT	GGA	TCA	GGG	ACA	GAT	TTC	ACA				
		CTC	AAG	ATC	AGC	AGA	GTG	GAG	GCT	GAG	GAT	GTG	GGA	ATT	TAT	TAC	TGC				
35		TTT	CAA	GGT	TCA	CAT	GTT	CCG	TAC	ACG	TTC	GGA	GGG	GGG	ACC	AAG	CTG				
		GAA	ATA	AAA	C (SEQ. ID. No.: 93)																

Table 53: Humanized anti-KC-4 Antibody V_H
Analogue DNA Sequences

		anti-KC-4 V _H FR-H2																			
		ATG	GAC	TTT	GGG	CTC	AGC	TTC	GTT	TTC	CTT	GTC	CTT	ATT	TTA	AAA	GGT				
5		GTC	CAG	TGT	GAA	GTG	CAG	ATG	GTG	GAG	TCT	GGG	GGA	GGC	TTA	GTG	CAG				
		CCT	GGA	GGG	TCC	CTG	AGA	CTC	TCC	TGT	GCA	GCC	TCT	GGC	TTT	GCT	TTC				
		AGT	AGC	TAT	GCC	ATG	TCT	TGG	GTT	CGC	CAG	GCT	CCA	GGG	AAG	GGG	CTG				
		GAG	TGG	GTC	GCA	GAA	ATT	AGT	AGT	GGT	GGT	AAT	TAC	GCC	TAT	CAA					
		GAC	ACT	GTG	ACG	GSC	CGA	TTC	ACC	ATC	TCC	AGA	GAC	AAT	TCC	AAG	AAC				
10		ACC	CTG	TAC	CTG	CAA	ATG	AAC	AGT	CTG	AGG	GCT	GAG	GAC	ACG	GCC	GTG				
		TAT	TAC	TGT	GCA	AGG	GAG	GAC	TAC	GGT	ATC	CCG	GCC	TGG	TTT	GCT	TAC				
		TGG	GGC	CAA	GGG	ACT	CTG	GTC	ACT	GTC	TCT	AGT	(SEQ. ID. No.: 94)								

Example 68: Expression of Anti-KC-4 Humanized Antibody

Two expression vectors pAG4622 and pAH4604 (Coloma, M.J., et al. (1992), *supra*) were used that were developed and provided by S.L. Morrison (Dept. of Microbiology and Molecular Genetics, UCLA). Any cDNA encoding a signal peptide and either the variable heavy chain or the variable light chain can, in principle, be inserted into these vectors resulting in a construction that encodes an IgG1, K, antibody with human constant regions. Correctly modified cDNAs were excised from pCR1000 with EcoRV and Sal I and inserted into pAG4622. These encode the modified light chain. The wild-type heavy chain was similarly excised from pCR1000 by digestion with EcoRV and NheI and inserted into pAH4604. The restriction and ligation reactions necessary to accomplish these operations were performed under the conditions stipulated by the enzyme manufacturers (New England Biolabs, Beverly, MA). Both the vectors and the inserts were purified from an agarose gel prior to ligation, using the Bio101 (La Jolla, CA) GeneClean kit (glass beads). The V_H and V_L regions in the final constructions were sequenced once again to verify that they were correct. The non-producer myeloma cell line SP2/O-Ag14, ATCC: CRL 1581, (Shulman M., et al. (1978), *supra*) was transfected with both plasmid constructions, and antibody producers were isolated following the recommendations outlined in Coloma et al. (Coloma, M.J. et al. (1992), *supra*) except that selection was done only for the uptake of hisD (by adding 5 mM histidinol to the medium and readjusting the pH to 7.4 with NaOH). Usually after ten days, stable transfectant colonies were established at a frequency of approximately 10⁻³ to 10⁻⁴. Colonies were then transferred to normal medium (without histidinol). The culture media were either Dulbecco's modified Eagle's medium (DME): fetal bovine serum (FBS), 90:10, v/v, or a mixture of DME:RPMI:FBS, 45:45:10, v/v/v. Penicillin and streptomycin were added to prevent bacterial growth.

The supernatants from stable transfectants were assayed for the presence of the antibodies. This was done by capturing the secreted chimeric

antibody with a plate-bound goat anti-human-kappa chain antibody and developing with goat anti-human-gamma chain antibody, essentially as described previously (Coloma, M.J. (1992), *supra*) except that the secondary antibody was radiolabeled with ^{125}I . The supernatants were also assayed for binding to human milk fat globule (HMFG) as described previously (Ceriani R.L., et al., "Diagnostic Ability of Different Human Milk Fat Globule Antigens in Breast Cancer", *Breast Cancer Res. Treat.*, 15:161-174 (1990)). HMFG is bound to the microtiter plates as described previously (Ceriani, R.L. (1984), *supra*). Usually most colony supernatants were positive by both assays. Colonies that secrete the highest level of antibody in the supernatants, as determined by these assays, were subcloned and subsequently adapted to serum-free medium for the purification of antibody. Serum free medium contains HL-1 supplement as directed by the manufacturer (Ventrex Labs., Portland, ME).

Example 69: Half Humanized-Half Chimeric anti-KC-4 Antibody

An anti-KC-4 humanized light chain was paired with an anti-KC-4 non-humanized chimeric heavy chain by co-transfection of SP2/O-Ag14 myeloma cells with hybrid plasmids carrying the respective DNA sequences and those of a human F_c . The cell line obtained, HuKC-4V1 was deposited with the ATCC on September 23, 1993 and awarded an accession number HB 11454.

In addition, an anti-KC-4 humanized heavy chain was paired with an anti-KC-4 non-humanized chimeric light chain as described in Example 69 above. The thus obtained cell, HuKC-4V3 was deposited with the ATCC on September 23, 1993 and awarded an accession number HB 11456.

Example 70: Fully Humanized anti-KC-4 Antibody

An anti-KC-4 fully humanized antibody was prepared by pairing fully humanized anti-KC-4 light and heavy chains by co-transfection as described in Example 69 above. The cell line, HuKC-4V2, was deposited with the ATCC on September 23, 1993 and awarded the accession number HB 11455.

Example 71: Determination of Affinity Constants for Half and Fully Humanized anti-KC-4 Antibodies

The secreted half humanized antibodies and the fully humanized antibody were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), *supra*). Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), *supra*) using successive layers of

methyated BSA, glutaraldehyde, anti- β -galactosidase and the bacterial fusion protein 11-2 (a hybrid of β -galactosidase and human mammary mucin). Each well contained 388 ng of the 11-2 fusion protein. To each well were added 25 μ l 125 I-KC-4 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % sodium azide pH 7.4, in phosphate buffer saline) and compete with 25 μ l of either unlabeled murine or chimeric antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM). Iodinations were performed with 125 I (17 Ci/mg, Nordion International). 50 μ g anti-KC-4 monoclonal antibody (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), supra).

The antibody-antigen affinity constants were determined by taking the reciprocal of the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. (1987), supra). The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to the antigen against the same radiolabeled antibody. The fully humanized antibody was shown to compete about as well or better with the anti-KC-4 murine antibody for the KC-4G3 antigen.

Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

Example 72: Histochemical Specificity of Half and Fully Humanized anti-KC-4 Antibodies
Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain the affinity for the KC-4G3 carcinoma antigen of the murine antibody. Breast carcinoma tissue sections were stained with the supernatant of the KC-4 murine and fully humanized transfected cells using the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies showed strong staining patterns.

The following Table 54 shows the results of the immunoperoxidase staining of five human breast carcinomas with either the standard anti-KC-4G3 murine or the fully humanized antibodies. Both stained the same tissues at a comparable level.

Table 54: immunoperoxidase Staining of Human Breast Carcinoma Tissue Sections with Murine and Fully Humanized anti-KC-4 Antibodies

Breast Tumor	Murine Antibody	Fully Humanized Antibody
5 1	++	++
2	+++	+++
3	-	-
4	++	++
5	+++	+++

- 10 **Example 73:** Binding to HMFG of Half Humanized and Fully Humanized anti-KC-4 Antibodies
Tissue culture supernatants from transfectants of all three anti-KC-4 variants of the humanized antibody were shown to bind the human milk fat globule (HMFG) as determined by radio-immunodetections.

- 15 **Example 74:** Half Humanized and Fully Humanized anti-KC-4 Antibodies Bind to Goat anti-Human κ or γ Antibodies

Tissue culture supernatants from transfectants of all three variants of the anti-KC-4 humanized antibody were shown to bind in sandwich
20 radioimmunodetections to both goat anti-human kappa chain antibody bound to microtiterplate wells (750ng/well), and to radio-iodinated 125 I-labeled goat anti-human gamma chain antibodies.

The results of these sandwich assays demonstrate that both chains of the humanized antibodies indeed possess human kappa and gamma constant
25 regions.

- Example 75:** Deduced Amino Acid Sequences of Humanized anti-KC-4 Variable Light and Heavy Chains

The amino acid sequences of the light and heavy chains of the analogue humanized antibody are shown in Tables 55 and 56 below. The actual amino
30 acid sequences may be varied either to increase affinity for the antigen or to decrease immunogenicity in humans. Numerous variants of this sequence may be engineered in accordance with the invention.

Table 55: Humanized anti-KC-4 Antibody V_L Analogue Sequenceanti-KC-4 V_L FR-HZ

MKLPVRLVL MFWIPASSSD VLMTQTPLSL PVTGPGEASI SCRSSQSIVH
 SNGNTYLEWY LQKPGQSPQL LIYKVSIRFS GVPDRFSGSG SGTDFTLKIS
 5 RVEAEDVGII YCFQGSHVPY TFGGGTKLEI K (Seq. ID No: 95)

Table 56: Humanized anti-KC-4 Antibody V_H Analogue Sequenceanti-KC-4 V_H FR-HZ

MDFGLSLVFL VLILKGVCCE VQMVESGGGL VQPGGSLRLS CAASGFAFSS
 10 YAMSWVROAP GKGLEWVAEI SSGGNYAYYQ DTVTGRFTIS RDNSKNTLYL
 QMNSLRAEDT AVYYCAREDY GIPAWFAYWG QGLTVTVSS (Seq. ID No: 96)

Example 76: Half Humanized-Half Chimeric anti-BrE-3 Antibodies

A humanized BrE-3 light chain was paired with a non-humanized chimeric
 15 BrE-3 heavy chain by co-transfection of SP2/O-Ag14 myeloma cells with hybrid
 plasmids carrying the respective DNA sequences and those of a human F_c . The
 cell line obtained, HuBrE3V1, was deposited with the ATCC on November 10,
 1993 and awarded an accession number HB _____.

In addition, a humanized BrE-3 heavy chain was paired with a non-
 20 humanized chimeric BrE-3 light chain as described in Example 69 above. The
 thus obtained cell, HuBrE3V3 was deposited with the ATCC on November 10,
 1993 and awarded an accession number HB _____.

Example 77: Fully Humanized BrE-3 Antibody

The fully humanized BrE-3 antibody was prepared by pairing fully humanized
 25 BrE-3 light and heavy chains by co-transfection as described in Example 69
 above. The thus obtained cell line, HuBrE3V2, was deposited with the ATCC on
 November 13, 1992 and awarded the accession number HB 1111200.

Example 78: Half Humanized and Fully Humanized
 BrE-3 Antibodies Bind to Goat
 30 anti-Human α or γ Antibodies

Tissue culture supernatants from transfectants of all three variants of the
 humanized BrE-3 antibody were shown to bind in sandwich
 radioimmunoassays to both goat anti-human kappa chain antibody bound to

microtiterplate wells (750ng/well), and to radio-iodinated ^{125}I -labeled goat anti-human gamma chain antibodies.

The results of these sandwich assays demonstrate that both chains of the humanized antibodies indeed possess human kappa and gamma constant regions.

Example 79: Binding to HMFG of Half Humanized and Fully Humanized BrE-3 Antibodies

Tissue culture supernatants from transfectants of all three BrE-3 variants of the humanized antibody were shown to bind the human milk fat globule (HMFG) as determined by radio-immunodetections.

Example 80: Competition Assays and Determination of Affinity Constants for Half Humanized BrE-3 Antibodies

The secreted half humanized antibodies were purified from culture supernatants using a Sepharose 4B-protein A column (Bio-Rad, Richmond, CA.) as described by Ey et al. (Ey, P.L., et al. (1978), supra). The fully humanized antibody (HuBrE3 V2) was not purified. Its concentration in the culture supernatant was determined by radioimmunodetection using a plate-bound goat anti-human kappa chain antibody as a capturing antibody and using radiolabeled goat anti-human gamma chain antibody as a detecting antibody. A parallel standard curve of human IgG, κ was used to determine the unknown concentration of HuBrE3 V2. Microtiter plates (Dynatech, Chantilly, VA) were prepared as described by Ceriani et al. (Ceriani, R.L., et al. (1992), supra) using successive layers of methylated BSA, glutaraldehyde, anti- β -galactosidase and the bacterial fusion protein 11-2 (a hybrid of β -galactosidase and human mammary mucin). Each well contained 388 nanograms of the 11-2 fusion protein. To each well were added 25 μl ^{125}I -BrE-3 in RIA buffer (10 % bovine calf serum, 0.3% triton X-100, 0.05 % sodium azide pH 7.4, in phosphate buffer saline) and competed with 25 μl of either unlabeled murine or humanized antibody in RIA buffer at the final concentrations of 130 pM, 850 pM, 1.3 nM, 4 nM, and 13 nM. Iodinations were performed with ^{125}I (17 Ci/mg, Nordion International). 50 μg BrE-3 monoclonal antibody (Coulter, Hialeah, FL) were labeled at a specific activity of 9.56 mCi/mg using the chloramine T method as described previously by Ceriani et al. (Ceriani, R.L., et al. (1988), supra).

All humanized versions of the BrE-3 antibody were shown to compete about as well or better with the labeled murine BrE-3 antibody for the antigen as did the murine BrE3 antibody. The protocol used to determine affinity constants was as described above except that in each case, an unlabeled antibody competed for binding to the antigen against the same radiolabeled antibody. The antibody-antigen affinity constants were determined by taking the reciprocal of

the concentration of competing unlabeled monoclonal antibody that produced 50 % binding as described by Sheldon et al. (Sheldon, K., et al. (1987), supra).

Polyacrylamide gel electrophoresis was performed to insure that the antibody chains migrated as expected. The affinity binding constants of the murine, chimeric, half humanized and humanized antibodies were determined in independent competition assays.

Example 81: Histochemical Specificity of Half Humanized BrE-3 Antibodies

Immunohistochemical staining using the immunoperoxidase technique of consecutive human breast carcinoma tissue sections was used as a test to verify that the analogue antibodies retain the affinity for the carcinoma antigen of the murine BrE-3 antibody. Breast carcinoma tissue sections were stained with the supernatant of cells transfected with the HuBrE3 V1 humanized antibody using the Vectastain ABC method (Vector Labs, Burlingame, CA). Both antibodies showed strong staining patterns.

The following Table 57 shows the results of the immunoperoxidase staining of five human breast carcinomas with either the standard BrE-3 murine or the half humanized HuBrE3 V1 antibody. Both stained the same tissues at a comparable level.

Table 57: Immunoperoxidase Staining of Human Breast Carcinoma Tissue Sections with Murine and Half Humanized BrE-3 Antibodies

Breast Tumor	Murine Antibody	Half Humanized Antibodies
1	++	++
2	+++	+++
3	-	-
4	++	++
5	+++	+++

Example 82: Hybridoma Cell Deposits

The following cell lines were deposited as present examples of the best mode of the invention. The hybridoma cell lines expressing the murine-human fully chimeric BrE-3 and anti-KC4 antibodies and the fully humanized BrE-3 antibody were deposited with the ATCC on November 13, 1992 under the Budapest Treaty, and have been assigned Accession Nos. HB 11199 (Chimeric BrE-3 A1C10), HB 11201 (Chimeric anti-KC-4 1E8) and HB 11200 (Humanized BrE-3 A1C10). The hybridoma cell line expressing the anti-KC-4 humanized

antibody was deposited with the ATCC on September 23, 1993 and has been assigned Accession No. HB 11455 (Humanized HuKC-4 V2). The half chimeric/ half humanized antibody cell lines deposited with the ATCC are as follows: ATCC No. HB 11454 (light humanized/ heavy chimeric chains, HuKC-4V1) and ATCC No. HB 11456 (Heavy humanized/ light chimeric chains, HuKC-4V3) were deposited on September 23, 1993, and ATCC No. HB _____ (light humanized/ heavy chimeric chains, BrE3V1) and ATCC No. HB _____ (heavy humanized/ light chimeric chains, BrE3V3) were deposited on November 10, 1993 under the Budapest Treaty as examples of the best mode of the invention known to the inventors.

The invention now being fully described, it will be apparent to one of ordinary skill in the art that many changes and modifications can be made thereto without departing from the spirit or scope of the invention as set forth herein.

**WHAT IS CLAIMED AS NOVEL AND UNOBVIOUS AND
DESIRED TO BE PATENTED IN LETTERS PATENT IS:**

1. A pure, isolated analogue peptide which selectively binds to an antigen that is present on the surface or in the cytoplasm of a neoplastic cell or that is released by the cell, the peptide being selected from the group consisting essentially of at least one CDR of the light or heavy chain of an antibody of a first species having affinity and specificity for the human mammary fat globule (HMFG) antigen and an antigen found on the surface or the cytoplasm of a neoplastic cell or that is released by the cell; at least one variable region of the light or heavy chains of an antibody of the first species having affinity and specificity for the HMFG antigen and an antigen found on the surface or the cytoplasm of a neoplastic cell or that is released by the cell, wherein about 1 to at least 46 amino acids in the FRs are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of a species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids; combinations thereof, wherein each analogue peptide is operatively linked to at least one other variable region peptide or analogue thereof; and mixtures thereof.

2. The anti-neoplastic analogue peptide of claim 1, wherein the antibody of the first species is selected from the group consisting of murine, rat, goat, rabbit, canine, primate, guinea pig, bovine, equine, feline and porcine antibodies.

3. The anti-neoplastic analogue peptide of claim 1, being able to compete with the antibody secreted by the hybridoma cell ATCC No. HB 10028 for the human mammary fat globule antigen.

4. The antineoplastic analogue peptide of claim 1, having an amino acid sequence selected from the group consisting of amino acid Sequence ID Nos: 67 through 73; amino acid Sequence ID Nos: 75 through 81; fragments thereof comprising at least one CDR or one CDR and two flanking regions thereof per chain; combinations thereof, wherein

each analogue peptide is operatively linked to at least one other analogue peptide; and mixtures thereof.

5. A glycosylated analogue peptide selectively binding to an antigen present on the surface or in the cytoplasm of a neoplastic cell or that is released by the cell, comprising the analogue peptide of claim 1; and at least one glycosyl residue operatively linked to the peptide.

6. A composition of matter, comprising the analogue peptide of claim 1; and a carrier.

7. A diagnostic kit for the in vivo therapy of neoplasms, comprising the composition of claim 6 in pharmaceutically-acceptable form; and instructions for its use.

8. The kit of claim 7, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

9. The kit of claim 7, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, DNA monomers, RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers or copolymers.

10. An in vitro carcinoma diagnostic kit, comprising the composition of claim 6; a solid support having operatively linked thereto an antigen which selectively binds to the analogue peptide; and instructions for its use.

11. The kit of claim 10, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

12. The diagnostic kit of claim 10, further comprising anti-constant region immunoglobulins, protein G, protein A, other antibody binding molecules or binding fragments thereof.

13. A hybrid analogue polymer, comprising at least one analogue peptide of claim 1 and at least one effector agent operatively linked to the

peptide; combinations thereof; and mixtures thereof.

14. The hybrid analogue polymer of claim 13, wherein the effector agent is selected from the group consisting of monomers, and non-peptide and peptide polymers other than the constant region of an antibody of the same species.

15. The hybrid analogue polymer of claim 13, wherein the effector agent comprises a radioisotope, an enzyme, or a phosphorescent or a fluorescent label.

16. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, phosphorescent and fluorescent agents, enzymes, DNA and RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers or copolymers.

17. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a non-peptide polymer selected from the group consisting of ester, ether, vinyl, amido, imido, alkylene, arylalkylene, cyanate, urethane, and isoprene polymers, halogenated polymers, DNA and RNA polymers, copolymers thereof, and copolymers thereof with peptide polymers or monomers.

18. The hybrid analogue polymer of claim 14, wherein the effector agent comprises a peptide polymer selected from the group consisting of the constant region of antibodies or fragments thereof, the CDRs or variable regions of antibodies, whole antibodies, Fab and Fab', (Fab')₂, or antibody fragments of a species other than the first species, analogues thereof, hormones, enzymes, peptide transmitters, combinations thereof, and combination thereof with non-peptide polymers, copolymers or monomers.

19. The hybrid analogue polymer of claim 18, wherein the analogue peptide comprises amino acid sequences of non-human origin; and the peptide polymer comprises at least one constant region of the light or heavy chains of a human antibody or fragments thereof capable

of binding to anti-constant region immunoglobulins, protein G or protein A or other antibody binding molecules

20. The hybrid analogue polymer of claim 19, being selected from the group consisting of an analogue peptide comprising two heavy and two light chains, each light and heavy chain comprising at least one non-human CDR or analogue variable region peptide and at least one human constant region or fragment thereof; at least one human-non-human analogue Fab, Fab', or (Fab')₂ fragment thereof; fragments thereof; combinations thereof; and mixtures thereof.

21. The hybrid analogue polymer of claim 20, wherein each pair of the analogue heavy and light chains has a different predetermined specificity.

22. The hybrid analogue polymer of claim 13, wherein the analogue peptide and the effector agent are operatively linked by a polymer.

23. The hybrid analogue polymer of claim 13, being selected from the group consisting of at least one CDR or analogue variable region of the heavy chain of a first antibody of the first species or fragment thereof comprising at least one CDR and two flanking regions thereof and a first effector agent; at least one CDR or analogue variable region of the light chain of a second antibody of the first species or fragments thereof comprising at least one CDR and two flanking regions thereof, and a second effector agent, wherein each chain has a predetermined specificity; combinations thereof; and mixtures thereof.

24. The hybrid analogue polymer of claim 13, wherein at least one first pair of light and heavy chains comprising at least one CDR or analogue variable region or fragments thereof is linked to at least one second pair of the analogue light and heavy chains comprising at least one CDR or one analogue variable region or fragments thereof.

25. An anti-carcinoma composition, comprising the hybrid analogue polymer of claim 13; and a carrier.

26. An in vivo carcinoma therapy and diagnosis kit, comprising the anti-carcinoma composition of claim 25 in pharmaceutically-

acceptable form; and instructions for its use.

27. The kit of claim 26, further comprising a molecule capable of selectively binding the analogue peptide and comprising an effector agent operatively linked thereto.

28. The kit of claim 27, wherein the effector agent comprises a molecule selected from the group consisting of therapeutic, immunogenic and diagnostic agents, radioisotopes, DNA monomers, RNA monomers, chemical linkers, transmitter molecules, combinations thereof, and combinations thereof with peptide and non-peptide polymers and copolymers.

29. A diagnostic kit for carcinomas, comprising the anti-carcinoma composition of claim 25; a solid support having operatively linked thereto an antigen which specifically binds to the hybrid analogue peptide; and instructions for its use.

30. The kit of claim 29, further comprising a molecule capable of selectively binding to the hybrid analogue polymer, the molecule having an effector agent operatively linked thereto.

31. The diagnostic kit of claim 29, further comprising anti-constant region immunoglobulins, protein G, protein A, other antibody binding molecules or binding fragments thereof.

32. A method of determining the presence of neoplastic cells in a tissue, comprising contacting a tissue suspected of comprising neoplastic cells with the anti-neoplastic analogue peptide of claim 1 and allowing the analogue peptide to bind to antigen associated with any neoplastic cells present in the tissue to form an analogue peptide-cell antigen complex; and detecting the presence of any complexes formed.

33. An in vitro method of determining the presence of neoplastic cells in a tissue comprising the method of claim 32, wherein the tissue is excised from a subject prior to being contacted with the analogue peptide in vitro.

34. An in vitro method of diagnosing a neoplasm comprising the method of claim 33, and comparing the results with a standard cut-off

value determined by comparing results obtained for normal and neoplastic samples, wherein a result above the cut-off value is diagnostic of a neoplasm.

35. The in vitro method of diagnosing a neoplasm of claim 34, conducted by contacting a biological sample obtained from a subject suspected of having a neoplasm with a known amount of the anti-tumor analogue peptide of claim 1 in the presence of a solid supported antigen molecule that is selectively bound by the analogue peptide and allowing the formation of solid supported analogue peptide-antigen molecule complexes and analogue peptide-sample antigen complexes with any neoplastic cell antigen present in the sample; detecting any complex formed between the analogue peptide and the solid supported neoplastic antigen; and comparing the result obtained with the standard cut-off value.

36. An in vivo method of imaging a neoplasm present in a subject, comprising the method of claim 32, wherein a pharmaceutically-acceptable composition comprising an effective amount of the analogue peptide in radiolabeled form to reach the neoplasm and bind thereto is administered to a subject suspected of having a primary or metastasized neoplasm; and the detection is of any binding of the labeled analogue peptide to the antigen on the surface or in the cytoplasm of the neoplastic cells.

37. An in vivo method of diagnosing the presence of a neoplasm in a subject comprising the method of claim 36, and comparing the results with a standard cut-off value determined by comparing results obtained for normal and neoplastic samples, wherein a result above the cut-off value is diagnostic of a neoplasm.

38. A method of inhibiting the growth or reducing the size of a primary or metastasized neoplasm in a subject comprising administering to a subject in need of the treatment an effective amount of the anti-tumor composition of claim 25 in pharmaceutically-acceptable form, wherein the effector agent comprises a therapeutic agent.

39. The method of claim 38, further comprising administering to

the subject a molecule capable of selectively binding to the analogue peptide and having an effector agent operatively linked thereto.

40. A method of purging neoplastic cells from a biological fluid, comprising the method of claim 32; and separating any analogue peptide-cell complexes present from the remainder of the fluid.

41. An ex vivo method of purging neoplastic cells from a biological sample obtained from a subject, comprising obtaining a biological sample from a subject suspected of having a neoplasm; conducting the method of claim 40 with the biological sample; and replenishing the purged biological sample to the subject.

42. An in vitro histochemical method of assessing the presence of neoplastic cells in a tissue, comprising obtaining from a patient suspected of having a neoplasm a tissue sample and preparing therefrom a tissue substrate; contacting the tissue substrate with the analogue peptide of claim 1 and allowing the analogue peptide to bind to any neoplastic cells present therein; and detecting the presence of any complexes formed.

43. A hybridoma cell expressing the analogue peptide of claim 1.

44. The hybridoma cell of claim 43, having the ATCC No. HB 11200.

45. The hybridoma cell of claim 43, having the ATCC No. HB 11455.

46. The hybridoma cell of claim 43, having the ATCC No. HB 11454.

47. The hybridoma cell of claim 43, having the ATCC No. HB 11456.

48. The hybridoma cell of claim 43, having the ATCC No. HB _____ (HuBrE3V1).

49. The hybridoma cell of claim 43, having the ATCC No. HB _____ (HuBrE3V3).

50. A composition, comprising the hybridoma cell of claim 43, and a diluent or carrier.

51. A pure, isolated analogue polydeoxyribonucleotide, comprising an analogue oligodeoxyribonucleotide encoding the analogue peptide of

claim 1.

52. The analogue polydeoxyribonucleotide of claim 51, wherein the analogue oligodeoxyribonucleotide comprises a DNA sequence selected from the group consisting of DNA Sequence ID No: 64; DNA Sequence ID No: 65; fragments thereof encoding at least one CDR or one CDR and two flanking regions per chain; redundant DNA sequences thereof; combinations thereof; and mixtures thereof.

53. A hybrid vector, comprising a vector having the analogue polydeoxyribonucleotide of claim 51 operatively linked thereto.

54. The hybrid vector of claim 53, further comprising a polydeoxyribonucleotide comprising an oligodeoxyribonucleotide encoding an effector peptide, the effector peptide-encoding polydeoxyribonucleotide being operatively linked to the vector.

55. A transfected host cell, carrying the hybrid analogue vector of claim 53.

56. A pure, isolated analogue polyribonucleotide, comprising an analogue oligoribonucleotide encoding the analogue peptide of claim 1.

57. A method of producing an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of neoplastic cell, or is released by the cell, comprising a) cloning the analogue polydeoxyribonucleotide of claim 40 into a vector to form a hybrid vector;

b) transfecting a host cell with the hybrid vector and allowing the expression of the analogue peptide; and c) isolating the analogue peptide or mixtures thereof.

58. The method of claim 57, further comprising site-specifically modifying the codon of at least one amino acid in the framework region of a non-human antibody prior to step a) to obtain the analogue polydeoxyribonucleotide.

59. The method of claim 57, wherein steps a) and b) are conducted by cloning analogue polydeoxyribonucleotides encoding analogue peptides selected from the group consisting of at least one CDR or analogue variable region of the first species or fragments thereof comprising at least one CDR and two flanking regions of the heavy or

light chains of the antibody of the first species; and the method further comprises d) allowing the expressed analogue peptides to interact with one another to form double chains.

60. A method of producing a hybrid analogue peptide comprising an analogue peptide which specifically binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or to an antigen released by the cell and an effector peptide, the method comprising a) transfecting a host cell with the hybrid analogue vector of claim 54 and allowing the expression of the anti-carcinoma hybrid analogue peptide; and b) isolating the anti-carcinoma hybrid analogue peptide or mixtures thereof.

61. An anti-idiotypic polymer, comprising polyclonal antibodies raised against anti-tumor antibodies or the analogue peptide of claim 1; monoclonal antibodies thereof capable of specifically binding the anti-neoplasm antibody; fragments thereof selected from the group consisting of CDRs, Fab, Fab', (Fab')₂, variable regions, and fragments thereof; analogues thereof selected from the group consisting of antibodies, Fab, Fab', (Fab')₂, and variable regions wherein about 1 to at least 46 amino acids in the FR are substituted per chain with amino acids selected from the group consisting of amino acids present in equivalent positions in antibodies of species other than the first species, or fragments thereof comprising 1 to 3 CDRs per chain and flanking regions thereof, each of about 1 to at least 10 amino acids, alone or with an N-terminal fragment of about 1 to at least 10 amino acids; combinations thereof, and combinations thereof with an oligopeptide comprising the amino acid sequence APDTRPAPG or fragments thereof comprising the TRP trimer hexamers thereof comprising the trimer, or tandem repeats thereof, wherein each analogue peptide is operatively linked to at least one other analogue peptide; and mixtures thereof.

62. A hybrid anti-idiotypic polymer, comprising the anti-idiotypic polypeptide of claim 61; and an effector agent operatively linked to the anti-analogue idiotypic polypeptide.

63. An anti-carcinoma vaccine, comprising the anti-idiotypic polypeptide of claim 61; and a pharmaceutically-acceptable carrier.

64. The anti-carcinoma vaccine of claim 63, in unit dose form.
65. A anti-tumor vaccination kit, comprising the vaccine of claim 63 and a diluent in separate sterile containers; and instructions for its use.
66. A method of vaccinating against neoplasms comprising administering to a subject an effective amount of the composition of claim 63.
67. The vaccination method of claim 66, wherein the anti-idiotypic polypeptide is administered in an amount of about 0.1 to 5000.00 $\mu\text{g/kg}$ body weight/dose.
68. A method of lowering the serum concentration of a circulating antibody or polypeptide that binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or that is released by the cell, comprising administering to a subject in need of the treatment a pharmaceutically-acceptable composition comprising an amount of the anti-idiotypic polypeptide of claim 61 effective to bind the circulating antibody or polypeptide and accelerate its clearance.
69. The method of claim 68, wherein the anti-idiotypic peptide is administered in an amount of about 0.01 to 5.000.00 mg/kg body weight/dose.
70. A method of inhibiting the growth or reducing the size of a primary or metastasized carcinoma tumor in a subject, comprising administering to the subject an effective amount of a pharmaceutically-acceptable composition comprising an anti-carcinoma hybrid polymer comprising an effector agent selected from the group consist of radioisotopes and therapeutic drugs, and an anti-carcinoma polypeptide which specifically binds to an antigen on the surface or in the cytoplasm of a carcinoma cell or that is released by the cell; allowing the hybrid polymer to reach the tumor and the anti-carcinoma polypeptide to bind thereto; and administering to the subject a pharmaceutically-acceptable composition comprising an amount of the anti-idiotypic polypeptide of claim 61 effective to bind any residual or unbound circulating anti-carcinoma hybrid polypeptide and accelerate the clearance of the hybrid polymer.

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